

**STUDIES ON POLY(LACTIC ACID) AND POLYACRYLATE
BASED ANISOTROPIC MICROPARTICLES MODIFIED
WITH POLYMER BRUSHES**

IFRA



**DEPARTMENT OF MATERIALS SCIENCE AND ENGINEERING
INDIAN INSTITUTE OF TECHNOLOGY DELHI
OCTOBER 2021**

© Indian Institute of Technology Delhi (IITD), New Delhi, 2021.

**STUDIES ON POLY(LACTIC ACID) AND POLYACRYLATE
BASED ANISOTROPIC MICROPARTICLES MODIFIED
WITH POLYMER BRUSHES**

by

IFRA

Department of Materials Science and Engineering

Submitted

in fulfillment of the requirements of the degree of Doctor of Philosophy

to the

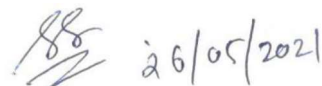


**DEPARTMENT OF MATERIALS SCIENCE AND ENGINEERING
INDIAN INSTITUTE OF TECHNOLOGY DELHI
OCTOBER 2021**

Dedicated to my family

CERTIFICATE

This is to certify that the thesis entitled, “**Studies on poly(lactic acid) and polyacrylate based anisotropic microparticles modified with polymer brushes**” being submitted by **Mrs. Ifra** to Indian Institute of Technology Delhi for the award of degree of **Doctor of Philosophy** is a record of bonafide research work carried out by her. Mrs. Ifra has worked under my guidance and supervision and has fulfilled the requirements for the submission of this thesis, which to my knowledge has reached the requisite standard. The results contained in this thesis are original and have not been submitted, in part or full, to any other University or Institute for the award of any other degree or diploma.



Dr. Sampa Saha

Associate Professor

Department of Materials Science
& Engineering, Indian Institute of
Technology Delhi, Hauz Khas,
New Delhi-110016

ACKNOWLEDGEMENTS

I wish to express my heartiest gratitude to my supervisor, **Prof. (Mrs.) Sampa Saha** for her invaluable guidance and constant encouragement. Her caring attitude and co-operation have been monumental throughout my research. Her constant inspiration and encouragement have been a great motivation for me, behind all the work which I have conducted. Through her wealth of knowledge, direction, and leadership I have been able to expand my knowledge in the area of interface and colloid science.

I am thankful to Prof. A.K. Ghosh, Prof. J. Jacob, Prof. B.K. Satapathy, Prof. Leena Nebhani, Prof. B. Tripathi, Prof. Manjeet Jassal, and all faculties of DMSE, IIT Delhi, for their constant encouragement and help throughout my research work. I express my thanks to Prof. Neetu Singh, Department of Biochemical Engineering and Prof. Hariprasad P., Centre for Rural Development and Technology, IIT Delhi, for providing facilities to carry out experiments in their lab.

It is a pleasure for me to express my gratitude towards all my labmates, an unbroken bond of friendship and brotherhood that prevails in the laboratory is due to the wonderful colleagues. Their warm affection, support during tough times, nice scientific interactions, involvement and conscious as well as unconscious help have rendered my research life a bountiful time. It was really fun to work in their association.

I would like to thank all my seniors Dr. Shivangi Sharma, Dr. Agni Kumar Biswal, Dr. Arun Sharma, Dr. Sabapathy S, Dr. Banpreet Kaur, Devendra Mogha, my batchmates, Smrutirekha Mishra, Dr. Sumbul Hafeez, Sucharita Sethi, Prajesh Nayak, Sahil Malhotra and juniors Srijita Purkayastha, Kalpana Pandey, Shaifali Dhingra, Nidhi Gupta, Meenaskhi Verma, Aiswarya TT, Shikha, Subhra, Aanchal, Tina, Anubhav, Amit from IIT Delhi for their constant encouragement.

My special thanks to Mr. Surender Sharma, Mr. Shivkant, Mr. Ashok Kapoor, Mr. Islam, Mr. Gajraj, Mrs. Shalini Arora, Mr. Subhash, Amit, Pramod, and Sudhir Pandey, for their immediate help whenever needed.

My family members deserve special attention for their support and persistent confidence in me, but I don't have words to express my gratitude for them. I am extraordinarily fortunate to have the blessings of my grandfather Late Mr. Ameerhasan, my grandmother Mrs. Aneesa, my father Mohd. Dilshad and my mother Mrs. Shaheen who always bless me with the shower of their love, providing inner strength, patience, emotional support and making sacrifices for my successful career. I express my heartiest thanks to my husband, Mr. Aamir for his constant support, encouragement, and patience which enable me to pursue my career. I am also thankful to my brother Tabish, Uzair, my sister Rimsha. A special thanks to my parent-in-laws Mr. Riazuddin and Mrs. Nafisha for their love and support.

Finally, I would like to thank everyone who helped me in the successful compilation of thesis, as well as expressing my apology that I could not mention personally one by one.

I would like to acknowledge the financial assistance received from CSIR which has helped me to pursue my Ph.D. without any stress.

Last but not the least I am thankful to the Almighty God in helping me to accomplish this task.

IFRA

ABSTRACT

Recently, biocompatible anisotropic particles have emerged as a powerful platform in the biomedical field, especially in the areas such as therapeutics, theranostics, biosensor, targeted drug delivery, triggered drug delivery, cellular uptake, targeted cellular internalization, etc., because of their unique characteristics and superior performances over their isotropic analogues. Macromolecular anisotropic particles can primarily be classified into three different categories, such as shape anisotropy, compositional anisotropy, and surface anisotropy, i.e., heterogeneous surface chemistry leading to anisotropic surface patches. We were particularly interested to fabricate biocompatible/biodegradable anisotropic particles based on biodegradable poly(lactic acid) and biocompatible polyacrylates.

In order to achieve the target, we have synthesized a random copolymer of methyl methacrylate (MMA) and 2-hydroxy ethyl methacrylate (HEMA) via Atom Transfer radical polymerization (ATRP) whose hydroxyl groups were later modified as bromo functionalized moieties named as poly(MMA-*co*-BEMA) (poly(methyl methacrylate-*co*-2-(2-bromopropionyloxy) ethyl methacrylate). In chapter 2, this bromo functionalized random copolymer, poly(MMA-*co*-BEMA) was blended with poly(lactic acid) by dissolving both in a mixture of chloroform and dimethyl formamide and the resultant polymer solutions were used to fabricate cup shaped microparticles of ~6 μm size via electrohydrodynamic jetting technique. The polymer solutions were jetted at different solution parameters (concentration, polymer composition, solvent system) and processing parameters (flow rate). And several properties of the jetted particles such as electrical conductivity, viscosity, surface tension etc. were studied to have a good understanding of the mechanism involved in cup shape formation. The mechanistic understandings also provide insight into the transition from cup, sphere to disc shape of the jetted particles.

In chapter 3, two hydrophilic brushes i.e., poly(HEMA) (poly(2-hydroxy ethyl methacrylate)) and poly(DMAEMA) (poly(2-(Dimethylamino) ethyl methacrylate)) were separately and individually grown from the surface of cup shaped microparticles. The shape-shifting of cup-shaped particles occurred when surface initiated ATRP of HEMA (2-hydroxy ethyl methacrylate) was carried out for 1 hour because of the immobilization of self-crosslinkable poly(HEMA) brushes onto the surface of the cup shaped particles. Post polymerization, these particles underwent controlled bending leading to the formation of spherical ball with one small opening (hole). However, no change of shape was observed while growing non-crosslinkable hydrophilic poly(DMAEMA) (poly(2-(Dimethylamino) ethyl

methacrylate)) brush from the surface of cup shaped particles. To understand the underlying phenomena of shape shifting, simulation studies were also performed. Dissipative particle dynamics (DPD) simulation of the ATRP process at the cup surface further confirmed that the use of HEMA monomer indeed led to the desired structure of the cup particle due to the crosslinking connectivity across the interface. We have also calculated the radial distribution function (RDF) and radius of gyration to study the structure of modified particles.

