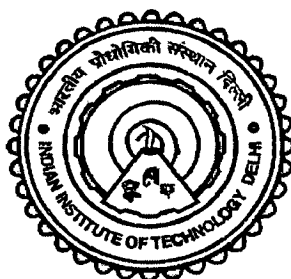


**SCREENING OF MICROORGANISMS AND  
EXPRESSION OF GENE INVOLVED IN DEGRADATION  
OF DIBENZOTHIOPHENE**

by  
**NIDHI GUPTA**  
Department of Biochemical Engineering and Biotechnology

Submitted  
In fulfillment of the requirements of the degree of  
**Doctor of Philosophy**  
to the



**Indian Institute of Technology, Delhi**  
**September 2005**

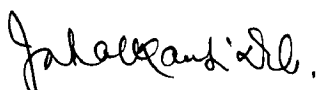
I. I. T. DELHI  
LIBRARY  
Acc. No TH-3229

I.I.T. DELHI  
PROCESSED  
LIBRARY

TH  
547.54  
GUP-5

## CERTIFICATE

*This is to certify that the thesis entitled “ Screening of microorganisms and expression of gene involved in degradation of Dibenzothiophene” being submitted by **Nidhi Gupta** to the Indian Institute of Technology, Delhi, for the award of Degree of **Doctor of Philosophy**, is a bonafide research work carried out by her under our supervision and guidance. The results presented in this thesis have not been submitted to any other University or Institute for the award of any other degree or diploma.*



**(J.K. Deb)**  
*Associate Professor*  
*Deptt. of Biochemical Engineering & Biotechnology*  
*Indian Institute of Technology*  
*New Delhi-110016*



**(P.K. Roychoudhury)**  
*Associate Professor*  
*Deptt. of Biochemical Engineering & Biotechnology*  
*Indian Institute of Technology*  
*New Delhi-110016*

## *Acknowledgment*

*This thesis marks the finalization of the PhD-study programme that I commenced December 2000. It has been an interesting, stimulating, challenging and occasionally frustrating process. Luckily, I have through out enjoyed the encouragement and support from numerous people. It is a pleasant aspect that I have now the opportunity express my gratitude to all of them.*

*First and foremost I would like to express my gratitude to my supervisors Dr. J.K. and Dr. P.K. Roychoudhury for their constant guidance and deep sense of involvement in my Ph.D work. I have enjoyed the creative way of thinking of Dr. Deb and ability to ask the right question at the right time has helped me on numerous occasions. I would like to thank Dr. Roychoudhury again for giving practical and friendly advice.*

*I am grateful to Dr. Lalit Garg, NII, New Delhi, Prof. Saroj Mishra and Prof. Subh Chand for their critical comments during development of this work on which this thesis is based.*

*I would also like to thank Dr. D.K. Adhikari (IIP) for providing various soil, water and sludge samples, carrying out GC-SCD analysis of my samples and for various scientific discussions, which we had together. Dr. Tsering Stobdan (IIP) deserves special mention for providing his insightful scientific expertise and many helpful suggestions.*

*I would also like to thank Dr. Ramesh, Department of Chemistry, IIT Delhi, for carrying out GC-MS spectral analysis of my samples.*

*I also feel responsible to express my gratitude to Mr. V.K. Ghosh for the critical analysis of bacterial strains and Mr. Mukesh Anand and Mr. Sapan Patra for their friendly manner and helping nature. Thanks to Ms. Renu Sethi, Mr. S.P. Rana, Mr. G.P. Yadav, Mr. I.A. Khan, Mr. Bhagwan Singh, Mr. Kishan, Mr. Prashant, Office staff, Sunita Dang, Mr. Rajeev, Ms. Meena Mathur, D.S. Tarzan and Ms. Neera Verma for their help.*

*Meherchandji deserves special thanks for keeping the lab clean and organized.*

*I would like to thank my labmates and colleagues Usha, Snehasis and Rupali for providing congenial environment in the lab, their undemanding help and unceasing support. They are more of friends to me. I think without their support and help my journey would have not been that memorable and pleasurable. Thanks to Shyam Shankari and Charu also for providing me their support and help.*

*I would also like to thank my friends and colleagues Ruchi, Gunjan, Rumpa, Sunita mam, Shilpa, Bhawna, Krishnan, Ashish, Ashwani for their support and help. Vibha deserves special mention. I can't express my feelings for her in words but I can definitely say that She is a true friend and I always found her standing next to me whenever I needed her. I can never forget the support, which she provided me during the initial days.*

*Finally and above all, I want to thank my parents, in laws, my husband Manish, my brother Neeraj and sister Namita. My parents always encouraged me to pursue what I enjoyed. They gave me the determination and moral support to accomplish objectives the honest (but often most difficult) way. They always provided me with the best of everything. Despite our aggressive interactions as children, my brother and sister blossomed into considerate and supportive siblings. I especially thank them for spending many hours tinkering with computers for doing corrections in my reports. The dreams of my parents would have never been fulfilled if I would have not got such supportive, loving and encouraging husband and in laws. Without their love, patience, understanding, joy, happiness and support my life would have been much poorer and this thesis would probably never have been finished.*

*At last, I want to devote this thesis to God. Thank him for giving me such loving and caring people. It would have not possible to complete this Ph. D programme without his grace.*

