

# **QUANTITATIVE DIFFUSION-WEIGHTED IMAGING IN CANCER APPLICATIONS**

**ARCHANA VADIRAJ MALAGI**



**CENTRE FOR BIOMEDICAL ENGINEERING  
INDIAN INSTITUTE OF TECHNOLOGY DELHI**

**DECEMBER 2022**

© Indian Institute of Technology Delhi (IITD), New Delhi, 2022

# **Quantitative Diffusion-Weighted Imaging in Cancer Applications**

*A thesis submitted in partial fulfilment of  
the requirements for the degree of*

**Doctor of Philosophy**

*in*

**Biomedical Engineering**

*by*

**Archana Vadiraj Malagi**

**Entry No. 2017BMZ8358**

*Under the guidance of*

**Dr. Amit Mehndiratta and Dr. Devasenathipathy Kandasamy**



**Centre for Biomedical Engineering,  
Indian Institute of Technology Delhi**

**December 2022**

*Dedicated to Mom, Dad,  
Aravind, Anil*

# Certificate

This is to certify that the work contained in this thesis titled “**Quantitative Diffusion-weighted Imaging in Cancer Applications**” is a bonafide record of the work done at Indian Institute of Technology, Delhi by **Archana Vadiraj Malagi** (2017BMZ8358) for the award of **Doctor of Philosophy** in Biomedical Engineering. The work in this thesis was conducted under our supervision and guidance. The matter submitted in this thesis has not been submitted elsewhere for the award of any other degree.



**Dr. Amit Mehndiratta**

Associate Professor  
Centre for Biomedical Engineering  
Indian Institute of Technology Delhi  
New Delhi-110016



**Dr. Devasenathipathy Kandasamy**

Additional Professor  
Department of Radiodiagnosis  
All India Institute of Medical Sciences  
New Delhi-110029

**Date:** 2<sup>nd</sup> August 2022

**Place:** New Delhi

## Acknowledgement

*It brings me great pleasure to convey my gratitude to everyone who has assisted and supported me over the past few years while I have been working on my PhD thesis.*

*It is a huge honor to have **Dr. Amit Mehndiratta** as my PhD supervisor. I would want to express my profound gratitude for his mentorship, advice, and unending support during the whole of my graduate studies. He has offered me extensive research resources, including limitless exposure to MRI techniques and clinical collaborations with radiologists and clinicians, for which I am very grateful. His wide expertise and excellent foundation in MR research helped me overcome project challenges. He has always pushed me to think independently as a researcher and complete all tasks in advance. His dedication of academic achievement, and his work ethic have inspired me beyond research and will undoubtedly be beneficial to my career after completing my PhD.*

*I am obliged to my PhD co-supervisor, **Dr. Devasenathipathy Kandasamy** Additional professor, Department of Radiodiagnosis, AIIMS, New Delhi. It was a major challenge to establish an MR procedure since early efforts to provide high-quality images for the lymphoma study. His extensive understanding of complex techniques assisted us in optimizing image quality. He has been an invaluable resource for my research, providing me with enough time on MR scanners, instructive feedback on my work and the medical imaging field.*

*I would like to thank my doctoral committee members in helping me better my dissertation through their insightful remarks and engaging conversations during presentations. I am grateful to my committee chairperson, **Dr. S. D. Joshi**, who have always encouraged to me think in-depth in the subject matter and provided his constructive comments. Dr. Joshi was consistently interested in learning new approaches throughout my presentation with engaging questions. I would like to convey my heartfelt gratitude to **Dr. Kedar Khare** for*

*his insightful analysis, recommendations, and direction in the development and refinement of my thesis.*

*I must thank **Dr. Anup Singh** for his constant support and encouragement by taking all lab members and engage us in fruitful discussions. His insightful advice throughout my doctoral dissertation improved my thesis. I would like to extend my gratitude to everyone on the faculty advisor as well as the staff members at our center.*

*Next, I would like to express my gratitude to **Dr. Sameer Bakhshi** and **Dr. Deepam Pushpam**, IRCH, AIIMS, New Delhi, for facilitating the recruitment of patients at their OPD at IRCH, AIIMS, Delhi. Dr. Bakhshi gave me invaluable guidance on "how to complete a successful PhD." He said, a successful completion of a PhD thesis requires completion of the following phases: project design; data collection (patient recruitment and data processing); novel technique development; data collation and publication.*

*I am grateful to **Dr. Rakesh Kumar**, Department of Nuclear Medicine, AIIMS, New Delhi, and staffs who endlessly helped me in data acquisition and collection of PET imaging. I would like to thank **Dr. Raju Sharma**, Department of Radiology, AIIMS, Delhi, for his insightful comments, advise, and encouragement during my PhD. I would also want to gratefully thank **Dr. Chandan J. Das** for his assistance in providing prostate imaging data and his contribution to the thesis from a clinical perspective.*

*I am grateful to the MR radiographers, notably **Mr. Udit** and **Mr. Lalit**, Department of Radiodiagnosis, AIIMS, New Delhi, who never refused to scan the patients for my thesis, even when they had to give their additional time. I must appreciate the nurse personnel and support staff of the medical oncology outpatient department (OPD) at IRCH, AIIMS, Delhi, for their assistance during patient recruitment. My doctoral thesis would be incomplete without my sincere appreciation for the patients who participated in this research by providing informed consent and cooperating with MRI acquisition.*

*It has been a great pleasure for me to work with the entire MedImg group of talented researchers and friendly members, **Dr. Neha**, **Dr. Sneha**, **Dr. Anirban**, **Dr. Ayan**, **Dr. Rupsa**, **Dr. Dinil**, **Dr. Rafeek**, **Sandeep**, **Dharmesh**, **Umang**, **Pradyumna**, **Virendra**, **Sneha**, **Debasish**, **Raufiya**, **Ankit**, **Himanshu**, **Piyush**, **Satyajit**, **Anshika**, **Ganeshkumar**, and **James** and my fellow batchmates, **Anjali**, **Ahana**, **Aayushi**, and **Sagar**. I would like to*

*specially thank **Dr. Esha** for helping me in developing methodologies and mentoring throughout my PhD journey.*

*I would like to express my sincere gratitude to the Ministry of Human Resource Development of the Government of India for giving the funding for my PhD research scholarship and SERB, Department of Science and Technology, the Government of India for funding the project. I also would like to thank the CSIR and IIT Delhi Research Scholar Fund for giving me travel funding so I could attend conferences internationally.*

*I am deeply indebted to my family – my proud parents, **Asha** and **Vadiraj Malagi**, who supported my desire to pursue an academic career and have been there through everything, for providing me with emotional support and guidance. Without your love and guidance, I would not be where I am today, for everything, thank you.*

*I want to express my gratitude to my brother, **Aravind** and **Akhila** for making me an aunt to my beautiful niece **Anvika** and for always being there for me. I am grateful to Anil's parents, **Pushpa** and **Raghavendra Mugali**, and **Anantha**, for welcoming me into their family. I am very grateful to my closest friends, **Chandrani**, **Pavitra**, **Karpur**, **Suprava**, **Dipanwita**, and **Charmi** for being by my side.*

*Finally, my lovely husband, **Anil**, I cannot express how thankful I am to have had you by my side during this whole process. When I succeed, you applaud me, and when I stumble, you encourage me. Your patience and kindness have made me a better person.*