In chapter 4, responsive properties of poly(DMAEMA) (poly(2-dimethylamino ethyl methacrylate)) brush modified cup shaped particles were studied and a small molecule (Rhodamine B) was adsorbed at different pH. An enhanced dye adsorption at specific pH was observed. Dye released study showed the burst release of dye at acidic pH due to the swelling of poly(DMAEMA) brushes. In another study, a model enzyme (macromolecule) such as α -glucosidase was immobilized onto poly(DMAEMA) (poly(2-dimethyl amino ethyl methacrylate)) brush modified anisotropic (cup and disc shaped) biocompatible polymeric particles. We have also demonstrated the role of swollen polymer brush grafted on the surface of cup shaped particles via surface initiated atom transfer radical polymerization, in immobilizing unprecedentedly high loading of enzyme (440 mg/g (cup) – 589 mg/g (disc) of particles, 15-20 times higher than literature reported system) as compared to non-brush modified particles. Circular dichroism spectroscopy was used to predict the structural changes of the enzyme upon immobilization onto the carrier particles. An enormously high amount of enzyme with preserved activity ($\sim 85 \pm 13\%$ for cups and $\sim 78 \pm 15\%$ for discs) was found to adhere onto brush modified particles at pH 7 via electrostatic adsorption and the enzyme loading capacity was observed to be progressively higher with increase of enzyme concentration and grafting density as well, till the saturation limit. In addition, accelerated particle separation was also achieved via magnetic force induced aggregation within 20 min (without centrifuge) by incorporating magnetic nanoparticles into disc shaped particles while electrojetting.

In last chapter, we have focused to make compositional anisotropic microparticles, i.e., Janus microparticles consisting of two different phases where poly(MMA-*co*-BEMA) was incorporated into one compartment only and hydrophilic brushes, poly(DMAEMA) was grown from the this particular compartment only, These selectively brush modified Janus particles which contained one part hydrophilic (poly(DMAEMA)) and another part hydrophobic (poly(lactic acid)). These types of amphiphilic particles were shown to be well suited for the Pickering emulsion stabilization to stabilize octanol(oil)/water emulsion. Metal nanoparticles such as gold nanoparticles and iron (0) nanoparticles were incorporated into two different

compartments of these brush modified Janus particles where gold and iron nanoparticles were simultaneously demonstrated to catalyze the reduction of *p*-nitrophenol to *p*-aminophenol (100% conversion within 1 min) and the dechlorination of trichloroethylene in same octanol (oil)/water emulsion, respectively.

सार

हाल ही में, बायोकम्पैटिबल अनिसोट्रोपिक कण बायोमेडिकल क्षेत्र में एक शक्तिशाली मंच के रूप में उभरे हैं, विशेष रूप से चिकित्सा विज्ञान, चिकित्सा विज्ञान, बायोसेंसर, लक्षित दवा वितरण, ट्रिगर दवा वितरण, सेलुलर तेज, लक्षित सेलुलर आंतरिककरण, आदि जैसे क्षेत्रों में, उनके अद्वितीय कारण उनके आइसोट्रोपिक एनालॉग्स पर विशेषताओं और बेहतर प्रदर्शन। मैक्रोमोलेक्यूलर अनिसोट्रोपिक कणों को मुख्य रूप से तीन अलग-अलग श्रेणियों में वर्गीकृत किया जा सकता है, जैसे कि आकार अनिसोट्रोपी, कंपोजिटल अनिसोट्रोपी, और सतह अनिसोट्रोपी, यानी विषम सतह रसायन जो अनिसोट्रोपिक सतह पैच के लिए अग्रणी है। हम विशेष रूप से बायोकम्पैटिबल/बायोडिग्रेडेबल अनिसोट्रोपिक कणों को बायोडिग्रेडेबल पॉली (लैक्टिक एसिड) और बायोकंपैटिबल पॉलीएक्रिलेट्स पर आधारित बनाने में रुचि रखते थे।

लक्ष्य को प्राप्त करने के लिए, हमने एटम ट्रांसफर रेडिकल पोलीमराइजेशन (एटीआरपी) के माध्यम से मिथाइल मेथैक्रिलेट (एमएमए) और 2-हाइड्रॉक्सी एथिल मेथैक्रिलेट (एचईएमए) के एक यादृच्छिक कॉपोलीमर को संश्लेषित किया है, जिसके हाइड्रॉक्सिल समूहों को बाद में पॉली (पॉली) नामक ब्रोमो फंक्शनल मोएट के रूप में संशोधित किया गया था। (पॉली(मिथाइल मेथैक्रिलेट-को-2 (2 ब्रोमोप्रोपियोनीलोक्सी) एथिल मेथैक्रिलेट))। अध्याय 2 में, इस ब्रोमो ने रैंडम कॉपोलीमर, पॉली (MMA-co-BEMA) को पॉली (लैक्टिक एसिड) के साथ मिश्रित किया था। क्लोरोफॉर्म और डाइमिथाइल फॉर्माइड के मिश्रण में घुलने और परिणामी बहुलक समाधानों का उपयोग इलेक्ट्रोहाइड्रोडायनामिक जेटिंग तकनीक के माध्यम से ~ 6 माइक्रोन आकार के कप के आकार के माइक्रोपार्टिकल्स बनाने के लिए किया गया था। बहुलक समाधान विभिन्न समाधान मापदंडों (एकाग्रता, बहुलक संरचना, विलायक प्रणाली) पर जेट किए गए थे। और प्रसंस्करण पैरामीटर (प्रवाह दर) और जेटेड कणों के कई गुणों जैसे विद्युत चालकता, चिपचिपाहट, सतह तनाव इत्यादि का अध्ययन किया गया ताकि उन्हें अच्छी समझ हो f कप के आकार के निर्माण में शामिल तंत्र। यंत्रवत समझ कप, गोले से डिस्क आकार में जेट कणों के संक्रमण में भी अंतर्दृष्टि प्रदान करती है।

अध्याय 3 में, दो हाइड्रोफिलिक ब्रश यानी, पॉली (एचईएमए) (पॉली (2-हाइड्रॉक्सी एथिल मेथैक्रिलेट)) और पॉली (डीएमईएमए) (पॉली (2- (डाइमिथाइलैमिनो) एथिल मेथैक्रिलेट)) अलग-अलग और व्यक्तिगत रूप से कप की सतह से उगाए गए थे। आकार के सूक्ष्म कण। कप के आकार के कणों का आकार-स्थानांतरण तब हुआ जब सतह पर शुरू हुआ HEMA (2-हाइड्रॉक्सी एथिल मेथैक्रिलेट) का

ATRP कप के आकार के कणों की सतह पर सेल्फ-क्रॉसलिंगेबल पॉली (HEMA) ब्रश के स्थिरीकरण के कारण 1 घंटे के लिए किया गया था। . पोलीमराइजेशन के बाद, इन कणों को नियंत्रित झुकाव से गुजरना पड़ा, जिससे एक छोटे से उद्घाटन (छेद) के साथ गोलाकार गेंद का निर्माण हुआ। हालांकि, कप के आकार के कणों की सतह से गैर-क्रॉसलिंगेबल हाइड्रोफिलिक पॉली (डीएमईएमए) (पॉली (2-डाइमिथाइलैमिनो) एथिल मेथैक्रिलेट)) ब्रश उगाते समय आकार में कोई बदलाव नहीं देखा गया। आकार बदलने की अंतर्निहित घटनाओं को समझने के लिए, सिमुलेशन अध्ययन भी किए गए थे। कप की सतह पर एटीआरपी प्रक्रिया के विघटनकारी कण गतिकी (डीपीडी) सिमुलेशन ने आगे पुष्टि की कि एचईएमए मोनोमर के उपयोग ने वास्तव में इंटरफ़ेस में क्रॉसलिंग कनेक्टिविटी के कारण कप कण की वांछित संरचना का नेतृत्व किया। हमने संशोधित कणों की संरचना का अध्ययन करने के लिए रेडियल डिस्ट्रीब्यूशन फंक्शन (आरडीएफ) और रेडियस रेडियस की भी गणना की है।