*I acknowledge Council of Scientific and Industrial Research (CSIR), New Delhi for providing financial support during the thesis work.*

*Nidhi Gupta*  
*Nidhi Gupta*  
*(2000 RBC 003)*

## **ABSTRACT**

Organic sulfur compounds in fossil fuels have long been one of the major sources of environmental pollution. Sulfur oxides emissions from the combustion of diesel have been shown to be responsible for acid rain, which could dissolve buildings, kill forests, and poison lakes. To avoid the production of such pollutants, sulfur must be removed from the diesel. Currently, the refining industry applies the energy intensive physico-chemical hydrodesulfurization (HDS) process in order to reduce the sulfur content. Due to its high costs and inherent inability to act upon low levels of aromatic sulfur compounds, biodesulfurization of hydrocarbon streams might represent an attractive complementary method as it can act on low concentration of aromatic sulfur containing component. Bacteria require relatively mild process conditions (pressure and temperature) and bacterial enzymes are very selective in converting target molecules. Moreover, biodesulfurized diesel has better lubricating property as compared to hydrodesulfurized diesel and this process does not result in the production of green house gas. Further, it has been reported that biodesulfurization can be competitive with hydrodesulfurization technology only with the concomitant production of sulfones that can be further used as surfactants and hydrotopes.

Dibenzothiophene (DBT) is considered to be model compound that is difficult to desulfurize using hydrodesulfurization technology. Keeping this in mind, we collected various soil, water and oil sludge samples for the screening of microorganisms, which could utilize DBT as the sulfur source. We were able to screen two different microorganisms, which utilized this compound efficiently. They have the ability to degrade 4,6 Dimethyl-DBT (DM-DBT) in addition to DBT, the former being present in diesel and is difficult to desulfurize than the latter. The two bacteria identified are *Citrobacter freundii* and *Rhodococcus* sp.

It is for the first time that *Citrobacter* is reported to be responsible for the degradation of DBT and DM-DBT. We have identified the intermediate of DBT degradation but its pathway is yet to be worked out. The desulfurization activity of diesel using *Citrobacter* is comparable to those of other bacteria.

*Rhodococcus* was found to desulfurize DBT following 4S pathway and contains all the three genes. The first step of 4S pathway is catalyzed by DBT monooxygenase and is able to convert DBT to DBT sulfone. Removal of DBT sulfone not only leads to desulfurization of diesel but also the production of valuable sulfones. With this objective in view

cloning and expression of *dsz C* was done. DBT monooxygenase was further purified and its activity studied.

A single step purification procedure resulted in 39 fold purification of the DBT monooxygenase. The basic kinetic parameter  $K_m$  and  $V_{max}$  of 0.012 mM and .002 mM/min respectively suggest that this enzyme has got high affinity for DBT and can even be used to remove sulfur from low sulfur diesel.

Thus both these microorganisms have the potential to be developed into an efficient biocatalyst for biodesulfurization.

# CONTENTS

	<b>Page No.</b>	
<b>Chapter 1</b>	<b>Introduction And Objectives</b>	<b>1-11</b>
1.1	Hydrodesulfurization	
1.2	Biodesulfurization	
1.2.1	Molecular Biology of 4S pathway	
1.3	Present status of Biodesulfurization	
1.4	Objectives	
<b>Chapter 2</b>	<b>Review of Literature</b>	<b>12-47</b>
2.1	Biodesulfurization	
2.2	Bacterial metabolism of Dibenzothiophene and its analogue	
2.2.1	Reductive desulfurization	
2.2.2	Oxidative degradation	
2.2.2.1	Attack on the benzene homocyclic ring not influenced by the electronegativity of the heteroatom attached	
2.2.2.2	Attack on dibenzothiophene influenced due to electronegative heteroatom	
2.3	Molecular biology of Kodama pathway	
2.4	Molecular biology of 4S pathway	
2.5	Enzymology of 4S pathway	
2.6	Biodesulfurization of fuels	
2.7	Commercialization of BDS	
<b>Chapter 3</b>	<b>Material and Methods</b>	<b>48-75</b>
3.1	Strains, plasmids and primers	
3.2	Screening and isolation of microorganisms for sulfur specific desulfurization of DBT	
3.3	Screening of soil isolates producing 2-HBP as the end product using dibenzothiophene as the sulfur source	
3.3.1	Gibb's assay	
3.3.2	Gas chromatography and High performance liquid chromatography	
3.4	Gas chromatography - Pulse Flame Photometric detector	
3.5	Gas chromatography - Mass spectrometry ( GC-MS)	
3.6	Isolation of chromosomal DNA from <i>Rhodococcus</i>	
3.7	Isolation of plasmid from <i>E.coli</i>	
3.7.1	Large scale isolation	
3.7.2	Small scale isolation	
3.8	Agarose gel electrophoresis	
3.9	DNA elution from agarose gel	
3.9.1	Freeze Squeeze Method	
3.9.2	AuPreP Gel extraction Method	
3.10	DNA clean up by Wizard system	
3.11	Colony lysis	
3.12	Identification of bacterial isolate	
3.12.1	Identification using api 20 E	
3.12.2	Random Amplification of Polymorphic DNA	