*Archana Vadiraj Malagi*

# Abstract

Quantitative magnetic resonance imaging (MRI) quantifies multiple factors utilizing tissue-specific physical units. These quantitative metrics are sensitive to physiological tissue changes and pathologies. In recent years, demand for non-contrast quantitative MRI has grown, such as intravoxel incoherent motion combined with diffusion kurtosis imaging (IVIM-DKI). IVIM-DKI is a diffusion-weighted imaging technique that uses low-to-high diffusion sensitizing gradient strengths ( $b$ -value). IVIM-DKI doesn't require contrast agents, ensuring application safety to a wide variety of patients and reducing MRI costs. Although IVIM-DKI is beneficial for clinical studies, it has not yet been widely used in routine clinical practices because of a few technical problems: (i) the signal-to-noise ratio (SNR) of the IVIM-DKI signal is poor, (ii) piecewise simultaneous fitting of parameters result in noisy and unreliable parameter maps, (iii) the diagnostic utility of IVIM-DKI in a various clinical setting at different MRI field strength has not been proven, and (iv) the absence of a standardized protocol for organ-specific imaging leads to prolonged scan times. Thus, this thesis aims to address some of these challenges by proposing novel strategies for improving IVIM-DKI parametric map estimation using parametric reconstruction method validation of the novel model at different MRI field strengths and optimization of  $b$ -values for protocol standardization, which can promote the role of IVIM-DKI in the improved quantitative lesion characterization technique in cancer applications.

The first objective was to develop a novel strategy to improve the parameter estimation of IVIM-DKI for producing high-quality parametric maps. To implement this objective, the total three-dimensional total variation (TV) penalty function was utilized in conjunction with non-linear least square optimization (NLLS) of the IVIM-DKI model (IDTV model). TV is an image-based reconstruction approach that corrects for NLLS error and adaptively removes any abrupt changes in the map while preserving the edges. Two models were implemented for the estimation of diffusion coefficient ( $D$ ), perfusion coefficient ( $D^*$ ), perfusion fraction ( $f$ ), and kurtosis ( $k$ ) using either: (1) standard model and (2) IDTV model. Experimental simulation results showed accurate and high-quality IVIM-DKI parameter maps, particularly at low signal-to-noise ratios in simulations. Additionally, TV regularization was resilient to changes in SNR, resulting in less error and bias in IVIM-DKI parameter maps.

The second objective was to examine the clinical usefulness of IVIM-DKI analysis using the IDTV model in prostate mass characterization at different MRI field strengths (1.5T and 3T). The IDTV model resulted in enhanced parameter estimation with minimal error at 1.5T and 3T. Furthermore, compared to the standard model at 3T, the IDTV model at 1.5T resulted in lower parameter estimate error and higher-quality parametric reconstruction.  $D$ ,  $f$ , and  $k$  estimated with the IDTV model showed higher diagnostic performance than the standard model. Hence, IVIM-DKI might play an essential role in prostate lesion diagnosis when paired with the IDTV model.

The third objective was to assess the efficacy of IVIM-DKI analysis with the IDTV model and machine learning-based multi-parametric texture analysis in characterizing pancreatic masses. Perfusion fraction was found to out-perform the diffusion measures (ADC and  $D$ ) in discriminating pancreatic adenocarcinoma (PDAC) from the pancreatic neuroendocrine

tumor (pNET). Whole-volumetric texture analysis of individual or combination IVIM-DKI parameters with machine learning-based classification of pancreatic masses can help with the non-invasive characterization of pancreatic lesions.

The fourth objective was to optimize the b-value for IVIM and IVIM-DKI analyses by combining different b-values. IVIM and IVIM-DKI signals were modeled utilizing the IVIM model with the TV method (BE+TV model) and the IDTV model, respectively. The experimental findings of simulation and clinical data demonstrated that the BE+TV and IDTV models were resistant to b-value combinations. Furthermore, both models were robust to any number and combination of b-values, and they improved the quality of all parametric maps at low SNR. The BE+TV approach significantly enhanced the quality of D and f maps. The IDTV model, on the other hand, improved the quality of D, f, and k maps. The acquisition of IVIM and IVIM-DKI with optimal 8 b-values may be employed for faster acquisition times and high-quality parameter maps with comparable accuracy as 13b-values using the standard model with less acquisition time.

Finally, this thesis concluded by using an IVIM-DKI model with an optimized b-value for diagnosing malignant lymph nodes compared to fluorodeoxyglucose-positron emission tomography with computed tomography (FDG-PET/CT) imaging. The preliminary findings revealed that the IDTV model outperformed PET imaging in identifying benign and malignant lymph nodes in lymphoma and further classifying lymphoma subtypes. In diagnosing malignant lymph nodes, IVIM-DKI parameters showed higher diagnostic performance than ADC and PET parameters. IVIM-DKI can potentially be used for lymph node evaluation in lymphoma and is comparable to PET imaging.

## सार

मात्रात्मक चुंबकीय अनुनाद इमेजिंग (एमआरआई) ऊतक-विशिष्ट भौतिक इकाइयों का उपयोग करने वाले कई कारकों को मापता है। ये मात्रात्मक मेट्रिक्स शारीरिक ऊतक परिवर्तन और रोग विकृति के प्रति संवेदनशील हैं। हाल के वर्षों में, गैर-विपरीत मात्रात्मक एमआरआई की मांग बढ़ी है, जैसे कि प्रसार कर्टोसिस इमेजिंग (आईवीआईएम-डीकेआई) के साथ संयुक्त इंट्रावॉक्सेल असंगत गति। आईवीआईएम-डीकेआई एक डिफ्यूजन-वेटेड इमेजिंग तकनीक है जो लो-टू-हाई डिफ्यूजन सेंसिटाइजिंग ग्रेडिएंट स्ट्रेंथ (बी-वैल्यू) का उपयोग करती है। आईवीआईएम-डीकेआई को कंट्रास्ट एजेंटों की आवश्यकता नहीं है, जो विभिन्न प्रकार के रोगियों के लिए आवेदन सुरक्षा सुनिश्चित करते हैं और एमआरआई लागत को कम करते हैं। हालांकि आईवीआईएम-डीकेआई नैदानिक अध्ययनों के लिए फायदेमंद है, लेकिन कुछ महत्वपूर्ण तकनीकी समस्याओं के कारण इसे अभी तक नियमित नैदानिक प्रथाओं में व्यापक रूप से उपयोग नहीं किया गया है: (i) आईवीआईएम-डीकेआई सिग्नल का सिग्नल-टू-शोर अनुपात (एसएनआर) खराब है, (ii) मापदंडों की एक साथ फिटिंग से शोर और अविश्वसनीय पैरामीटर मानचित्र उत्पन्न होते हैं, (iii) विभिन्न एमआरआई क्षेत्र की ताकत पर विभिन्न नैदानिक सेटिंग में आईवीआईएम-डीकेआई की नैदानिक उपयोगिता, और (iv) अंग के लिए एक मानकीकृत प्रोटोकॉल की अनुपस्थिति- विशिष्ट इमेजिंग लंबे समय तक स्कैन समय की ओर ले जाती है। इस प्रकार, इस थीसिस का उद्देश्य आईवीआईएम-डीकेआई पैरामीट्रिक मानचित्र अनुमान में सुधार के लिए नई रणनीतियों का प्रस्ताव करके इन चुनौतियों का समाधान करना है, जो विभिन्न