अध्याय 4 में, पॉली (डीएमईएमए) (पॉली (2 डाइमिथाइलैमिनो एथिल मेथैक्रिलेट)) ब्रश संशोधित कप के आकार के कणों के प्रतिक्रियाशील गुणों का अध्ययन किया गया था और एक छोटे अणु (रोडामाइन बी) को विभिन्न पीएच में सोख लिया गया था। विशिष्ट पीएच पर एक बढ़ाया डाई सोखना देखा गया। डाई जारी किए गए अध्ययन ने पॉली (डीएमईएमए) ब्रश की सूजन के कारण अम्लीय पीएच पर डाई के फटने को दिखाया। एक अन्य अध्ययन में, एक मॉडल एंजाइम (मैक्रोमोलेक्यूल) जैसे कि α -ग्लूकोसिडेज़ को पॉली (डीएमईएमए) (पॉली (2-डाइमिथाइल एमिनो एथिल मेथैक्रिलेट)) ब्रश संशोधित अनिसोट्रोपिक (कप और डिस्क आकार) बायोकंपैटिबल पॉलीमेरिक कणों पर स्थिर किया गया था। हमने सतह पर शुरू किए गए परमाणु हस्तांतरण कट्टरपंथी पोलीमराइजेशन के माध्यम से कप के आकार के कणों की सतह पर ग्राफ्ट किए गए सूजे हुए बहुलक ब्रश की भूमिका का भी प्रदर्शन किया है, जो एंजाइम (440 मिलीग्राम / जी (कप) - 589 मिलीग्राम / जी (डिस्क) के अभूतपूर्व उच्च लोडिंग को स्थिर करता है कण, गैर-ब्रश संशोधित कणों की तुलना में साहित्य रिपोर्ट प्रणाली से 15-20 गुना अधिक)। वाहक कणों पर स्थिरीकरण पर एंजाइम के संरचनात्मक परिवर्तनों की भविष्यवाणी करने के लिए सर्कुलर डाइक्रोइज्म स्पेक्ट्रोस्कोपी का उपयोग किया गया था। इलेक्ट्रोस्टैटिक सोखना के माध्यम से पीएच 7 पर ब्रश संशोधित कणों का पालन करने के लिए संरक्षित गतिविधि ($\sim 85 \pm 13\%$ कप के लिए और $\sim 78 \pm 15\%$ डिस्क के लिए) के साथ एंजाइम की एक बड़ी मात्रा में पाया गया था और एंजाइम लोडिंग क्षमता को उत्तरोत्तर देखा गया था। संतृप्ति सीमा तक एंजाइम सांद्रता और ग्राफ्टिंग घनत्व में वृद्धि के साथ उच्च। इसके अलावा, इलेक्ट्रोजेटिंग करते समय डिस्क के आकार के कणों में चुंबकीय नैनोकणों को शामिल करके 20 मिनट (अपकेंद्रित के बिना) के भीतर चुंबकीय बल प्रेरित एकत्रीकरण के माध्यम से त्वरित कण पृथक्करण भी प्राप्त किया गया था।

पिछले अध्याय में, हमने कंपोजिटल अनिसोट्रोपिक माइक्रोपार्टिकल्स बनाने पर ध्यान केंद्रित किया है, यानी, दो अलग-अलग चरणों से युक्त जानूस माइक्रोपार्टिकल्स जहां पॉली (एमएमए-को-बीईएमए) को केवल एक डिब्बे में शामिल किया गया था और हाइड्रोफिलिक ब्रश, पॉली (डीएमईएमए) को इसी से उगाया गया था। केवल विशेष डिब्बे, ये चुनिंदा ब्रश संशोधित जानूस कण जिसमें एक भाग हाइड्रोफिलिक (पॉली (डीएमईएमए)) और दूसरा भाग हाइड्रोफोबिक (पॉली (लैक्टिक एसिड)) होता है। इस प्रकार के एम्फीफिलिक कणों को ऑक्टेनॉल (तेल)/पानी इमल्शन को स्थिर करने के लिए पिकरिंग इमल्शन स्थिरीकरण के लिए अच्छी तरह से अनुकूल दिखाया गया था। सोने के नैनोकणों और लोहे (0) नैनोकणों जैसे धातु के नैनोकणों को इन ब्रश संशोधित जानूस कणों के दो अलग-अलग डिब्बों में शामिल किया गया था, जहां सोने और लोहे के नैनोकणों को एक साथ पी-नाइट्रोफेनॉल की कमी को पी-एमिनोफेनोल में उत्प्रेरित करने के लिए प्रदर्शित किया गया था (1 के भीतर 100% रूपांतरण मिनट) और एक ही ऑक्टेनॉल (तेल)/पानी के पायस में क्रमशः ट्राइक्लोरोइथिलीन का डीक्लोरीनीकरण।

TABLE OF CONTENTS

	Page No.
CERTIFICATE	i
ACKNOWLEDGEMENTS	ii
ABSTRACT	iv
TABLE OF CONTENTS	x
LIST OF FIGURES	xv
LIST OF TABLES	xxvii
LIST OF ABBREVIATIONS	xxviii
 CHAPTER 1: INTRODUCTION AND LITERATURE SURVEY	 1-70
1.1 Motivation and Background	2
1.2 Shape anisotropy	5
1.2.1 Droplet Microfluidics	6
1.2.2 Electrospraying	10
1.2.3 Self-assembly of block copolymers	12
1.2.4 Film Stretching Method	15
1.2.5 Particle Replication in Non-wetting Templates (PRINT)	17
1.3 Compositional anisotropy	19
1.3.1 Electrohydrodynamic co-jetting technique	20
1.3.2 Emulsion technique	26
1.3.3 Droplet microfluidics	29
1.4 Surface anisotropy	34
1.4.1 Anisotropic Patchy Particles	34
1.4.2 Polymer brush modified anisotropic particles	38
1.5 Applications	43
1.5.1 Drug Delivery	43
1.5.2 Biodistribution	47
1.5.3 Cellular uptake	51
1.5.4 Catalysis	54
1.6 Research Gap	56
1.7 Objective	56

1.8	Plan of Work	56
1.9	Thesis format	57
	References	59
CHAPTER 2: FABRICATION OF ANISOTROPIC CUP SHAPED MICROPARTICLES COMPOSED OF POLY(LACTIC ACID) AND POLY(ACRYLATES)		71-94
2.1	Introduction	72
2.2	Results and Discussions	73
	2.2.1 Synthesis and characterization of macroinitiator	73
	2.2.2 Fabrication of cup shaped particles	76
2.3	Conclusions	87
2.4	Experimental Section	87
	2.4.1 Materials	87
	2.4.2 Characterization Methods	88
	2.4.3 Synthesis of Poly(methyl methacrylate-co-2-hydroxy ethyl methacrylate)(poly(MMA-co-HEMA))	88
	2.4.4 Synthesis of macroinitiator [poly[methyl methacrylate-co-2-(2-bromopropionyloxy) ethyl methacrylate] (poly(MMA-co-BEMA))](macroinitiator)	89
	2.4.5 Fabrication of particles using electrojetting	89
	References	91
CHAPTER 3: SHAPE SHIFTING OF ANISOTROPIC CUP SHAPED PARTICLES ON GROWING POLY(2-HYDROXY ETHYL METHACRYLATE) BRUSHES BY “GRAFTING FROM” APPROACH		95-121
3.1	Introduction	96
3.2	Results and Discussions	98
	3.2.1 Polymer brush growth from surface of cup shaped particles	98
	3.2.2 Simulation studies	106
3.3	Conclusions	112
3.4	Experimental Section	112

3.4.1 Materials	112
3.4.2 Characterization Methods	113
3.4.3 Surface initiated polymerization of 2-hydroxy ethyl methacrylate (HEMA) and 2-(Dimethylamino)ethyl methacrylate (DMAEMA) from surface of cup shaped particles	113
3.4.4 Dye (Rhodamine B) adsorption onto the particles for CLSM study	114
3.4.5 Simulation Studies	114
References	116
 CHAPTER 4A: SURFACE MODIFICATION OF CUP SHAPED PARTICLES POLY(2-DIMETHYLAMINO ETHYL METHACRYLATE) BRUSH AND THEIR pH RESPONSIVE BEHAVIOUR	 123-144
4A.1 Introduction	123
4A.2 Results and discussion	124
4A.3 Conclusions	137
4A.4 Experimental Section	137
4A.4.1 Materials	137
4A.4.2 Characterizations Techniques	137
4A.4.3 Surface initiated polymerization of DMAEMA from surface of cup shaped particles	138
4A.4.4 Adsorption of water soluble dye (Rhodamine B)	139
4A.4.5 Adsorption kinetics and adsorption isotherm	139
4A.4.6 Dye release study	139
References	141
 CHAPTER 4B: ADSORPTION OF α- GLUCOSIDASE ON POLY(2-DIMETHYLAMINO ETHYL METHACRYLATE) BRUSH MODIFIED CUP SHAPED PARTICLES AND THE IMPACT OF BRUSH ON ENZYME LOADING CAPACITY	 145-180
4B.1 Introduction	145
4B.2 Results and discussion	147

