

POLYPHENOLS AND THEIR EFFECT ON THE AGGREGATION OF ALS-LINKED Cu-Zn SUPEROXIDE DISMUTASE (SOD1)

NIDHI KAUR BHATIA



**DEPARTMENT OF CHEMISTRY
INDIAN INSTITUTE OF TECHNOLOGY DELHI**

MAY 2018

©Indian Institute of Technology Delhi (IITD), New Delhi, 2018

POLYPHENOLS AND THEIR EFFECT ON THE AGGREGATION OF ALS-LINKED Cu-Zn SUPEROXIDE DISMUTASE (SOD1)

by

NIDHI KAUR BHATIA
Department of Chemistry

Submitted

in fulfillment of the requirements of the degree of Doctor of Philosophy

to the



INDIAN INSTITUTE OF TECHNOLOGY DELHI

MAY 2018

CERTIFICATE

This is to certify that the thesis entitled ‘**Polyphenols and their effect on the aggregation of ALS-linked Cu-Zn superoxide dismutase (SOD1)**’ being submitted by Ms. **Nidhi Kaur Bhatia** to the Indian Institute of Technology Delhi for the award of the degree of **Doctor of Philosophy** in chemistry is a record of a bona fide research work carried out by her. Ms. Nidhi Kaur Bhatia has worked under my guidance and supervision and has fulfilled the requirements for the submission of this thesis, which to my knowledge has reached the requisite standard.

The results contained in this dissertation have not been submitted in part or full to any other University or Institute for the award of any degree or diploma.

(Dr. Shashank Deep)

Associate Professor,
Department of Chemistry
Indian Institute of Technology Delhi
New Delhi-110016

ACKNOWLEDGEMENTS

I am indebted to a number of individuals who guided, supported, or encouraged me over the course of my education in general and through my graduate studies in particular.

*Firstly, I would like to express my sincere gratitude to my supervisor **Dr. Shashank Deep** for the tremendous support, motivation, and patience he has shown throughout my Ph.D. His guidance in my research and thesis writing was priceless. I acknowledge him for encouraging independent and creative thought in his students. His efforts towards shaping up my career give me the privilege to proudly accept that I could not have found a better advisor and mentor.*

*I am highly grateful to **Prof. P.J. Hart** (University of Texas Health Science Center, San Antonio) for providing the plasmid for SOD1, without which my thesis could not be accomplished.*

*I am grateful to the members of my 'Scientific Research Committee', **Prof. Sunil Kumar Khare, Dr. Sameer Sapra and Prof. Rajiv Bhat** (School of Biotechnology, JNU) for their thoughtful suggestions and criticism, which have helped me to shape the work presented in this thesis. I wish to extend my thanks to **Dr. Bishwajit Kundu** (KSBS, IIT Delhi) for his valuable suggestions on my research work.*

I express my humble thanks to the past and the current Heads of Department and DRC chairpersons for their support. I am thankful to the Department of Chemistry and all its staff for providing an efficient infrastructure and working facilities for the research.

*The work presented in this thesis could not have been completed without the expertise and effort of all my collaborators over the years. I would like to thank **Dr. Shyam Kishor** (Department of chemistry, J.V college, Baraut, U.P.) for sparing his precious time in conducting the DFT studies that would prove critical in this work; **Dr. Samudrala Gourinath** (SLS, JNU) for letting me hang out in his lab for some months and giving me an opportunity to*

learn protein crystallization and **Dr. Rohit Kumar** for teaching me the protein crystallization techniques. I would like to convey my heartiest thanks to **Dr. Ankit Srivastava** (KSBS, IIT Delhi), for his help in AFM and cytotoxicity experiments and for his insightful discussions and suggestions.

I wish to extend my thanks to **Sneha** for sharing her knowledge regarding toxicity experiments and **Anu** for conducting time-resolved experiments. I would also like to thank **Sushma** for being always helpful and supportive during my thesis writing.