एमआरआई क्षेत्र की ताकत पर उपन्यास मॉडल के पैरामीट्रिक पुनर्निर्माण विधि सत्यापन और प्रोटोकॉल मानकीकरण के लिए बी-वैल्यू के अनुकूलन का उपयोग कर सकते हैं, जो बढ़ावा दे सकते हैं कैंसर अनुप्रयोगों में बेहतर मात्रात्मक घाव लक्षण वर्णन तकनीक में आईवीआईएम-डीकेआई की भूमिका।

पहला उद्देश्य उच्च गुणवत्ता वाले पैरामीट्रिक मानचित्रों के निर्माण के लिए आईवीआईएम-डीकेआई के पैरामीटर अनुमान में सुधार के लिए एक नई रणनीति विकसित करना था। इस उद्देश्य को लागू करने के लिए, आईवीआईएम-डीकेआई मॉडल (आईडीटीवी मॉडल) के नॉन-लीनियर लीज स्क्वायर ऑप्टिमाइज़ेशन (एनएलएलएस) के संयोजन के साथ त्रि-आयामी टोटल टोटल वेरिएशन (टीवी) पेनल्टी फ़ंक्शन का उपयोग किया गया था। टीवी एक छवि-आधारित पुनर्निर्माण दृष्टिकोण है जो एनएलएलएस त्रुटि के लिए सुधार करता है और किनारों को संरक्षित करते हुए मानचित्र में किसी भी अचानक परिवर्तन को अनुकूल रूप से हटा देता है। प्रसार गुणांक (डी), छिड़काव गुणांक (डी \*), छिड़काव अंश (एफ), और कर्टोसिस (के): (1) मानक मॉडल और (2) आईडीटीवी मॉडल के आकलन के लिए दो मॉडल लागू किए गए थे। प्रायोगिक सिमुलेशन परिणामों ने सटीक और उच्च-गुणवत्ता वाले आईवीआईएम-डीकेआई पैरामीटर मानचित्र दिखाए, विशेष रूप से सिमुलेशन में कम सिग्नल-टू-शोर अनुपात पर। इसके अतिरिक्त, टीवी नियमितीकरण एसएनआर में परिवर्तन के लिए लचीला था, जिसके परिणामस्वरूप आईवीआईएम-डीकेआई पैरामीटर मानचित्रों में कम त्रुटि और पूर्वाग्रह था।

दूसरा उद्देश्य विभिन्न एमआरआई क्षेत्र शक्तियों (1.5 टेस्ला और 3 टेस्ला) में प्रोस्टेट मास लक्षण वर्णन में आईडीटीवी मॉडल का उपयोग करके आईवीआईएम-डीकेआई विश्लेषण की नैदानिक उपयोगिता की जांच करना था। आईडीटीवी मॉडल ने 1.5 टेस्ला और 3 टेस्ला पर न्यूनतम त्रुटि के साथ पैरामीटर अनुमान बढ़ाया। इसके अलावा, 3 टेस्ला पर मानक मॉडल की तुलना में, 1.5 टेस्ला पर आईडीटीवी मॉडल के परिणामस्वरूप कम पैरामीटर अनुमान त्रुटि और उच्च गुणवत्ता वाला पैरामीट्रिक पुनर्निर्माण

हुआ। आईटीटीवी मॉडल के साथ अनुमानित डी, एफ और के ने मानक मॉडल की तुलना में उच्च नैदानिक प्रदर्शन दिखाया। इसलिए, आईटीटीवी मॉडल के साथ जोड़े जाने पर आईवीआईएम-डीकेआई प्रोस्टेट घाव के निदान में एक आवश्यक भूमिका निभा सकता है।

तीसरा उद्देश्य आईटीटीवी मॉडल और मशीन लर्निंग-आधारित बहु-पैरामीट्रिक बनावट विश्लेषण के साथ आईवीआईएम-डीकेआई विश्लेषण की प्रभावकारिता का आकलन करना था। अग्राशयी न्यूरोएंडोक्राइन ट्यूमर (पीएनईटी) से विभेदक अग्राशयी एडेनोकार्सिनोमा (पीडीएसी) में प्रसार उपायों (एडीसी और डी) को बाहर करने के लिए छिड़काव अंश पाया गया था। अग्राशयी द्रव्यमान के मशीन लर्निंग-आधारित वर्गीकरण के साथ व्यक्तिगत या संयोजन आईवीआईएम-डीकेआई मापदंडों का संपूर्ण-वॉल्यूमेट्रिक बनावट विश्लेषण अग्राशय के घावों के गैर-आक्रामक लक्षण वर्णन में मदद कर सकता है।

चौथा उद्देश्य विभिन्न बी-मानों को मिलाकर आईवीआईएम और आईवीआईएम-डीकेआई विश्लेषणों के लिए बी-वैल्यू का अनुकूलन करना था। आईवीआईएम और आईवीआईएम-डीकेआई सिग्नलिंग का विश्लेषण आईवीआईएम मॉडल का उपयोग करके क्रमशः टीवी विधि (बीई+टीवी मॉडल) और आईटीटीवी मॉडल के साथ किया गया था। सिमुलेशन और क्लिनिकल डेटा के प्रयोगात्मक निष्कर्षों ने प्रदर्शित किया कि बीई+टीवी और आईटीटीवी मॉडल बी-वैल्यू संयोजनों के प्रतिरोधी थे। इसके अलावा, दोनों मॉडल किसी भी संख्या और बी-मानों के संयोजन के लिए मजबूत थे, और उन्होंने कम एसएनआर पर सभी पैरामीट्रिक मानचित्रों की गुणवत्ता में सुधार किया। बीई + टीवी दृष्टिकोण ने डी और एफ मानचित्रों की गुणवत्ता में काफी वृद्धि की है। दूसरी ओर, आईटीटीवी मॉडल ने डी, एफ और के मानचित्रों की गुणवत्ता में सुधार किया। इष्टतम 8 बी-मानों के साथ आईवीआईएम और आईवीआईएम-डीकेआई के अधिग्रहण को कम अधिग्रहण समय के साथ मानक मॉडल का उपयोग

करते हुए 13 बी-मानों के रूप में तुलनीय सटीकता के साथ तेजी से अधिग्रहण के समय और उच्च गुणवत्ता वाले पैरामीटर मानचित्रों के लिए नियोजित किया जा सकता है।

अंत में, मानक एफडीजी-पीईटी/सीटी इमेजिंग की तुलना में घातक लिम्फ नोड्स के निदान के लिए एक अनुकूलित बी-मान के साथ आईवीआईएम-डीकेआई मॉडल का उपयोग करके इस थीसिस का निष्कर्ष निकाला गया। प्रारंभिक निष्कर्षों से पता चला है कि आईडीटीवी मॉडल ने लिम्फोमा में सौम्य और घातक लिम्फ नोड्स की पहचान करने और लिम्फोमा उपप्रकारों को वर्गीकृत करने में पीईटी इमेजिंग से बेहतर प्रदर्शन किया। घातक लिम्फ नोड्स के निदान में, आईवीआईएम-डीकेआई मापदंडों ने एडीसी और पीईटी मापदंडों की तुलना में उच्च नैदानिक प्रदर्शन दिखाया। आईवीआईएम-डीकेआई संभावित रूप से लिम्फोमा में लिम्फ नोड मूल्यांकन के लिए इस्तेमाल किया जा सकता है और पीईटी इमेजिंग के बराबर है।