4B.2.1 Enzyme immobilization on brush modified anisotropic particles	147
4B.2.2 Impact of brush grafting density on enzyme immobilization	155
4B.2.3 Impact of shape of anisotropic particles on enzyme immobilization	158
4B.2.4 Characterization of enzyme immobilized particles	161
4B.2.5 Facile separation of enzyme immobilized particles induced by magnetic force	164
4B.2.6 Storage stability of immobilized enzyme	169
4B.3 Conclusions	169
4B.4 Experimental Section	170
4B.4.1 Materials	170
4B.4.2 Characterization Methods	170
4B.4.3 Fabrication of cup shaped particles (P ₁ & P ₃) and surface modification of P ₁ with Poly(DMAEMA) brushes (P ₂)	171
4B.4.4 Fabrication and surface modification of Disc shaped particles	172
4B.4.5 Fabrication of poly(DMAEMA) brush modified discoid particles entrapped with magnetic nanoparticles	172
4B.4.6 Immobilization of α -glucosidase enzyme onto the particles	173
References	175
 CHAPTER 5: DEVELOPMENT OF pH RESPONSIVE ANISOTROPIC (JANUS) PARTICLES DECORATED WITH GOLD NANOPARTICLES AS PICKERING EMULSION STABILIZER AND THEIR APPLICATION IN CATALYSIS	 181-212
5.1 Introduction	182
5.2 Results and discussion	185
5.2.1 Fabrication of bicompartmental/Janus particles and spatio-selective surface modification	185

5.2.2	Stabilization of Pickering emulsion based on Octanol/Water	193
5.2.3	Synthesis of Gold nanoparticles onto poly(DMAEMA) brush modified bicompartmental particles	196
5.2.4	Simultaneous catalytic reduction of p-nitrophenol and dechlorination of trichloroethylene	199
5.3	Conclusions	203
5.4	Experimental section	204
5.4.1	Materials	204
5.4.2	Characterization techniques	204
5.4.3	Fabrication of bicompartmental/Janus particles	205
5.4.4	Selective surface modification via growing poly(2-dimethyl amino ethyl methacrylate)(poly(DMAEMA)) brushes	206
5.4.5	Octanol in water based pickering emulsion stabilized by P ₁₀ , P ₂₅ and P ₅₀	206
5.4.6	In Situ synthesis of gold nanoparticles onto poly(DMAEMA) brushes	206
5.4.7	Catalytic reduction of p-nitrophenol	207
5.4.8	Loading of Iron (0) nanoparticles and dechlorination of trichloroethylene (TCE)	207
5.4.9	Simultaneous reduction of PNP and TCE	208
	References	209
CHAPTER 6: SUMMARY, CONCLUSIONS AND FUTURE SCOPE		213-216
6.1	Summary	214
6.2	Future scope	216
	List of Publications	217

LIST OF FIGURES

	Page No.
Figure 1.1 A schematic diagram indicating the emulsion formation by PDMS microfluidic device and its solidification via UV induced polymerization.	7
Figure 1.2 (a)-(c) Schematic representation of deformed droplets (disk, ellipsoid, rod). (d) polyTPGDA disk shaped particles. (e) polyTPGDA elliptical shaped particles. (f) polyTPGDA rod shaped particles. (g) PEG-DA hexagonal shaped particles. (h) PEG-DA circular shaped. (i) PEG-DA triangular shaped. (j) PLA based crescent shaped particles.	9
Figure 1.3 Scanning Electron Microscopy (SEM) images of (a) PLA/poly(MMA- <i>co</i> -BEMA) cup shaped particles. (b) PLA/poly(MMA- <i>co</i> -BEMA) disc shaped particles. (c) PLA/poly(MMA- <i>co</i> -BEMA) spherical particles with one hole. (d) PMMA cups. (e) chitosan based red blood shaped particles.	12
Figure 1.4 (a) Ellipsoidal particles having an axially stacked lamellar structure and nanosheets having hexagonal structure of PFS cylinders composed of PFS- <i>b</i> -P2VP). (b)-(e) Cuboids made from PMAA ₁₁₂ - <i>b</i> -PMAAZ ₆₆ (subscript indicates DP (degree of polymerization) of particular block), short belts made from PMAA ₁₁₂ - <i>b</i> -PMAAZ ₈₉ , lamellae made from PMAA ₁₁₂ - <i>b</i> -PMAAZ ₁₁₅ and ellipsoidal vesicles made from PMAA ₁₁₂ - <i>b</i> -PMAAZ ₁₄₂ , respectively.	14
Figure 1.5 Scanning Electron Microscopy (SEM) of (a)-(b) PLGE nanoellipsoids fabricated by cold stretching (100% and 200%, respectively), (c)-(f) PLGE nanorods fabricated by hot stretching (60%, 120%, 120% and 150%, respectively) (g)-(i) PLGE nanodisks fabricated by hot compressing (200%, 200% and 100%, respectively). (j) PLGA lipid coated particles. (k) PLGA microrods fabricated from different ratio of PEMA (poly (ethylene-alt-maleic acid))/PVA.	16
Figure 1.6 Schematic diagrams for PRINT Technique. Scanning Electron Microscopy (SEM) images of (a) Conical triacrylate particles. (b) PLA trapezoidal particles. (c), (d), (e), (f), (g) PLGA particles (80nm × 320nm cylinders, 200nm × 200nm cylinders, 200nm × 600nm cylinders, 2µm cubes with ridges, and 3µm particles with centre fenestrations, respectively). (h) Acid labile drug containing PLGA particles.	19
Figure 1.7 Common forms of compositionally anisotropic particles or janus particles.	20
Figure 1.8 (a) General setup for electrohydrodynamic co-jetting technique. (b) & (c) DIC image of PLA/PLGA biphasic particles encapsulated with levodopa and	22

carbidopa. (c), (d) and (e) are SERS tagged gold nanoparticles encapsulated biphasic particles composed of PLGA.

Figure 1.9 (a) Basic setup to fabricate triphasic particles (b) Scanning Electron Microscopy (SEM) image of PEO based triphasic particles. 23

Figure 1.10 (a)-(d) Shape shifting of bicompartamental PLGA/PLGA cylindrical microparticles (30 μm in length) ((c) is intermediate stage) by ultrasound heat treatment (e)-(f) Shape shifting of bicompartamental PLGA/PLGA cylindrical microparticles (70 μm in length) ((c) is intermediate stage) by ultrasound heat treatment (g)-(h) Shape shifting of tricompartmental cylindrical particles by ultrasound heat treatment (i)-(l) Shape shifting of bicompartamental PLGA/PMMA cylindrical microparticles by heat treatment: only PLGA part gets reconfigure. (m)-(n) Reversible shifting of hydrogel/PLGA (core/shell) microcylinders by swelling/deswelling mechanisms. (o)-(p) Reversible shape bending of PEO/PVCi microcylinders in presence/absence of dioxane. 25

Figure 1.11 (a) Helically compartmentalized microfibers having diameter around 30 μm (b) Helically compartmentalized microcylinders after cryosectioning of microfibers (c) Shape reconfiguration of microcylinders (with different degree of helicity) to spheres. 26

Figure 1.12 Scanning Electron Microscopy Images of (a) Polymer/Precirol particles. (b) PLGA/PCL particle. (c) Glibenclamide-loaded microparticles composed of PLGA/PCL. (d) Cross sectional view of bi-compartmental with a PLGA/PCL ratio (20/10). (e) PLGA/PLLA biphasic particles. (f) Cross sectional view of bi-compartmental PLGA/PLLA biphasic particles. 28

Figure 1.13 (a) Fluorescent microscopic image of polyacrylamide particles containing PNIPAM nanoparticles on one side. (b) & (c) Scanning Electron Microscopic Images of pectin/pectin homojanus and alginate/pectin hetero janus particles. (d) PLGA/PCL bicompartamental particles. 30

Figure 1.14 (a) Acetylene –modified containing bicompartamental microparticles which were modified with PEG-amine-FITC. (b) Tetracompartmental microcylinders containing acetylene functionalized PLGA in one of the compartment and selectively modified with streptavidin 35

Figure 1.15 In (A)–(C), microparticles displaying polymers 3 to 5 in one hemisphere only are selectively surface functionalized through Staudinger ligation (A), photo-immobilization (B), and alkyne/azide click chemistry (C). 37

Figure 1.16 Selective surface functionalization of three-patch microparticles (B1) Schematic (B2) CLSM images to of surface modified three patch particles 38

(particles containing poly 1, 4 and 5) (B3) 3 dimensional reconstruction of the microparticles collected in the z-direction.