I would like to express my generous regards to all my colleagues and lab mates so far. I am thankful to all my seniors, **Komal ma'am, Unnati ma'am** and **Shahid Sir** for their supportive nature. I am grateful to **Ashhar sir, Amrita ma'm, Vinay, Preeti, Shivnetra, Aayushi, Nitika, Sonam, Meena and Fatima** for enriching my Ph.D. experience and for being cooperative with me in using Lab's Instrument. I would also like to thank all the M.Sc. and M.tech. Students who worked with me during this tenure. I am very thankful to **Priya** for assisting me in conducting experiments in the last year of my Ph.D. and for being always supportive.

I am extremely grateful to **Pardeep sir** for always being a great mentor and cheering me up during my rough moments. A very special thanks to **Nidhi Katyal** for the stimulating discussions, providing courage in tough times, for enormous support during the sleepless nights we were working together in order to meet the deadlines, for being extremely patient during my worse mood, for all the fun we had together in the last five years and for being the best listener I have ever had. I fall short of words to mention how much she means to me and the way she always stood by me.

I am also thankful to **Dr. Pramit Chaudhury** and **Dr. Siddharth Pandey** and their group for allowing me to use instruments in their lab whenever needed.

*I feel indebted to **Shruti Trivedi** for her immense moral support, motivation, and guidance and for always taking care of me like an elder sister. Great thanks to **Ankita, Nisha, Shweta, Neha, Utkarsh,** and **Ritika** for providing me with family-like feeling and comfort in IIT Delhi and for the unforgettable memories made together. I would like to thank **Ambika, Ashish, Sridhar, Nitish, Haimanti** and **Sohini ma'm, Samrat** and **Sourabh Sir**, whom I made wonderful friends with during my Ph.D. journey.*

*I would like to thank my **family**: my parents and my brother, who has been an invaluable support to me throughout my life. I would like to thank **my parents** for all their unconditional love, guidance, encouragement and patience. I am extremely grateful to **my brother Navjot**, for always being protective and assuring me that he is there in every situation of my life. I fall short of words thanking my parents and my brother as they were my greatest pillars of support and strength all this time. Heartfelt thanks to **my sister-in-law Mansi** and my adorable **niece Nadar** for all the lighter and cheerful moments we have shared. I owe my thanks to **my father-in-law, mother-in-law and brother-in-law Vinit** for their immense love, support, and patience.*

*Last but not the least, I would like to thank **my loving husband, Nitin Sharma**, for always believing in me, for all his love and care and for being a constant source of support and encouragement during the challenges of graduate school and life. He has been my great mentor as well as my best friend since even I began my graduate school. I feel extremely lucky for having him in my life.*

Finally, I would like to dedicate this thesis to my muma and papa, as without their blessings and enormous support, I would not have been able to accomplish my goals.

(Nidhi Kaur Bhatia)

ABSTRACT

Amyotrophic lateral sclerosis (ALS) is fatal and progressive neurodegenerative disease which is associated with the aggregation of Cu-Zn Superoxide Dismutase (SOD1). Unfortunately, effective therapeutics against ALS has not yet been developed. In the past few years, small molecules like polyphenols have been identified as potential anti-amyloidogenic agents and their biological properties are governed by the relative concentrations of their different conformational and ionic forms. Therefore, a thorough understanding of the conformational properties and mechanism of action of these polyphenols, which may serve as a potential inhibitor against SOD1 aggregation is needed. The thesis entitled '**Polyphenols and their effect on the aggregation of ALS-linked Cu-Zn superoxide dismutase (SOD1)**' is concerned with the understanding of physicochemical properties of three naturally occurring polyphenols (curcumin, quercetin, and baicalein) in nearly aqueous solution and their role as an anti-amyloidogenic agent against aggregation of ALS-linked SOD1. An attempt has also been made to understand the mechanism of their action.