# Contents

<b>CERTIFICATE</b> .....	<b>I</b>
<b>ACKNOWLEDGEMENT</b> .....	<b>III</b>
<b>ABSTRACT</b> .....	<b>VII</b>
<b>CONTENTS</b> .....	<b>XV</b>
<b>LIST OF FIGURES</b> .....	<b>XXI</b>
<b>LIST OF TABLES</b> .....	<b>XXIX</b>
<b>GLOSSARY</b> .....	<b>XXXI</b>
<b>CHAPTER 1: INTRODUCTION AND MOTIVATION</b> .....	<b>1</b>
1.1    DIFFUSION-WEIGHTED IMAGING IN CANCER APPLICATIONS .....	1
1.2    CHALLENGES OF IVIM-DKI IN CLINICAL ROUTINE STANDARDS .....	4
1.3    THESIS OBJECTIVES .....	5
1.4    DATA INFORMATION AND MRI PROTOCOL .....	7
1.4.1 <i>Prostate cancer</i> .....	7
1.4.1.1    Clinical data information.....	7
1.4.1.2    MRI acquisition for prostate gland.....	8
1.4.2 <i>Pancreatic masses</i> .....	9
1.4.2.1    Clinical data information.....	9
1.4.2.2    MRI acquisition for pancreas .....	10
1.4.3 <i>Lymphoma</i> .....	11
1.4.3.1    Clinical data information.....	11
1.4.3.2    Whole-body FDG-PET/CT protocol .....	12
1.4.3.3    MRI acquisition for lymphoma .....	13
1.5    DETAILS OF SOFTWARE USED IN THE FRAMEWORK DEVELOPMENT .....	13
1.6    THESIS OUTLINE .....	14
<b>CHAPTER 2: FUNDAMENTALS OF MAGNETIC RESONANCE IMAGING AND DIFFUSION-WEIGHTED IMAGING IN CANCER MANAGEMENT</b> .....	<b>17</b>
2.1    GLIMPSE OF NUCLEAR MAGNETIC RESONANCE (NMR) PHYSICS: PRINCIPLES OF NUCLEAR MAGNETISM, RESONANCE, AND NUCLEAR SPIN RELAXATION .....	18
2.2    MR IMAGE RECONSTRUCTION .....	22
2.3    PULSE SEQUENCES AND ACQUISITION TECHNIQUES .....	25
2.3.1 <i>Spin echo</i> .....	25
2.3.2 <i>Gradient echo</i> .....	26
2.4    DIFFUSION-WEIGHTED IMAGING .....	27
2.4.1 <i>Pulsed Gradient Spin Echo</i> .....	29
2.4.2 <i>Quantification of physiological parameters using Diffusion models</i> .....	30
2.4.2.1    Gaussian diffusion models .....	30

2.4.2.2	Non-Gaussian diffusion models .....	34
2.4.2.3	BE model with parametric reconstruction methods .....	39
2.5	PROSTATE GLAND.....	41
2.5.1	<i>Anatomy and histology</i> .....	41
2.5.2	<i>Models of DWI in diagnosis and characterization of prostatic lesions</i> .....	42
2.6	PANCREAS .....	45
2.6.1	<i>Anatomy and histology</i> .....	45
2.6.2	<i>Models of DWI in diagnosis and characterization of pancreatic lesions</i> .....	46
2.7	LYMPHATIC SYSTEM AND LYMPH NODES .....	48
2.7.1	<i>Anatomy and histology</i> .....	48
2.7.2	<i>Diagnosis of lymphoma using DWI, CT and FDG-PET/CT imaging</i> .....	50
2.8	RADIOMICS, TEXTURE ANALYSIS AND MACHINE LEARNING IN CANCER DIAGNOSTIC IMAGING.....	53
2.9	CONCLUSION .....	56

**CHAPTER 3: IVIM-DKI PARAMETRIC MAP RECONSTRUCTION USING 3D TOTAL VARIATION MINIMIZATION.....57**

3.1	INTRODUCTION .....	58
3.2	MATERIALS AND METHODS.....	59
3.2.1	<i>Proposed IVIM-DKI image analysis</i> .....	59
3.2.2	<i>Evaluation of standard and IDTV model performance using digital phantom</i>	61
3.2.3	<i>Error estimation</i> .....	62
3.2.4	<i>Statistical tests</i> .....	63
3.3	RESULTS.....	63
3.3.1	<i>Qualitative assessment of IVIM-DKI parameters estimated using the standard and IDTV models</i> .....	63
3.3.2	<i>Quantitative assessment of IVIM-DKI parameters estimated using standard and IDTV models</i> .....	65
3.4	DISCUSSION.....	67
3.5	CONCLUSION .....	69

**CHAPTER 4: COMPARISON BETWEEN NOVEL IVIM-DKI ANALYSIS AT 1.5T AND 3T MRI FOR PROSTATE CANCER AND BENIGN PROSTATIC HYPERPLASIA .....71**

4.1	INTRODUCTION .....	71
4.2	MATERIALS AND METHODS .....	73
4.2.1	<i>Patients and Clinical Data Acquisition</i> .....	73
4.2.2	<i>Quantitative analysis of DWI</i> .....	73
4.2.3	<i>Region of Interest localization</i> .....	74
4.2.4	<i>Statistical tests</i> .....	75
4.3	RESULTS.....	76
4.3.1	<i>Tumor size</i> .....	76
4.3.2	<i>Within-scanner performance comparison between standard and IDTV model for IVIM-DKI parameter estimation</i> .....	76
4.3.3	<i>Performance comparison between the standard model at 3T and the IDTV model at 1.5T for IVIM-DKI parameter estimation</i> .....	80

4.3.4	<i>Qualitative and quantitative comparison of parameter estimation at 1.5T &amp; 3T</i>	81
4.3.5	<i>Diagnostic performance of IVIM-DKI parameters using the IDTV model at 1.5T and standard model at 3T in PCa detection</i>	84
4.4	DISCUSSION	85
4.5	CONCLUSIONS	89
<b>CHAPTER 5: PANCREATIC MASS CHARACTERIZATION USING IVIM-DKI MR IMAGING AND MULTI-PARAMETRIC TEXTURE ANALYSIS</b>		<b>91</b>
5.1	INTRODUCTION	92
5.2	NON-INVASIVE CHARACTERIZATION OF PANCREATIC MASSES USING IVIM-DKI WITH IDTV MODEL	93
5.2.1	<i>Introduction</i>	93
5.2.2	<i>Materials and Methods</i>	93
5.2.2.1	Patients and Clinical Data Acquisition	93
5.2.2.2	Quantitative analysis of DWI	94
5.2.2.3	Region of interest localization	94
5.2.2.4	Statistical tests	95
5.2.3	<i>Results</i>	96
5.2.3.1	Tumor volume	96
5.2.3.2	Model performance in pancreatic masses	96
5.2.3.3	Quantitative comparison between subtypes of pancreatic masses	102
5.2.3.4	Differential diagnosis of pancreatic masses using ROC analysis	104
5.3	MACHINE LEARNING-BASED TEXTURE ANALYSIS OF IVIM-DKI FOR CLASSIFICATION OF PANCREATIC MASSES	105
5.3.1	<i>Introduction</i>	105
5.3.2	<i>Materials and Methods</i>	105
5.3.2.1	Patients and Clinical Data Acquisition	105
5.3.2.2	Quantitative analysis of DWI	105
5.3.2.3	Region of interest localization	105
5.3.2.4	Texture feature calculation and machine learning-based classification	106
5.3.2.5	Statistical tests	107
5.3.3	<i>Results</i>	107
5.3.3.1	Multi-parametric texture analysis with machine learning-based classification of pNET vs. non-pNET	108
5.3.3.2	Multi-parametric texture analysis with machine learning-based classification of PDAC vs. non-PDAC	108
5.4	DISCUSSION	113
5.5	CONCLUSION	117
<b>CHAPTER 6: EFFECT OF COMBINATIONS AND NUMBER OF B-VALUE IN IVIM AND IVIM-DKI ANALYSIS WITH PARAMETRIC RECONSTRUCTION METHOD</b>		<b>119</b>
6.1	INTRODUCTION	120
6.2	EFFECT OF COMBINATIONS AND NUMBER OF B-VALUE ON IVIM ANALYSIS WITH CONSTRAINED RECONSTRUCTION METHOD IN PROSTATE CANCER	120