Figure 1.17 Confocal Laser Scanning Microscopic images of (a) microcylinders having 120 μ m length before polymerization (b) microcylinders having 120 μ m length before polymerization. 40

Figure 1.18 Scanning electron microscopic image of (a) cup shaped particle before polymerization (b) cup shaped particle after growing PDMAEMA brushes, Confocal Laser Scanning Microscopic images of (c) & (d) Rhodamine B dye adsorption on cup shaped particles before polymerization at pH-7 & pH-4, respectively, (e) & (f) Rhodamine B dye adsorption on cup shaped particles after polymerization at pH-7 & pH-4, respectively. 41

Figure 1.19 Scanning electron microscopic image of cup shaped particle polymerized at different intervals to grow PHEMA brushes (a) 0h (b) 15min (c) 1h (d) 3h and (e) 19h. 42

Figure 1.20 (a) Schematic for the selective degradation of PEO containing poly(AAM-*co*-AA) compartment at pH 7.4 (b) % release of FITC-PEO at different pH. 45

Figure 1.21 Drug release profile for LD and CD encapsulated biphasic PLGA/PLLA particles (a) when LD is in PLA and CD is in PLGA (b) when LD is in PLGA and CD is in PLA. 46

Figure 1.22 (a) Doc concentration Vs Time curve at different time intervals and for various organs (b) % release of Doc with respect to time. 49

Figure 1.23 (a) Fabrication scheme of RBC coated spherical, prolate ellipsoid and oblate ellipsoid particles (b) Half-life of RBC coated and uncoated spherical, prolate ellipsoid and oblate ellipsoid particles (c) Biodistribution of RBC coated and uncoated spherical, prolate ellipsoid and oblate ellipsoid particles. 50

Figure 1.24 (a) Cellular-uptake kinetics of 200nm discs, 325nm discs 400nm rods and 800nm rods in various cell lines. (a) HeLa cells, (b) HEK 293 cells, (c) BMDCs, and (d) HUVEC cells. 52

Figure 1.25 (a) Internalization profile of cubic and cylindrical particles prepared via PRINT technique with HeLa cells over an incubation period of 4h. Reproduced from ref 31. Copyright 2008, with permission from PNAS. (b) PVPON/TA spherical and hemispherical particles internalized per cells. 53

Figure 1.26 Catalytic reduction of *p*-nitrophenol by Ag embedded poly(St-*co*-VA)/(PTA) Janus particles. 55

Figure 1.27 (a) UV-Vis spectra for reduction of <i>p</i> -nitrophenol catalysed by Au embedded SiO ₂ @PDVB/PS Janus particles (b) UV-Vis spectra for reduction of <i>p</i> -nitrophenol catalysed by Au embedded SiO ₂ @PDVB/PS Janus particles (c) UV-Vis spectra for reduction of methylene blue dye by Ag embedded PAA/PS Janus particles	55
Figure 2.1 ¹ HNMR spectrum of random copolymer of MMA and HEMA [poly(MMA- <i>co</i> -HEMA)]	75
Figure 2.2 ¹ HNMR spectrum of random copolymer of MMA and BEMA (poly(MMA- <i>co</i> -BEMA))	76
Figure 2.3 SEM images of electrojetted samples at a concentration of 1 wt% having (a) sample 1 (0% PLA) (b) sample 2 (25% PLA) (c) sample 3 (50% PLA) (d) sample 4 (75% PLA) (e) sample 5 (90% PLA) and (f) sample 6(100% PLA).	77
Figure 2.4 DSC thermograms of poly (MMA- <i>co</i> -BEMA) and sample 4.	81
Figure 2.5 SEM images of electrojetted samples from 1 wt% polymer (sample 4) solution in different solvents (a) Dichloromethane (b) Chloroform (c) Chloroform : DMF (98:2) (d) Chloroform : DMF (96:4) (e) Chloroform : DMF (95:5) (f) Chloroform : DMF (94:6)	82
Figure 2.6 SEM images of electrojetted samples (sample 4) at different concentration with same solvent system (chloroform: DMF – 95:5) (a) 0.5wt% (b) 1wt% (c) 2wt% (d) 3wt%.	85
Figure 2.7 SEM images of electrojetted solutions of sample 4 at a flow rate of (a) 0.5 mL/h (b) 1 mL/h (c) 1.5 mL/h.	85
Figure 2.8 SEM image of cup shaped particles electrojetted from 1 wt% solution of poly (MMA- <i>co</i> -DMAEMA) and PLA blend at a ratio of (1:3) prepared in a solvent system of chloroform:DMF (95:5) (flow rate: 1 mL/h and applied voltage: 9 KV).	87
Figure 3.1 Representative Scanning Electron Microscopic images of dry (a) cup shaped particles (b) Poly(HEMA) brush modified cup shaped particles for 1h (c) Poly(DMAEMA) brush modified cup shaped particles for 1h.	99
Figure 3.2 Representative Scanning Electron Microscopic images of dry poly(HEMA) brushes modified cup shaped particles for (a) 0h (b) 15 min (c) 1h (d) 3h (e) 19h and (f) variation of hole size (opening) in cup shaped particles over polymerization time (a portion of the graph shown in the inset).	102

Figure 3.3 Representative Scanning Electron Microscopic images of dry poly(HEMA) brush modified cup shaped particles for (a) 0 min (b) 15 min (c) 60 min.	103
Figure 3.4 Raman spectra of (A) cup shaped particles (black line) (B) Poly(DMAEMA) brushes modified cup shaped particles (red line). (C) Poly(HEMA) brushes modified cup shaped particles (blue line).	103
Figure 3.5 FTIR (Fourier Transform Infrared) spectra of (a) Cup shaped particles (blue line) (b) Poly(HEMA) brushes modified cup shaped particles (red line) (c) Poly(DMAEMA) brushes modified cup shaped particles (black line).	104
Figure 3.6 CLSM (Confocal Laser Scanning Microscopic) and DIC (Differential Interference Contrast) Images of Rhodamine B adsorbed (a) and (d) cup shaped particles, (b) and (e) Poly(HEMA) brush modified cup shaped particles, (c) and (f) Poly(DMAEMA) brush modified cup shaped particles, respectively.	106
Figure 3.7 (a) The monomer conversion (in black) and $\ln([M]_0/[M]_t)$ (in red) as a function of time during the polymerization for DMAEMA (as in Figure 1a) where $[M]_t$ is the current concentration of unreacted monomer. The dimensionless time (reaction time), t , is the number of simulation steps multiplied by the time step, $\Delta t = 0.02$. (b) The conversion (black curves) and $\ln([M]_0/[M]_t)$ (red curves) as a function of time during the polymerization for HEMA monomers and crosslinker for the evolution shown in Figure 1b.	107
Figure 3.8 Time evolution of the alteration observed in the cup particle due to the growth of polymer brushes via atom transfer radical polymerization (ATRP) in the presence of (a) DMAEMA monomers and (b) HEMA monomers for $[I]_0/[X]_0/[M]_0 = 1/5/20$, and $[I]_0/[X]_0/[M]_0 = 1/5/75$ at the time displayed in the figure. The orange and purple beads represent the cup particle (PLA) and the embedded initiators (poly(MMA-co-BEMA), respectively. The initiators are distributed equally on both the inner and outer surfaces of cup particle. The green bead represents an active monomer radical, and the blue bead shows the reactive monomer. Partially active and entirely reactive crosslinked HEMA beads are denoted by yellow and red, respectively.	108
Figure 3.9 Comparison of the radial distribution function ($g(r)$) and radius of gyration (R_g) of the modified cup particle. The black curve represents $g(r)$ and R_g for poly(DMAEMA) brush; the red and green curves represent $g(r)$ and R_g for poly(HEMA) brush for $[I]_0/[X]_0/[M]_0 = 1/5/20$, and $[I]_0/[X]_0/[M]_0 = 1/5/75$ respectively.	109
Figure 3.10 Time evolution of the alteration observed in the cup particle when initiators are distributed in a ratio 1:2 on both the inner and outer surfaces of cup	111

particle; $[I]_0/[X]_0/[M]_0 = 1/5/20$. Comparison of the radial distribution function ($g(r)$) and radius of gyration (R_g) of the modified cup particle.