The thesis is composed of seven chapters. **Chapter 1** (Introduction) provides an overview of SOD1 and its implication in ALS pathogenesis. detail review of aggregation of SOD1 and strategies are known so far, to inhibit SOD1 aggregation is also presented. The chapter then describes polyphenols and their therapeutic role against many chronic diseases as an anti-bacterial, anti-carcinogenic and anti-amyloidogenic agent. This chapter finally deals with the origin of the scientific problem associated with polyphenols and aggregation of SOD1 and the outline of the present research work done in this thesis in the context of addressing these problems. **Chapter 2** (Materials and Methodologies) deals with chemical procurement and protein expression, purification and various analytical techniques used for the investigation of aggregation of SOD1 and physicochemical properties of curcumin, baicalein, and quercetin. **Chapter 3** (Effect of pH and temperature on conformational equilibria and aggregation behaviour of curcumin in aqueous binary mixtures of ethanol) provides a detailed study on the effect of various factors like temperature, pH and solvent composition on keto-enol conformational equilibria and aggregation of curcumin. **Chapter 4** (Curcumin binds to the pre-fibrillar aggregates of

Cu/Zn superoxide dismutase (SOD1) and alters its amyloidogenic pathway resulting in reduced toxicity) describes, in details, the inhibition of DTT-induced SOD1 fibrillation by curcumin at physiological pH and temperature using ThT binding assay, DLS, AFM, TEM, ATR-FTIR. Binding parameters with native SOD1 were also estimated by tryptophan quenching experiments using steady state and time-resolved fluorescence spectroscopy. Docking studies demonstrated that putative binding sites of curcumin are the aggregation-prone regions of SOD1. MTT- assay was performed on THP1 cells to compare the cytotoxicity of SOD1 aggregates formed in the presence and absence of curcumin.

Chapter 5 (Effect of pH and temperature on physicochemical properties and aggregation behaviour of quercetin and baicalein in nearly aqueous media) deals with the detailed investigation of the effect of different solution conditions (pH and temperature) on physicochemical and aggregation behaviour of two naturally occurring flavonoids having same core structure (quercetin and baicalein) but varying hydroxyl groups in nearly aqueous media. **Chapter 6** (Quercetin and baicalein act as a potent anti-amyloidogenic and fibril destabilizing agents for SOD1 fibrils) presents a detailed study on the effect of quercetin and baicalein on the DTT-EDTA induced fibrillation of SOD1 at 37 °C and pH 7.4. The anti-amyloidogenic and disaggregating effect of both the flavonoids against SOD1 fibrillation was revealed by using ThT fluorescence, TEM and SDS PAGE. Binding parameters of these polyphenols were also estimated by quenching of tryptophan fluorescence and UV absorption spectra of quercetin and baicalein in the presence of varying concentration of SOD1. Putative binding sites of quercetin and baicalein were predicted by docking studies. The MTT assay was performed to determine the toxicity of SOD1 fibrils formed without and with flavonoids. **Chapter 7** (Summary and future perspectives) **contains** salient highlights of this work. In a nutshell, our findings have been instrumental in discovering and understanding the role of naturally occurring polyphenols (curcumin and quercetin) in the inhibition of aggregation of immature forms of SOD1 and in exploring the physicochemical properties of these polyphenols. Thus, our studies will help in providing the pathway for effective therapeutics against ALS and also have utility in terms of the enhancement of the bioavailability and therapeutic potential of polyphenols.

सार

एमीट्रोफिक पार्श्व स्वलेरोसिस (एएलएस) घातक और प्रगतिशील न्यूरोडिजेनेरेटिव बीमारी है जो Cu-Zn सुपरऑक्साइड डिस्मूटेज (एसओडी 1) के एकत्रीकरण से जुड़ा हुआ है। दुर्भाग्यवश, एएलएस के खिलाफ प्रभावी चिकित्सीय विकास अभी तक विकसित नहीं हुआ है। पिछले कुछ वर्षों में, पॉलीफेनॉल जैसे छोटे अणुओं को संभावित एंटी-एमीलाइडोजेनिक एजेंटों के रूप में पहचाना गया है और उनके जैविक गुणों को उनके विभिन्न संरचनात्मक और आयनिक रूपों के सापेक्ष सांद्रता द्वारा नियंत्रित किया जाता है। इसलिए, इन पॉलीफेनॉल की संरचना के गुणकारी गुणों और तंत्र की पूरी तरह से समझ, जो एसओडी 1 एकत्रीकरण के खिलाफ संभावित अवरोधक के रूप में कार्य कर सकती है, की आवश्यकता है। थीसिस 'पॉलीफेनॉल और एलएस-लिंकड क्व Cu-Zn सुपरऑक्साइड डिस्मूटेज (एसओडी 1) के एकत्रीकरण पर उनके प्रभाव को लगभग जलीय घोल में तीन स्वाभाविक रूप से होने वाले पॉलीफेनॉल (कर्व्यूमिन, क्वार्सीटिन और बाइकेलिन) के भौतिक रसायन गुणों की समझ से संबंधित है। एलएस-लिंकड एसओडी 1 के एकत्रीकरण के खिलाफ एंटी-एमीलाइडोजेनिक एजेंट के रूप में उनकी भूमिका। उनकी कार्रवाई के तंत्र को समझने के लिए भी एक प्रयास किया गया है।