6.2.1	<i>Introduction</i> .....	120
6.2.2	<i>Materials and methods</i> .....	121
6.2.2.1	Simulations .....	121
6.2.2.2	Patients and Clinical Data acquisition .....	122
6.2.2.3	Quantitative Analysis of DWI.....	123
6.2.2.4	Error calculations .....	123
6.2.2.5	Region of Interest localization .....	123
6.2.2.6	Statistical tests .....	124
6.2.3	<i>Results</i> .....	124
6.2.3.1	Simulations.....	124
6.2.3.2	Clinical data.....	126
6.3	IMPACT OF B-VALUE COMBINATIONS ON IVIM-DKI ANALYSIS WITH CONSTRAINED RECONSTRUCTION METHOD IN PANCREATIC ADENOCARCINOMA .....	131
6.3.1	<i>Introduction</i> .....	131
6.3.2	<i>Materials and Methods</i> .....	131
6.3.2.1	Simulations.....	131
6.3.2.2	Patient information and Clinical data acquisition .....	133
6.3.2.3	Quantitative analysis of DWI.....	133
6.3.2.4	Error calculations .....	133
6.3.2.5	Region of Interest localization .....	134
6.3.2.6	Statistical tests .....	134
6.3.3	<i>Results</i> .....	135
6.3.3.1	Comparison between parameters estimated at different number and combination of b-values in digital phantoms.....	135
6.3.3.2	Comparison between parameters estimated at different number and combination of b-values in PDAC and normal parenchyma .....	140
6.3.3.3	Agreement between parameters estimated at different numbers and combinations of b-values in PDAC and normal parenchyma.....	145
6.4	DISCUSSION.....	147
6.5	CONCLUSIONS .....	151

**CHAPTER 7: IVIM-DKI FOR LYMPH NODE EVALUATION AND CHARACTERIZATION IN LYMPHOMA COMPARED TO FDG-PET/CT ..... 153**

7.1	INTRODUCTION .....	154
7.2	MATERIALS AND METHODS.....	155
7.2.1	<i>Patients and Clinical Data Acquisition</i> .....	155
7.2.2	<i>Quantitative analysis of IVIM-DKI and PET imaging</i> .....	156
7.2.3	<i>Multimodal 3D-registration between PET and IVIM-DKI volume</i> .....	156
7.2.4	<i>Localization of ROI</i> .....	157
7.2.5	<i>Statistical analysis</i> .....	158
7.3	RESULTS.....	159
7.3.1	<i>Qualitative assessment of malignant lymph nodes in lymphoma using IVIM-DKI and PET imaging</i> .....	159
7.3.2	<i>Quantitative characterization of benign and malignant lymph nodes in lymphoma using IVIM-DKI and ROC analysis</i> .....	160

7.3.3	<i>Quantitative characterization of malignant lymphoma subtypes using IVIM-DKI, PET imaging and ROC analysis</i> .....	163
7.3.4	<i>Correlation between ADC and IVIM-DKI parameters with PET parameters in lymphoma</i> .....	164
7.4	DISCUSSIONS .....	165
7.5	CONCLUSION .....	169
<b>CHAPTER 8: CONCLUSIONS AND FUTURE DIRECTIONS</b> .....		<b>171</b>
8.1	THESIS CHAPTER SUMMARY .....	171
8.2	MAJOR CONTRIBUTION .....	173
8.3	FUTURE ASPECTS .....	174
<b>APPENDIX</b> .....		<b>177</b>
	APPENDIX I: IVIM-DKI ANALYSIS WITH TV METHOD TOOLBOX SPECIFICATIONS .....	177
	APPENDIX II: SIMULATION RESULTS FOR EFFECT OF COMBINATIONS AND NUMBER OF B-VALUE IN IVIM-DKI ANALYSIS WITH CONSTRAINED RECONSTRUCTION METHOD.....	179
<b>REFERENCES</b> .....		<b>189</b>
<b>AUTHOR'S PUBLICATIONS</b> .....		<b>203</b>
	JOURNAL PUBLICATIONS.....	203
	CONFERENCE PROCEEDINGS .....	203
<b>ABOUT THE AUTHOR</b> .....		<b>205</b>

# List of Figures

Figure 1.1: Thesis summary of implementation of IVIM-DKI model with total variation minimization method in various cancer applications.

Figure 1.2: Clinical data information of patients suffering from prostate cancer, (a) Gleason grade and (b) Prostate Imaging Reporting & Data System score.

Figure 1.3: Clinical data information of patients suffering from pancreatic adenocarcinoma (PDAC), pancreatic neuroendocrine tumor (pNET), mass-forming chronic pancreatitis (MFCP), and solid papillary epithelial neoplasm (SPEN).

Figure 1.4: Clinical data information of patients suffering from Hodgkin lymphoma (HL) and non-Hodgkin lymphoma (NHL).

Figure 2.1: The alignment of the spinning nucleus with or against an external magnetic field depends on where it is positioned. A lesser energy is seen in parallelly aligned nuclei compared to anti parallel oriented nuclei. Inspired from ref: [21].

Figure 2.2: MRI principle where (a) the protons' spins are randomly oriented, but when exposed to a strong magnetic field  $B_0$  ( $Z$ ) that is always on, (b) they become aligned. (c) The spins from the alignment are flipped using a radiofrequency pulse. (d) When the radiofrequency is removed, the spins return to equilibrium. Inspired from ref: [21].

Figure 2.3: Simulated T1 and T2 relaxation curves in 3T MRI, T1 and T2 relaxation curves were simulated for a prostate cancer.

Figure 2.4: When the transverse magnetization is reduced to zero, the RF coil signal oscillates sinusoidal, returning the net magnetization to equilibrium. This is a free induction decay simulation (FID).

Figure 2.5: A variable to denote spatial frequency is  $k$ . Therefore, 2D-space in which Fourier transform operation is defined, is called  $k$ -space, with its space variables,  $k_x$  and  $k_y$ . Inspired from ref: [21]

Figure 2.6: Sequence of spin echo pulses. Inspired from ref. [23].

Figure 2.7: For spoiled gradient echo (SPGR) sequence, the gradient spoiling crushing gradients and varying RF-carrier phase are represented in blue. Inspired from ref. [23]

Figure 2.8: The flow of water molecules throughout tissues of varied cellularity, (a) free diffusion, (b) hindered diffusion, (c) restricted diffusion, and (d) Diffusion signal attenuation based on diffusion based on in vivo environment.