Figure 4A.1. (a) SEM image of dry cup shaped particles (average particle diameter - $5.2\mu\text{m} \pm 0.60$) (b) AFM image (phase image) of dry cup shaped particles (c) SEM image of PDMAEMA brushes modified cup shaped particles for 1h (average particle diameter - $9.0\mu\text{m} \pm 2.1$). (d) AFM image (Height image) of PDMAEMA brushes modified cup shaped particles for 1h. Scale bar is $5\mu\text{m}$. **Note:** For elongated cup shaped particles, major axis has been measured as diameter. 125

Figure 4A.2 Brightfield images of (a) cup shaped particles (average particle diameter - $5.9\mu\text{m} \pm 0.6$) (b), (c) and (d) particles after growing PDMAEMA brushes from the surface for 15 min (average particle diameter - $6.8\mu\text{m} \pm 1.6$), 30 min (average particle diameter - $7.6\mu\text{m} \pm 1.5$) and 1h (average particle diameter - $9.2\mu\text{m} \pm 1.8$) respectively. 126

Figure 4A.3 Increase in average particle diameter of PDMAEMA brush modified cup shaped particles with respect to polymerization time. 127

Figure 4A.4 Swelling ratios obtained from diameter change of the PDMAEMA brush modified particles compared to unmodified one with respect to polymerization time. 127

Figure 4A.5 Confocal Laser Scanning Microscope images of cup shaped particles (average particle diameter - $5.9\mu\text{m} \pm 0.6$) and particles modified with PDMAEMA brushes from the surface for 15 min (average particle diameter - $6.8\mu\text{m} \pm 1.6$), 30 min (average particle diameter - $7.6\mu\text{m} \pm 1.5$) and 1h (average particle diameter - $9.2\mu\text{m} \pm 1.8$). Scale bar is $5\mu\text{m}$. Dye adsorption was done at pH 7. 128

Figure 4A.6 Representative Raman spectra of (A) neat PLA (black) (B) poly(MMA-co-BEMA) (Red line) (C) cup shaped particles (blue line) (D) cup shaped particles modified with PDMAEMA brush (light blue line). Zoom in images of characteristic bands were shown in left- and right-hand sides. 129

Figure 4A.7 Brightfield image of cup shaped particles modified with PDMAEMA brush for 1 h at (a) pH - 4 (average particle diameter: $14.8\mu\text{m} \pm 3.4$) and (b) pH -7 (average particle diameter: $9.2\mu\text{m} \pm 1.8$). 131

Figure 4A.8 Confocal Laser Scanning Microscope images of dye adsorbed unmodified cup shaped particles (a) at pH -7 (b) at pH -4; dye adsorbed modified cup shaped particles (c) at pH-7 and (d) at pH -4. Scale bar is $10\mu\text{m}$. Dye adsorption experiments were performed at pH 4 and pH 7. 132

Figure 4A.9 Adsorption kinetics data for (A) Cup shaped particles (B) Modified cup shaped particles (C) Disc shaped particles (D) Modified disc shaped particles at pH – 7.	133
Figure 4A.10 Linear fitting plots of kinetic data using pseudo-second order kinetic model for (A) unmodified cup shaped particles (B) polymer brush modified cup shaped particles (C) unmodified disc shaped particles (D) polymer brush modified disc shaped particles. All experiments were carried out at pH 7.	134
Figure 4A.11 Dye adsorption at different concentration for (a) - (A) Cup shaped particles at pH – 7 (B) Cup shaped particles at pH – 4 (C) Polymer brushes modified cup shaped particles at pH – 7 (D) Polymer brushes modified cup shaped particles at pH – 4. (b) - (A) Disc shaped particles at pH – 7 (B) Polymer brushes modified disc shaped particles at pH – 7 (C) Disc shaped particles at pH – 4 (D) Polymer brushes modified disc shaped particles at pH – 4.	135
Figure 4A.12 Fit of the equilibrium data obtained at pH 7 (Figure 10) with Freundlich Isotherm model for (A) unmodified cup shaped particles and (B) polymer brush modified cup shaped particles. [R^2 is the correlation coefficient].	135
Figure 4A.13 (A) Cup shaped particles at pH – 4 (B) Cup shaped particles at pH – 7 (C) Cup shaped particles at pH – 9 (D) Polymer brushes modified cup shaped particles at pH – 4 (E) Polymer brushes modified cup shaped particles at pH – 7 (F) Polymer brushes modified cup shaped particles at pH – 9.	136
Figure 4B.1 NMR spectrum of random copolymer of methyl methacrylate and DMAEMA (2-dimethyl amino ethyl methacrylate) [poly(MMA- <i>co</i> -DMAEMA)].	149
Figure 4B.2 (a) Cup shaped particles fabricated from PLA and poly(MMA- <i>co</i> -BEMA) (75:25) (P₁), (b) Cup shaped particles fabricated from PLA and poly(MMA- <i>co</i> -DMAEMA) (75:25) (P₃) (c) Poly(DMAEMA) brush modified (polymerized for 1h) cup shaped particles (P₂).	149
Figure 4B.3 TEM images of (a) cups (P₁) (b) poly(DMAEMA) brush modified cups (P₂) (c) Enzyme immobilized P₂ particles.	152
Figure 4B.4 Evolution of amount of immobilized α -glucosidase enzyme on cup shaped particles fabricated from a blend of PLA and poly(MMA- <i>co</i> -BEMA) (75:25) (P₁), cup shaped particles fabricated from a blend of PLA and poly(MMA- <i>co</i> -DMAEMA) (75:25) (P₃) and poly(DMAEMA) brush modified cup shaped particles (P₂). Each data point represents the average of three runs.	153
Figure 4B.5 Evolution of Zeta Potential of (a) poly(DMAEMA) brush modified P₂ particles with respect to pH (b) poly(DMAEMA) brush modified particles with varying initiator (poly(MMA- <i>co</i> -BEMA) content (A) before enzyme	154

immobilization **(B)** after enzyme immobilization. Each data point represents the average value of three readings.

Figure 4B.6 Bright field images of poly(DMAEMA) brush modified cups polymerized for (a), (b) 5 min (c), (d) 15 min and (e), (f) 30 min before (left column) and after (right column) enzyme immobilization, respectively. 156

Figure 4B.7 Bright field images of (a), (b) poly(DMAEMA) brush modified cups containing 10% poly(MMA-*co*-BEMA) and (c), (d) poly(DMAEMA) brush modified cups containing 50% poly(MMA-*co*-BEMA) polymerized for 1 h; before (left column) and after (right column) enzyme immobilization, respectively. 156

Figure 4B.8 Evolution of amount of adsorbed α -glucosidase on poly(DMAEMA) brush modified cup shaped particles with varying initiator content: (poly(MMA-*co*-BEMA) – 0% (**P₁**), 10% (**P₄**), 25% (**P₂**) and 50% (**P₅**)). Each data point represents the average of three runs. 157

Figure 4B.9 SEM (Scanning Electron Microscopic) images of particles composed of PLA and poly(MMA-*co*-BEMA) (75:25) **(a)** Cups **(b)** Discs. 158

Figure 4B.10 Evolution of amount of adsorbed α -glucosidase on poly(DMAEMA) brush modified cup shaped particles polymerized for different time periods [5 min (**P₆**), 15 min (**P₇**), 30 min (**P₈**) and 60 min (**P₂**)]. Each data point represents the average of three runs. 158

Figure 4B.11 Bright field images of **(a)** cups **(b)** poly(DMAEMA) brush modified cups (c) Enzyme immobilized poly(DMAEMA) brush modified cups **(d)** Discs (e) poly(DMAEMA) brush modified discs (f) Enzyme immobilized poly(DMAEMA) brush modified discs. 160

Figure 4B.12 Evolution of amount of immobilized α -glucosidase on poly(DMAEMA) brush modified cups (**P₂**) and brush modified discs (**P₉**). Each data point represents the average of three runs. 160

Figure 4B.13 Evolution of amount of active enzyme on poly(DMAEMA) brush modified cups (**P₂**) and poly(DMAEMA) brush modified discs (**P₉**). Each data point represents the average of three runs. 161

Figure 4B.14 FTIR-ATR spectra of (A) free α -glucosidase enzyme (B) poly(DMAEMA) brush modified cups (C) α -glucosidase immobilized, poly(DMAEMA) brush modified cups. 162

Figure 4B.15 (a) Circular dichroism (CD) spectra of (A) native α -glucosidase enzyme (B) blank poly(DMAEMA) brush modified cup shaped particles (without adsorbing any enzyme) (C) desorbed enzyme from enzyme immobilized, 163

poly(DMAEMA) brush modified cup shaped particles (D) desorbed enzyme from enzyme immobilized, poly(DMAEMA) brush modified disc shaped particles (b) Enlarged part of spectra shown in (a) and (c) Percentage of α -helix, β -sheet and Random coil in neat enzyme (A), enzyme desorbed from cup shaped particles (B) and enzyme desorbed from disc shaped particles (C).