थीसिस सात अध्यायों से बना है। अध्याय 1 (परिचय) एसओडी 1 और एएलएस रोगजन्य में इसके निहितार्थ का एक सिंहावलोकन प्रदान करता है। एसओडी 1 और रणनीतियों के एकत्रीकरण की विस्तृत समीक्षा अब तक ज्ञात है, एसओडी 1 एकत्रीकरण को बाधित करने के लिए भी प्रस्तुत किया जाता है। अध्याय में पॉलीफेनॉल और कई पुरानी बीमारियों के खिलाफ एंटी-बैक्टीरिया, एंटी-कैंसरजन्य और एंटी-एमीलाइडोजेनिक एजेंट के रूप में उनकी चिकित्सीय भूमिका का वर्णन किया गया है। अंततः यह अध्याय पॉलीफेनॉल से जुड़े वैज्ञानिक समस्या की उत्पत्ति और एसओडी 1 के एकत्रीकरण और इन समस्याओं को हल करने के संदर्भ में इस शोध में किए गए वर्तमान शोध कार्य की रूपरेखा से संबंधित है। अध्याय 2 (सामग्रियों और पद्धतियों) रासायनिक खरीद और प्रोटीन अभिव्यक्ति, शुद्धिकरण और विभिन्न विश्लेषणात्मक तकनीकों से संबंधित है जो एसओडी 1 के एकत्रीकरण और कर्व्यूमिन, बाइकेलिन और क्वार्सीटिन के भौतिक रसायन गुणों की जांच के लिए उपयोग किए जाते हैं। अध्याय 3 (पीएच का प्रभाव और इथेनॉल के जलीय बाइनरी मिश्रण में कर्व्यूमिन के समेकित संतुलन और तापमान के एकत्रीकरण व्यवहार पर तापमान) केटो-एनोल संरचनात्मक संतुलन और curcumin के एकत्रीकरण पर तापमान, पीएच और