Figure 2.9: Pulse sequence for diffusion-weighted imaging. Inspired from ref. [23].

Figure 2.10: Illustration of a standard ADC fitting procedure using a monoexponential model with three diffusion-weighted images at  $b=0$  s/mm<sup>2</sup>,  $b=500$  s/mm<sup>2</sup>, and  $b=1000$ s/mm<sup>2</sup>.

Figure 2.11: Illustration of a standard IVIM fitting procedure using a biexponential model with 10 diffusion-weighted images at  $b=0$  - 1000s/mm<sup>2</sup>.

Figure 2.12: Illustration of a standard IVIM fitting procedure using an IVIM-DKI model with 14 diffusion-weighted images at  $b=0$  - 2000s/mm<sup>2</sup>.

Figure 2.13: Relative signal intensity variations for DWI signal using the IVIM-DKI model with varying parameters.

Figure 2.14: A representative image of patient with pancreatic ductal adenocarcinomas (PDAC); where qualitative comparison between parameter maps (D, D\*, and f maps) calculated using biexponential (BE) and biexponential with TV (BE+TV) models.

Figure 2.15: Zones of prostate gland. Inspired from ref [64]

Figure 2.16: Multi-parametric comparison of prostate cancer appearance on (a) T2-weighted (hypointense), (b) high b-value DWI (hyperintense), and (c) ADC (hypointense) pointed by arrow (blue).

Figure 2.17: Anatomy of pancreas. Inspired from ref [75]

Figure 2.18: Neuroendocrine tumor is (a) hyperintense in T2-weighted, high b-value DWI (b), hyperintense mass with ADC showing restricted diffusion coefficient (c).

Figure 2.19: Schematics of lymphatic system across the body. (a) A network of lymphatic vessels and (b) Afferent lymph. Inspired from ref [83]

Figure 2.20: A representative images showing maximum-intensity projection (MIP) of PET images and CT images used for primary tumor localization in lymphoma. (a-b) MIP and CT image of a 31 year old male patient with Diffuse large B-cell lymphoma (DLBCL) at stage IV and primary tumor being in anterior chest wall (orange arrow). (c-d) MIP and CT image of a 26 year old male patient with follicular lymphoma at stage IV and primary tumor being in rectosigmoid bowel loop (orange arrow). (e-f) MIP and CT image of a 28 year old female patient with Classical Hodgkin lymphoma, Nodular Sclerosis type at stage IV and primary tumor being in left cervical node (orange arrow).

Figure 2.21: Workflow for the classification of benign and malignant prostate lesions using texture analysis and machine learning.

Figure 3.1: Flow chart describing the total variation minimization of IVIM-DKI parametric maps using IDTV model.

Figure 3.2: Representation of digital phantom of the kurtosis (k) map used in Simulation IV.

Figure 3.3: IVIM-DKI parametric maps (a) D map, (b) D\* map, (c) f map, and (d) k map estimated by standard and IDTV models using 13 b-value combination. 1st row represents the reference map of each parameter. Succeeding rows represent the results at different SNRs: 15, 20, 25, 30, 40, and 50.

Figure 3.4: Error estimation of D, D\*, f, and k using (a) RRMSE, (b) RB, and (c) SSIM of standard and IDTV models

Figure 3.5: Parameter-wise comparison between reference and estimated parameters (a) D, (b) D\*, (c) f, and (d) k maps estimated using standard and IDTV models using different SNR 15, 20, 25, 30, 40, and 50

Figure 4.1: ROI localization of tumor (black circle), healthy PZ (red circle) and BPH (blue circle) using IVIM-DKI at (a)  $b=2000 \text{ s/mm}^2$ , (b) ADC map, and (c)  $b=0 \text{ s/mm}^2$  for one representative patient (65 years old) having prostate cancer with Gleason score of 7 (4+3) acquired at 1.5T.

Figure 4.2: Box-whisker plot showing comparison between IDTV model (yellow), and standard model (red) estimated parameters (b) D, (c) D\*, (d) f and (e) k for healthy PZ, tumor, and BPH ROI using IVIM-DKI image and (a) ADC for 1.5T and 3T data.

Figure 4.3: Box-whisker plot showing CVcomb for (a) healthy PZ, (b) tumor, and (c) BPH obtained from standard (red) and IDTV (yellow) model using IVIM-DKI image from 1.5T and 3T.

Figure 4.4: Fitting of mean DWI signal at b-values 0 to  $2000 \text{ s/mm}^2$  using standard and IDTV models at (a-c) 1.5T and (d-f) 3T in (a,d) healthy PZ, (b,e) tumor, and (c,f) BPH ROI.

Figure 4.5: Coefficient of Variation (CV (%)) of D, D\*, f, and k in healthy PZ, tumor and BPH obtained from IVIM-DKI parameter estimation using IDTV model at 1.5T and standard model at 3T.

Figure 4.6 IVIM-DKI parameter maps of representative patients ((a) male, 61 years and (b) male, 60 years) with PIRADS 4 lesion encircled by black circle. Tumor appearing hypointense on D and f map and hyperintense on D\* and k map obtained from standard and IDTV models at (a) 1.5T and (b) 3T.

Figure 4.7 Box-whisker plot of ADC and IVIM-DKI parameters (D, D\*, f, k) in healthy PZ, tumor and BPH obtained using (a) IDTV model at 1.5T and (b) standard model at 3T.

Figure 5.1: Representation of ROI localization in 27 years old female patient with pancreatic mass (blue line highlighting the region of SPEN mass) as viewed on IVIM-DKI image at  $b = 0 \text{ s/mm}^2$ .

Figure 5.2: Coefficient of Variation (CV (%)) of D, D\*, f, and k in healthy PZ, tumor and BPH obtained from IVIM-DKI parameter estimation using IDTV model at 1.5T

Figure 5.3: A representative of a 72 year old male patient with PDAC; demonstrating qualitative comparison between parameter maps estimated using standard (d, f, h, and j) and IDTV model (e, g, i, k). The encircled zone (white) represents the tumor-affected region and (c) curve fitting the mean of IVIM-DKI signal decay for tumor region.

Figure 5.4: A representative image of a 28 year old female patient with solid papillary epithelial neoplasm (SPEN); where qualitative comparison between parameter maps (D, D\*, f and k maps) estimated using standard and IDTV model. The encircled zone (white) represents the tumor-affected region.

Figure 5.5: A representative image of a 63 year old male patient with mass-forming chronic pancreatitis (MFCP); where qualitative comparison between parameter maps (D, D\*, f and k maps) estimated using standard and IDTV models. The encircled zone (white) represents the tumor-affected region.

Figure 5.6: A representative image of a 43 year old male patient with pancreatic neuroendocrine tumor (pNET); where qualitative comparison between parameter maps (D, D\*, f and k maps) estimated using standard and IDTV models. The encircled zone (white) represents the tumor-affected region.

Figure 5.7: Box-whisker's plot shows the distribution of ADC and IVIM-DKI parameters estimated using IDTV model in PDAC, MFCP, SPEN, and pNET.

Figure 5.8: Flow chart representing machine learning-based classification of pancreatic masses using texture features of ADC and IVIM-DKI parameters.

Figure 6.1: Two-dimensional digital phantom with varying parameter map of f (matrix-size:64x64), consisting of concentric circles; f varied from 0.03 in center to 0.43 periphery, as indicated.