Figure 4B.16 CLSM images of α -glucosidase immobilized (a), (c) 164
poly(DMAEMA) brush modified cups (P₂) before and after enzyme
immobilization, respectively. (b), (d) poly(DMAEMA) brush modified discs (P₉)
before and after enzyme immobilization, respectively.

Figure 4B.17 (a) SEM images of magnetic nanoparticles loaded discs (b) EDX 165
(Energy dispersive X-ray) spectra of magnetic nanoparticles loaded discs.

Figure 4B.18 (a), (b) Bright field images and (c), (d) TEM images of magnetic 166
nanoparticles loaded discs and poly(DMAEMA) brush modified (polymerized for
1 h) magnetic nanoparticles loaded discs, respectively.

Figure 4B.19 TGA of (A) iron (0) magnetic nanoparticles loaded disc shaped 166
particles (B) poly(DMAEMA) brush modified iron (0) magnetic particles loaded
disc shaped particles

Figure 4B.20 Photos of (a), (c) & (e) brush modified particles (polymerized for 168
5 min) without any magnetic nanoparticles, kept undisturbed for 0 min, 20 min
and 50 min, respectively; Photos of (b), (d) and (f) brush modified particles
(polymerized for 5 min) with magnetic nanoparticles, kept undisturbed for 0 min,
20 min and 50 min, respectively.

Figure 4B.21 Bright field image of poly(DMAEMA) brush modified magnetic 168
nanoparticles loaded disc particles (polymerized for 5 min).

Figure 4B.22 Retention of activity of enzyme after storing the P₂ particles for 30 169
days. Each data point represents the average of three runs.

Figure 5.1 Scanning electron microscopic (SEM) images of Janus particles (a) 186
PLA in one compartment and PLA/poly(MMA-co-BEMA) (90/10) in second
compartment (P₁₀) (b) PLA in one compartment and PLA/poly(MMA-co-BEMA)
(75/25) in second compartment (P₂₅) (c) PLA in one compartment and
PLA/poly(MMA-co-BEMA) (50/50) in second compartment (P₅₀).

Figure 5.2 Brightfield images of Janus particles (a) PLA in one compartment and 187
PLA/poly(MMA-co-BEMA) (90/10) in second compartment (P₁₀) (b)) PLA in
one compartment and PLA/poly(MMA-co-BEMA) (75/25) in second
compartment (P₂₅) (c) PLA in one compartment and PLA/poly(MMA-co-BEMA)
(50/50) in second compartment (P₅₀) (d) brush modified Janus microparticles,
having PLA in one compartment and PLA/poly(MMA-co-BEMA) (90/10) in

second compartment (**P₁₀**) (e) brush modified Janus microparticles, PLA in one compartment and PLA/poly(MMA-co-BEMA) (75/25) in second compartment (**P₂₅**) (f) brush modified Janus microparticles, PLA in one compartment and PLA/poly(MMA-co-BEMA) (50/50) in second compartment (**P₅₀**).

Figure 5.3 Confocal Laser Scanning Microscopic images of Janus particles (a) 188
 PLA in one compartment and PLA/poly(MMA-co-BEMA) (90/10) in second
 compartment (**P₁₀**) (b)) PLA in one compartment and PLA/poly(MMA-co-
 BEMA) (75/25) in second compartment (**P₂₅**) (c) PLA in one compartment and
 PLA/poly(MMA-co-BEMA) (50/50) in second compartment (**P₅₀**) (d) brush
 modified Janus microparticles, having PLA in one compartment and
 PLA/poly(MMA-co-BEMA) (90/10) in second compartment (**P₁₀**) (e) brush
 modified Janus microparticles, PLA in one compartment and PLA/poly(MMA-
 co-BEMA) (75/25) in second compartment (**P₂₅**) (f) brush modified Janus
 microparticles, PLA in one compartment and PLA/poly(MMA-co-BEMA)
 (50/50) in second compartment (**P₅₀**).

Figure 5.4 Differential Interference Contrast images of Janus particles (a) 189
 in one compartment and PLA/poly(MMA-co-BEMA) (90/10) in second
 compartment (**P₁₀**) (b)) PLA in one compartment and PLA/poly(MMA-co-
 BEMA) (75/25) in second compartment (**P₂₅**) (c) PLA in one compartment and
 PLA/poly(MMA-co-BEMA) (50/50) in second compartment (**P₅₀**) (d) brush
 modified Janus microparticles, having PLA in one compartment and
 PLA/poly(MMA-co-BEMA) (90/10) in second compartment (**P₁₀**) (e) brush
 modified Janus microparticles, PLA in one compartment and PLA/poly(MMA-
 co-BEMA) (75/25) in second compartment (**P₂₅**) (f) brush modified Janus
 microparticles, PLA in one compartment and PLA/poly(MMA-co-BEMA)
 (50/50) in second compartment (**P₅₀**).

Figure 5.5 Scanning electron microscopic (SEM) images of Janus particles (a) 190
 PLA in one compartment and PLA/poly(MMA-co-BEMA) (90/10) in second
 compartment (**P₁₀**) (b) brush modified Janus microparticles, having PLA in one
 compartment and PLA/poly(MMA-co-BEMA) (90/10) in second compartment
 (**P₁₀**).

Figure 5.6 Fourier Transform Infrared Spectroscopy (FTIR) spectra of Janus 190
 particles (a) PLA in one compartment and PLA/poly(MMA-co-BEMA) (50/10)
 in second compartment (**P₅₀**) (b) brush modified Janus microparticles, having
 PLA in one compartment and PLA/poly(MMA-co-BEMA) (50/10) in second
 compartment (**P₅₀**).

Figure 5.7 Brightfield images of Janus particles (a) PLA in one compartment and 191
 PLA/poly(MMA-co-BEMA) (50/50) in second compartment (**P₅₀**). Brush
 modified Janus microparticles, having PLA in one compartment and

PLA/poly(MMA-co-BEMA) (50/50) in second compartment (**P₅₀**), polymerized for (b) 5 min, (c) 15 min, (d) 30 min and (e) 60 min.

Figure 5.8 Brightfield images of Janus particles (a) PLA in one compartment and PLA/poly(MMA-co-BEMA) (90/10) in second compartment (**P₅₀**). Brush modified Janus microparticles, having PLA in one compartment and PLA/poly(MMA-co-BEMA) (90/10) in second compartment (**P₅₀**), polymerized for 0h, 15 min, 30 min and 60 min. 192

Figure 5.9 (A) Schematic for Pickering emulsion stabilized by Janus particles composed of PLA in one compartment and PLA/poly(MMA-co-BEMA) in second compartment (B) Digital photos of Pickering emulsion stabilized by brush modified Janus microparticles **P₁₀**, **P₂₅** and **P₅₀**. 194

Figure 5.10 Brightfield images of (A) (a) Pickering emulsion stabilized by Janus particles **P₁₀** (b) zoom in image (B) (a) Pickering emulsion stabilized by Janus particles **P₂₅** (b) zoom in image (c) fluorescent image (B) a) Pickering emulsion stabilized by Janus particles **P₅₀** (b) zoom in image. 195

Figure 5.11 Top image: Digital photos of emulsions made at different pHs; Bottom image: Brightfield images of (a) brush modified Janus particles **P₅₀** at pH – 4 (b) brush modified Janus particles **P₅₀** at pH – 7 (b) brush modified Janus particles **P₅₀** at pH – 10. 195

Figure 5.12 Thermal Gravimetric analysis of (A) GNP synthesized onto brush modified Janus particles **P₁₀** at H_{AuCl₄} concentration of (a) 5mM, (b) 15mM (c) 30mM (d) 50mM (e) 0mM, (B) GNP synthesized brush modified Janus particles (a) **P₁₀** (b) **P₂₅** (c) **P₅₀** at H_{AuCl₄} concentration of 50mM, (C) GNP synthesized onto brush modified Janus particles **P₅₀**, polymerized for (a) 5 min (b) 15 min (c) 30min (d) 60 min at H_{AuCl₄} concentration of 50mM. 197

Figure 5.13 Field Emission Scanning Electron Microscope (FESEM) of (a) Janus particles **P₁₀** (b) GNP synthesized brush modified Janus particles **P₁₀** (c) Transmission electron microscopic image of GNP synthesized brush modified Janus particles **P₁₀**. 198

Figure 5.14 (a) Energy Dispersive X- ray (EDX) mapping image of Janus particles **P₁₀** (b) Field Emission Scanning Electron Microscope (FESEM) of Janus particles **P₁₀** (c) EDX spectra. 198

Figure 5.15 UV-Vis spectra of *p*-nitrophenol reduction to *p*-nitroaniline reduction catalysed by GNPs synthesized **P₅₀** at H_{AuCl₄} concentration of (A) 5 mM (B) 15mM and (C) 50mM. 200

Figure 5.16 Brightfield images of (a) iron (0) nanoparticles (in PLA compartment) loaded Janus particles (b) brush modified iron (0) nanoparticles (in 201

PLA compartment) loaded Janus particles **P₅₀** (polymerized for 15 min) (c) GNPs synthesized (15mM HAuCl₄) brush modified iron (0) nanoparticles (in PLA compartment) loaded Janus particles **P₅₀** (polymerized for 15 min).