विलायक संरचना जैसे विभिन्न कारकों के प्रभाव पर एक विस्तृत अध्ययन प्रदान करता है। अध्याय 4 (Curcumin पूर्व-फाइब्रिलर योग के लिए बांधता है क्यू / जेएन सुपरऑक्साइड विघटन (एसओडी 1) और इसके अमीलाइडोजेनिक मार्ग को बदलता है जिसके परिणामस्वरूप कम विषाक्तता होती है) विवरण में, डीटीटी प्रेरित एसओडी 1 फाइब्रिलेशन का अवरोध शारीरिक पीएच और तापमान पर टीसी बाध्यकारी परख, डीएलएस, एएफएम, टीईएम, एटीआर का उपयोग करके तापमान से होता है। -FTIR। देशी एसओडी 1 के साथ बाध्यकारी पैरामीटर का अनुमान स्थिर राज्य और समय-हल फ्लोरोसेंस स्पेक्ट्रोस्कोपी का उपयोग करके ट्राइपोफान क्वेंचिंग प्रयोगों द्वारा भी किया गया था। डॉकिंग अध्ययनों से पता चला है कि कर्क्यूमिन की रखरखाव बाध्यकारी साइटें एसओडी 1 के एकत्रीकरण-प्रवण क्षेत्र हैं। टीटीपी 1 कोशिकाओं पर एमटीटी-परख प्रदर्शन किया गया था ताकि curcumin की उपस्थिति और अनुपस्थिति में गठित एसओडी 1 योग की साइटोटोक्सिसिटी की तुलना की जा सके। अध्याय 5 (पीएच का प्रभाव और भौतिक रसायन गुणों पर तापमान और लगभग जलीय मीडिया में क्वार्सेटिन और बाइकेलिन का एकत्रीकरण व्यवहार) भौतिक रसायन पर दो अलग-अलग समाधान स्थितियों (पीएच और तापमान) के प्रभाव की विस्तृत जांच से संबंधित है और दो स्वाभाविक रूप से होने वाले फ्लेवोनोइड्स के एकत्रीकरण व्यवहार लगभग मूल संरचना (क्वार्सेटिन और बाइकेलिन) होने पर लगभग जलीय मीडिया में हाइड्रोक्साइल समूह अलग-अलग होते हैं। अध्याय 6 (क्वार्सेटिन और बाइकेलिन एसओडी 1 फाइब्रिल के लिए एक शक्तिशाली एंटी-एमिलाइडोजेनिक और फाइब्रिल अस्थिर एजेंटों के रूप में कार्य करते हैं) एसओडी 1 के डीटीटी-ईडीटीए प्रेरित फाइब्रिलेशन पर 37 डिग्री सेल्सियस और पीएच 7.4 पर क्वार्सेटिन और बाइकेलिन के प्रभाव पर विस्तृत अध्ययन प्रस्तुत करते हैं। एसओडी 1 फाइब्रिलेशन के खिलाफ फ्लेवोनोइड्स के एंटी-एमिलाइडोजेनिक और असंतुलित प्रभाव को टीटी फ्लोरोसेंस, टीईएम और एसडीएस पेज का उपयोग करके पता चला था। इन पॉलीफेनॉल के बाध्यकारी मानकों का भी एसओडी 1 की विभिन्न एकाग्रता की उपस्थिति में ट्राइपोफान फ्लोरोसेंस और क्वार्सेटिन और बाइकेलिन के यूवी अवशोषण स्पेक्ट्रा को बुझाने का अनुमान लगाया गया था। Quercetin और baicalein की रखरखाव बाध्यकारी साइटों डॉकिंग अध्ययन द्वारा भविष्यवाणी की गई थी। एमटीटी परख flavonoids के बिना और उसके साथ गठित एसओडी 1 फाइब्रिल की विषाक्तता निर्धारित करने के लिए किया गया था। अध्याय 7 (सारांश और भविष्य के दृष्टिकोण) में इस काम की मुख्य हाइलाइट्स शामिल हैं। संक्षेप में, हमारे निष्कर्ष एसओडी 1 के अपरिपक्व रूपों के एकत्रीकरण और इन पॉलीफेनॉल के भौतिक रसायन गुणों की खोज में प्राकृतिक रूप से

होने वाले पॉलीफेनॉल (कवर्क्यूमिन और क्वार्सेटिन) की भूमिका को खोजने और समझने में महत्वपूर्ण भूमिका निभा चुके हैं। इस प्रकार, हमारे अध्ययन एएलएस के खिलाफ प्रभावी चिकित्सकीय मार्ग के मार्ग प्रदान करने में मदद करेंगे और जैव उपलब्धता और पॉलीफेनॉल की चिकित्सकीय क्षमता के संवर्द्धन के संदर्भ में उपयोगिता भी करेंगे।

TABLE OF CONTENTS

CERTIFICATE	I
ACKNOWLEDGEMENTS	II
ABSTRACT	V
TABLE OF CONTENTS	VII
LIST OF FIGURES	XIII
LIST OF TABLES	XXIII
LIST OF ABBREVIATIONS	XXV
Chapter 1 Introduction	1
1.1 Protein folding, misfolding and aggregation	1
1.1.1 Protein folding pathway	1
1.1.2 Protein misfolding diseases: Neurodegenerative diseases	2
1.1.3 Toxicity of protein aggregates and neurodegeneration	5
1.1.4 Protein folding versus aggregation	5
1.2 Mechanism of formation of amyloid fibrils	7
1.2.1 Nucleation dependent polymerization (NDP)	6
1.2.2 Templated assembly model	8
1.2.3 Monomer directed conversion model	8
1.2.4 Nucleated conformation conversion model	9
1.3 Diagnostic tools for structural characterization and elucidation of fibrils and their precursors in amyloid fibril formation pathway	9
1.4 Factors affecting protein aggregation	14
1.4.1 Intrinsic factors	14