Figure 6.2: Prostate cancer PIRADS 4 lesion in left transitional zone (Tz) appearing hyperintense on high b-value IVIM image and hypointense on corresponding ADC map.

Example ROIs, corresponding to tumor (black), and healthy tissue drawn on transition zone (blue) and peripheral zone (red).

Figure 6.3: Estimated parameter maps using various combinations of b-values using BE and BE+TV for Simulation 1, 2 and 3. D, D\* and f maps are the reference maps.

Figure 6.4: Bar plots showing RRMSE of simulated D map, D\* map and f map generated using different combinations of b-values.

Figure 6.5: A representative patient with prostate cancer PIRADS 4 lesion (Black circle) in left transitional zone (Tz) and parametric maps (D, D\*, f) using BE+TV model using 4 b-values, 8 b-values, 13 b-values.

Figure 6.6: Correlation and Bland-Altman plots of parameter generated from BE+TV using 8 b-values and 13 b-values in Tumor, transition zone and peripheral zone.

Figure 6.7: Coefficient of variation (CV) of IVIM parameters, (a) D map, (b) D\* map and (c) f map generated from BE (blue) and BE+TV (orange) using 4, 8 and 13 b-values.

Figure 6.8: Parametric maps (a) D map, (b) D\* map, (c) f map, and (d) k map estimated by standard and IDTV model using different b-value combination sets at SNR 20. 1<sup>st</sup> row represents reference map of each parameter. Succeeding rows represent the results of different b-value combinations.

Figure 6.9: Parameter-wise RRMSE of (a) D, (b) D\*, (c) f, and (d) k estimated using standard and IDTV model at different b-values combinations and SNR 20.

Figure 6.10: Mean SSIM index of (a) D, (b) D\*, (c) f, and (d) k estimated using standard and IDTV model at different b-values combinations and all SNR.

Figure 6.11: Total parameter error in percentage of standard and IDTV model using different b-values combinations at SNR (a) 15, (b) 20, (c) 30, and (d) 50.

Figure 6.12: Parameter-wise comparison between reference and estimated parameters by using standard and IDTV models and different b-values combinations at (a) SNR20, (b) SNR30, and (c) SNR50.

Figure 6.13: Coefficient of variation (CV) of (a,e) D, (b,f) D\*, (c,g) f, and (d,h) k estimated using standard (blue) and IDTV (red) model and 6,8,10, and 13 b-values in (a-d) tumor and (e-h) normal parenchyma ROI.

Figure 6.14: Mean differences between (a,f,k,p) ADC, (b,g,l,q) D, (c,h,m,r) D\*, (d,i,n,s) f, and (e,j,o,t) k parameters estimated using standard and IDTV model and (a-e) 6 b-values, (f-j) 8 b-values, (k-o) 10 b-values, and (p-t) 13 b-values in tumor (blue) and

normal parenchyma (red) ROI. (l) D, (m) D\*, and (d,i,n,s) f parameters estimated using IDTV model.

Figure 7.1: Schematics of multimodality registration between IVIM-DKI and PET images.

Figure 7.2: Representative images of (a-d) of 32 year old male patient suffering from Diffuse large B-cell lymphoma (DLBCL) at stage IV, (e-h) 45 year old male patient suffering from follicular lymphoma at stage III, and (i-l) 28 year old female patient suffering from classical Hodgkin lymphoma (HL), (a,e,i) IVIM-DKI at  $2000 \text{ s/mm}^2$ , (b,f,j) SUV map, (c,g,k) STIR image, and (d,h,l) images showing fusion between SUV and STIR images.

Figure 7.3: Representative images of 32 year old male patient suffering from Diffuse large B-cell lymphoma (DLBCL) at stage IV showing (a) IVIM-DKI at  $2000 \text{ s/mm}^2$ , (b) SUV map, (c) STIR images, and (d) ADC map. (e) D, (f, j) D\*, (g, k) f, and (h, l) k maps.

Figure 7.4: Boxplot showing comparison between ADC and IVIM-DKI parameters quantified from IDTV model in benign and malignant lymph node.

Figure 7.5: Boxplot showing comparison between ADC and SUV parameters and IVIM-DKI parameters quantified from IDTV model in Hodgkin lymphoma (HL) and non-Hodgkin lymphoma (NHL).

Figure 7.6: Correlation plots showing correlation between ADC and IVIM-DKI parameters,  $\text{SUV}_{\max}$  with  $\text{SUV}_{\text{mean}}$  of the patients with lymphoma.

Appendix I figure 1: Overview of open access IVIM-DKI with TV analysis toolbox (v1.4) implementing IDTV model. (a) Input Data and Analysis, (b) Calculate ROI statistics and Save the results in excel format, and (c) Output tab

Appendix II figure 1: Parametric maps (a) D map, (b) D\* map, (c) f map, and (d) k map estimated by standard and IDTV model using b-value combination3 and 4 sets at SNR 20. 1<sup>st</sup> row represents reference map of each parameter. Succeeding rows represent the results of different b-value combinations.

Appendix II figure 2: Parametric maps D map estimated by standard and IDTV model using (a-d) b-value combination1 and 2 and (e-h) b-value combination3 and 4 at SNR (a,e) 15, (b,f) 25, (c,g) 30, (d,h) 50. 1<sup>st</sup> row represents reference map of each parameter. Succeeding rows represent the results of different b-value combinations.

Appendix II figure 3: Parametric maps  $D^*$  map estimated by standard and IDTV model using (a-d) b-value combination1 and 2 and (e-h) b-value combination3 and 4 at SNR (a,e) 15, (b,f) 25, (c,g) 30, (d,h) 50. 1<sup>st</sup> row represents reference map of each parameter. Succeeding rows represent the results of different b-value combinations.

Appendix II figure 4: Parametric maps  $f$  map estimated by standard and IDTV model using (a-d) b-value combination1 and 2 and (e-h) b-value combination3 and 4 at SNR (a,e) 15, (b,f) 25, (c,g) 30, (d,h) 50. 1<sup>st</sup> row represents reference map of each parameter. Succeeding rows represent the results of different b-value combinations.

Appendix II figure 5: Parametric maps  $k$  map estimated by standard and IDTV model using (a-d) b-value combination1 and 2 and (e-h) b-value combination3 and 4 at SNR (a,e) 15, (b,f) 25, (c,g) 30, (d,h) 50. 1<sup>st</sup> row represents reference map of each parameter. Succeeding rows represent the results of different b-value combinations.

Appendix II figure 6: Parameter-wise RRMSE (a)  $D$ , (b)  $D^*$ , (c)  $f$ , and (d)  $k$  estimated using standard and IDTV model at different b-values combinations and SNR 15.

Appendix II figure 7: Parameter-wise RRMSE (a)  $D$ , (b)  $D^*$ , (c)  $f$ , and (d)  $k$  estimated using standard and IDTV model at different b-values combinations and SNR 25.

Appendix II figure 8: Parameter-wise RRMSE (a)  $D$ , (b)  $D^*$ , (c)  $f$ , and (d)  $k$  estimated using standard and IDTV model at different b-values combinations and SNR 30.

Appendix II figure 9: Parameter-wise RRMSE (a)  $D$ , (b)  $D^*$ , (c)  $f$ , and (d)  $k$  estimated using standard and IDTV model at different b-values combinations and SNR 40.