Figure 5.17 Transmission electron Microscopy (TEM) images of (a) iron (0) nanoparticles (in PLA compartment) loaded Janus particles (b) brush modified iron (0) nanoparticles (in PLA compartment) loaded Janus particles **P₅₀** (polymerized for 15 min) (c) GNPs synthesized (15mM HAuCl₄) brush modified iron (0) nanoparticles (in PLA compartment) loaded Janus particles **P₅₀** (polymerized for 15 min). 202

Figure 5.18 (a) Proton NMR study for dechlorination of trichloroethylene (TCE) in octanol phase of octanol/water emulsion by iron (0) nanoparticles loaded (in PLA compartment only) brush modified **P₅₀** particles (polymerized for 15 min) **(b)** % conversion of TCE with respect to time. 202

Figure 5.19 Proton NMR study for dechlorination of trichloroethylene (TCE) present in octanol phase of octanol/water emulsion by GNPs immobilized (15mM of HAuCl₄), iron (0) nanoparticles loaded (in PLA compartment only) brush modified **P₅₀** (polymerized for 15 min) **(b)** % conversion of TCE with respect to time. 203

Figure 5.20 Proton NMR study for dechlorination of trichloroethylene (TCE) by iron (0) nanoparticles loaded (in PLA compartment only) brush modified **P₅₀** particles (polymerized for 15 min) present in water medium **(b)** % conversion of TCE with respect to time. 203

LIST OF TABLES

	Page No.
Table 1.1 Fabrication methods, compositions, shapes and applications of shape anisotropic and compositional anisotropic particles.	31
Table 1.2 Fabrication methods, compositions, surface modifications and applications of surface anisotropic particles.	42
Table 2.1 Morphology and particle size of electrojetted samples at a concentration of 1 wt% with different PLA content*	78
Table 2.2 Electrical conductivity, viscosity and surface tension of electrojetted solutions with different content of PLA*	80
Table 2.3 Morphology and particle size obtained for sample 4 jetted from different solvent systems*	80
Table 2.4 Electrical conductivity, surface tension and viscosity of electrojett solutions made in different solvent systems*	81
Table 2.5 Morphology and particle size of eletrojetted samples (sample 4) at different polymer concentrations made in chloroform:DMF (95:5)*	83
Table 2.6 Electrical conductivity, surface tension and viscosity of electrojetted solutions made at different concentrations*	83
Table 2.7 Morphology and particle size of electrojetted samples (sample 4) jetted at different flow rates*	86
Table 3.1 Raman characteristic bands	104
Table 3.2 DPD interaction parameters between different components at T=300 K	109
Table 4A.1 Raman characteristic bands	129
Table 4A.2 Linear fit data for the dye adsorption kinetics (pseudo second order, PSO).	133
Table 4B.1 Average diameter of different particle systems before and after enzyme immobilization.	150
Table 5.1 Average size of different Janus particle system.	192

LIST OF ABBREVIATIONS

RNA	Ribonucleic Acid
DNA	Deoxyribonucleic Acid
PLGA	Poly(Lactic-Co-Glycolic Acid
PDMS	Poly(dimethylsiloxane)
TPGDA	Tripropyleneglycol Diacrylate
DVB	Divinylbenzene
DMOS	Dimethacrylate Oxypropyldimethylsiloxane
PEG-DA	Poly(ethylene glycol) Diacrylate
PLA	Poly(lactic acid)
DMAEMA	2-dimethylamino Ethyl Methacrylate
PDMAEMA	Poly(2-dimethylamino ethyl methacrylate)
ATRP	Atom Transfer Radical Polymerization
SIATRP	Surface Initiated Atom Transfer Radical Polymerization
PMMA	Poly(methylmethacrylate)
SEM	Scanning Electron Microscopy
TMEDA	Tetramethylethylenediamine
SORP	Self-organization Rapid Precipitation
PFS	Poly(ferrocenylsilane)
P2VP	Poly(2-vinylpyridine)
CTAB	Cetyl Trimethylammonium Bromide
CTAB-OH	Hydroxy-cetyl Trimethylammonium Bromide
PISA	Polymerization Induced Self-assembly
PMAAz	Azobenzene Containing Poly(methacrylate)
PMAA	Poly(methacrylic acid)
LC	Liquid Crystal
T _g	Glass Transition Temperature
PLGE	Poly(lactide- <i>co</i> -glycolide- <i>b</i> -ethylene glycol- <i>b</i> -lactide- <i>co</i> -glycolide)
NR	Nile Red
PEMA	Poly(ethylene-alt-maleic acid)
PVA	Poly(vinyl alcohol)
DOPC	1,2-dioleoyl-sn-glycero-3-phosphocholine
PRINT	Particle Replication in Non-wetting Templates
PFPE	Perfluoropolyether
PCL	Poly(ϵ -caprolactone)
PVP	Poly(vinyl pyrrolidone)
PEO	Poly(ethylene oxide)
PEG	Poly(ethylene glycol)
EHDC	Electrohydrodynamic Co-jetting
PD	Parkinson's Disease
LD	Levodopa
CD	Carbidopa

PEO	Poly(ethylene oxide)
CLSM	Confocal Laser Scanning Microscope
PEI	Poly(ethyleneimine)
SERS	Surface-Enhanced Raman Scattering
PVCi	Poly(vinyl cinnamate)
PLLA	Poly(L-lactic acid)
O/W	Oil-in-Water
W/O	Water-in-Oil
PAAm	Poly(acrylamide)
PNIPAM	Poly(N-isopropylacrylamide)
DCC	N,N'-dicyclohexylcarbodiimide
DMAP	4-dimethylaminopyridine
TRITC	Tetramethyl Rhodamine Isothiocyanate
BSA	Bovine Serum Albumin
RAFT	Reversible Addition-Fragmentation Chain Transfer
ROMP	Ring Opening Metathesis Polymerization
NMP	Nitroxide Mediated Polymerization
HEMA	2-Hydroxy Ethyl Methacrylate
pHEMA	Poly(2-hydroxy ethyl methacrylate)
LAMMPS	Large-scale Atomic/Molecular Massively Parallel Simulator
DPD	Dissipative Particle Dynamics
MPS	Mononuclear Phagocyte System
ID	Injected Dose
HUVEC	Human Umbilical Vein Endothelial Cells
BMDC	Bone Marrow Dendritic Cells
EDTA	Ethylene Diamine Tetraacetate
NMR	Nuclear Magnetic Resonance
GPC	Gel Permeation Chromatography
HPLC	High Performance Liquid Chromatography
DSC	Differential Scanning Calorimetry
DMF	Dimethylformamide
THF	Tetrahydrofuran
FTIR	Fourier Transform Infrared Spectroscopy
BEMA	2-(2-Bromopropionyloxy) Ethyl Methacrylate
DIC	Differential Interference Contrast
DLS	Dynamic Light Scattering
OM	Optical Microscopy
AFM	Atomic Force Microscopy
TGA	Thermal Gravimetric Analysis
<i>p</i> -NP	<i>p</i> -nitrophenol
<i>p</i> -PNPG	<i>p</i> -nitrophenyl- α -D-glucopyranoside
PMDTA	Pentamethyl Diethylene Tetraacetate
EDX	Energy Dispersive X-ray

CD	Circular Dichroism Spectroscopy
ATR-FTIR	Attenuated Total Reflectance-Fourier Transform Infrared
TEM	Transmission Electron Microscope
ZP	Zeta Potential
SR	Swelling Ratio
TMPTA	Trimethylopropane triacrylate
LPS	Linear Polystyrene
FESEM	Field Emission Scanning Electron Microscopic
GNPs	Gold Nanoparticles
TCE	Trichloroethylene