1.4.2 Extrinsic factors in vivo	14
1.4.3 Physicochemical factors	14
1.4.4 External factors in vitro (Excipients)	16
1.5 Inhibition of amyloid formation (Strategies and inhibitors)	17
1.6 Polyphenols	18
1.7 Amyotrophic Lateral Sclerosis (ALS)	19
1.8 Cu/Zn Superoxide Dismutase (SOD1)	20
1.9 ALS pathogenesis	22
1.9.1 Hypothesis for SOD1 mediated toxicity	22
1.9.2 Neurodegeneration in SOD1 mediated ALS	25
1.9.3 Treatment of ALS (Inhibition of aggregation, a facile strategy)	27
1.10 Origin of the problem in the context of this thesis	28
1.11 Outline of the present research problem	31
1.12 References	33
Chapter 2 Materials and Methodologies	48
2.1 Introduction	48
2.2 Chemicals	48
2.3 Expression and purification of SOD1 protein	49
2.3.1 Expression and purification of holo wt-SOD1	49
2.3.2 Expression and purification of N ¹⁵ labeled holo wt SOD1	50
2.4 Column Chromatography	50
2.4.1 Immobilized metal ion affinity chromatography	50
2.4.2 Size Exclusion Column (SEC) chromatography	51
2.5 Ultraviolet-Visible (UV-Vis) Spectroscopy	51
2.6 Circular Dichroism (CD) spectroscopy	53
2.7 Fluorescence Spectroscopy	55
2.8 Nuclear magnetic resonance (NMR) spectroscopy	58
2.9 Attenuated Total Reflection (ATR) Fourier Transform Infrared spectroscopy (FTIR) spectroscopy	59

2.10 Dynamic Light Scattering (DLS)	60
2.11 Transmission electron microscopy (TEM)	61
2.12 Multivariate curve resolution (MCR) analysis	62
2.13 References	63
Chapter 3 Effect of pH and temperature on conformational equilibria and aggregation behaviour of curcumin in aqueous binary mixtures of ethanol	66
3.1 Introduction	66
3.2 Methods and materials	68
3.2.1 Chemicals	68
3.2.2 Preparation of samples	68
3.2.3 UV-Vis Absorption Spectroscopy	69
3.2.4 Steady-state fluorescence spectroscopy	69
3.2.5 Dynamic light scattering (DLS)	69
3.2.6 Density functional theory (DFT) calculations	70
3.3 Results and Discussion	70
3.3.1 DFT calculations of absorption and emission behaviour of different forms of curcumin	70
3.3.2 Effect of the increase in ethanol content on conformational equilibria of the curcumin	76
3.3.3 Effect of pH on the absorption and emission behaviour of curcumin in 5% ethanolic solution	82
3.3.4 Effect of temperature on the absorption and the emission behaviour of curcumin in 5% ethanolic solution	85
3.3.5 Effect of concentration on the physiochemical properties of curcumin in 5% ethanolic solution	89
3.3.6 Estimation of thermodynamic parameters	96
3.4 Conclusions	99
3.5 References	101

Chapter 4 Curcumin binds to the pre-fibrillar aggregates of Cu/Zn superoxide dismutase (SOD1)	105
4.1 Introduction	105
4.2 Methods and materials	106
4.2.1 Chemicals	106
4.2.2 Expression and purification of hSOD1	106
4.2.3 Preparation of samples for aggregation studies	107
4.2.4 Fluorescence assays	109
4.2.5 Fluorescence quenching studies	110
4.2.6 Time-resolved fluorescence lifetime decay measurements	111
4.2.7 Attenuated total reflection Fourier transform infrared spectroscopy (ATR-FTIR)	111
4.2.8 Atomic Force Microscopy (AFM)	111
4.2.9 Transmission electron microscopy (TEM)	112
4.2.10 Docking of SOD1 and curcumin using Autodock	112
4.2.11 Cell culture and MTT assay	113
4.3 Results and Discussion	114
4.3.1 Curcumin inhibits the fibril formation propensity of reduced SOD1	114
4.3.2 DTT-treated SOD1 forms disordered aggregates in the presence of Curcumin	115
4.3.3 Comparatively smaller prefibrillar aggregates are formed in curcumin containing samples	118
4.3.4 Curcumin binds to SOD1 with moderate affinity	121
4.3.5 Docking suggests that the binding of curcumin is primarily to the aggregation-prone region of SOD1	123
4.3.6 Curcumin binds to the oligomers and pre-fibrillar aggregates of SOD1 with higher affinity	126
4.3.7 Curcumin reduces the cytotoxicity of the aggregates	127