Appendix II figure 10: Parameter-wise RRMSE (a)  $D$ , (b)  $D^*$ , (c)  $f$ , and (d)  $k$  estimated using standard and IDTV model at different b-values combinations and SNR 50.

Appendix II figure 11: Error while estimating (a)  $D$ , (b)  $D^*$ , (c)  $f$ , and (d)  $k$  parameters using IDTV model at different SNRs and b-values combination1. IDTV model error linearly decreases with increase in SNR and numbers of b-values.

Appendix II figure 12: Parameter-wise comparison between reference and estimated parameters by standard and IDTV models using different b-value combinations at (a) SNR15, (b) SNR25, and (c) SNR40.

## List of Tables

Table 4.1: Diagnostic performance of IVIM-DKI parameters obtained from IDTV model at 1.5T and standard model at 3T in PCa detection

Table 5.1: Comparison between mean of DWI and IVIM-DKI parameters quantified using new approach in pancreatic masses

Table 5.2: P-values obtained from Tukey-Kramer Multiple comparison test between ADC and IVIM-DKI parameters values estimated using IDTV model in pancreatic masses

Table 5.3: ROC analysis of characterizing pancreatic masses using ADC and IVIM-DKI parameters quantified using IDTV model

Table 5.4: Texture features extracted from Global, GLCM, GLRLM, and NGTDM

Table 5.5: Machine learning-based classification performance of all texture features (30 features) and top ten features selected from ADC, combined and individual IVIM-DKI parameters, and ADC combined with IVIM-DKI parameters to classify pNET from non-pNET pancreatic masses.

Table 6.1: Different number and combinations of b-values used in simulations

Table 6.2: ADC, D, D\*, and f (mean±SD) values for Tumor, transition zone and peripheral zone in prostate using different combinations of b-values.

Table 6.3: Different numbers and combinations of b-values were used to estimate IVIM-DKI parameters. 6 to 10 b-values and 4 different combinations for each set were used and 13b-values1 combination was used as standard b-values combination.

Table 6.4: Mean±SD of ADC, D, D\*, F, K parameters estimated using standard and IDTV model and different combinations of b-values such as 6,8,10,13b-values in tumor and normal parenchyma ROI

Table 7.1: Quantitative comparison between mean of ADC and IVIM-DKI parameters in benign and malignant lymph node.

Table 7.2: Characterizing of ADC and IVIM-DKI parameters estimated using IDTV model in benign and malignant lymph nodes in the lymphoma using ROC analysis

Table 7.3: Quantitative comparison between mean of ADC, IVIM-DKI, and SUV parameters in NHL and HL.

Table 7.4: Characterizing of ADC, PET and IVIM-DKI parameters estimated using IDTV model in HL and NHL using ROC analysis

# Glossary

<b>DWI</b>	Diffusion-weighted imaging
<b>PCa</b>	Prostate cancer
<b>TRUS</b>	Transrectal ultrasonography
<b>MRI</b>	Magnetic resonance imaging
<b>CT</b>	Computed tomography
<b>FDG-PET/CT</b>	Fluorodeoxyglucose-Positron emission tomography/CT
<b>ADC</b>	Apparent diffusion coefficient
<b>IVIM</b>	Intra-voxel incoherent motion
<b>ME</b>	Monoexponential
<b>BE</b>	Bisexponential
<b>DKI</b>	Diffusion kurtosis imaging
<b>IVIM-DKI</b>	IVIM combined with DKI
<b>SNR</b>	Signal-to-noise ratio
<b>2D</b>	Two-dimensional
<b>TV</b>	Total variation
<b>BE+TV</b>	BE model with TV method
<b>3D</b>	Three-dimensional
<b>BPH</b>	Benign prostatic hyperplasia
<b>AIIMS</b>	All India Institute of Medical Sciences
<b>EPI</b>	Echo planar imaging
<b>PI-RADS</b>	Prostate Imaging Reporting & Data System
<b>SE</b>	Spin Echo
<b>TR</b>	Repetition time
<b>TE</b>	Time of echo
<b>SSFP</b>	Steady-state free precession
<b>FS</b>	Fat-suppressed
<b>TSE</b>	Turbo SE
<b>MRCP</b>	Magnetic resonance cholangiopancreatography
<b>HASTE</b>	Half-Fourier Acquisition Single-shot Turbo spin Echo imaging
<b>SE-EPI</b>	Spin echo-echo planar imaging

<b>SPIR</b>	Spectral presaturation with inversion recovery
<b>MFCP</b>	Mass-forming chronic pancreatitis
<b>SPEN</b>	Solid papillary epithelial neoplasm
<b>PDAC</b>	Pancreatic adenocarcinoma
<b>pNET</b>	Pancreatic neuroendocrine tumor
<b>FNAC</b>	Fine-needle aspiration cytology
<b>NMR</b>	Nuclear magnetic resonance
<b>RF</b>	Radiofrequency
<b>1D</b>	One-dimensional
<b>GRE</b>	Gradient echo
<b>FSE</b>	Fast spin echo
<b>PGSE</b>	Pulsed gradient spin echo
<b>ADC</b>	Apparent diffusion coefficient
<b>DDC</b>	Distributed diffusion coefficient
<b>MCP</b>	Microcirculatory perfusion
<b>D</b>	Diffusion coefficient
<b>D*</b>	Pseudo-diffusion coefficient
<b>f</b>	Perfusion fraction
<b>k</b>	Kurtosis
<b>CV</b>	Coefficient of variation
<b>wCV</b>	Within-subject CV
<b>RRMSE</b>	Relative root means square error
<b>RB</b>	Relative bias
<b>SD</b>	Standard deviation
<b>Pz</b>	Peripheral zone
<b>Tz</b>	Transition zone
<b>AFS</b>	Anterior fibromuscular stroma
<b>DCE</b>	Dynamic contrast-enhanced
<b>mp-MRI</b>	Multiparametric-MRI
<b>bp-MRI</b>	Biparametric-MRI
<b>AUC</b>	Area under curve
<b>ROC</b>	Receiver-operating-characteristics
<b>GS</b>	Gleason score

<b>AIP</b>	Autoimmune pancreatitis
<b>HL</b>	Hodgkin lymphoma
<b>NHL</b>	Non-Hodgkin lymphoma
<b>MIP</b>	Maximum intensity projection
<b>DLBCL</b>	Diffuse large B-cell lymphoma
<b>MALT</b>	Mucosa-associated lymphoid tissue
<b>CLL/SLL</b>	Chronic lymphocytic leukemia/small lymphocytic lymphoma
<b>HNSCC</b>	Head and neck squamous cell carcinomas
<b>NAC</b>	Neo-adjuvant chemotherapy
<b>NPC</b>	Nasopharyngeal carcinoma
<b>MRMR</b>	Minimum Redundancy Maximum Relevance Algorithm
<b>LASSO</b>	Least absolute shrinkage and selection operator
<b>GLCM</b>	Gray-level co-occurrence matrix
<b>NGTDM</b>	Neighbourhood gray-tone difference matrix
<b>GLRLM</b>	Gray-level run length matrix
<b>IDTV</b>	IVIM-DKI model with TV method
<b>SSIM</b>	Structural similarity index
<b>CV<sub>comb</sub></b>	Combined CV
<b>ANOVA</b>	Analysis of variance
<b>ANN</b>	Artificial neural network
<b>LOA</b>	Limit of agreement