4.4 Conclusions	129
4.5 References	131
Chapter 5 Effect of pH and temperature on physicochemical properties and aggregation behaviour of quercetin and baicalein in nearly aqueous media	136
5.1 Introduction	136
5.2 Materials and methods	138
5.2.1 Chemicals	138
5.2.2 Preparation of samples	138
5.2.3 UV-Vis absorption and steady state fluorescence spectroscopy	138
5.2.4 Dynamic light scattering (DLS)	138
5.2.5 Density functional theory (DFT)	139
5.3 Results and Discussion	140
5.3.1 Effect of pH on the absorption behaviour of quercetin and baicalein	140
5.3.2 Effect of temperature on the absorption behaviour of quercetin and baicalein	143
5.3.3 Effect of pH on emission behaviour of quercetin and baicalein	146
5.3.4 Effect of pH, temperature and concentration of quercetin or baicalein on their particle size distribution	149
5.4 Conclusions	153
5.5 References	154
Chapter 6 Quercetin and baicalein act as a potent anti-amyloidogenic and fibril destabilizing agents for SOD1 fibrils	157
6.1 Introduction	157
6.2 Methods and materials	158
6.2.1 Chemicals	158
6.2.2 Expression and purification of SOD1	158
6.2.3 Sample preparation	158
6.2.4 Fluorescence assays	158
6.2.5 Quenching Experiments	159

6.2.6 Time-resolved fluorescence lifetime decay measurements	159
6.2.7 Transmission Electron Microscopy (TEM)	160
6.2.8 Docking	160
6.2.9 SDS PAGE	160
6.2.10 UV- Vis absorption spectroscopy	161
6.2.11 Cytotoxicity assay: MTT assay	161
6.3 Results and Discussion	162
6.3.1 Both the flavonoids (quercetin and baicalein) inhibited the fibrillation of SOD1	162
6.3.2 Quercetin and baicalein affects the rate of elongation significantly	163
6.3.3 Both the flavonoids modulate the aggregation pathway of SOD1	165
6.3.4 Quercetin and baicalein destabilizes the SOD1 fibrils	166
6.3.5 Cytotoxicity of aggregates and the fragmented fibrils formed in the presence of quercetin and baicalein	167
6.3.6 Flavonoids reduced the exposure of hydrophobic patches in the aggregates of SOD1	170
6.3.7 Quercetin and baicalein interact differently with native SOD1	171
6.3.8 Quercetin and baicalein binds moderately to native SOD1	172
6.3.9 Docking	175
6.4 Conclusions	180
6.5 References	182
Chapter 7 Summary and Future Prospectives	186
Curriculum Vitae	191

LIST OF FIGURES

Figure No.	Figure Caption	Page No.
Figure 1.1	Combined energy landscape of protein folding and aggregation. Adapted from reference.	2
Figure 1.2	Illustration of nucleation dependent polymerization (NDP) model of protein aggregation .	7
Figure 1.3	Illustration of templated assembly model of protein aggregation	8
Figure 1.4	Illustration of monomer directed conversion model of protein Aggregation.	8
Figure 1.5	Illustration of nucleated conformation conversion model of protein aggregation.	9
Figure 1.6	Structure of human superoxide dismutase 1 (SOD1). The metal copper and zinc depicted by red and green spheres respectively. The intramolecular disulphide bond between cys 57 and cys 146 is coloured in black.	21
Figure 1.7	Generalized model for aggregation of SOD1 based on various in-vivo and in-vitro studies.	24
Figure 1.8	Generalized model depicting the various cell pathways and organelles being affected leading to neurodegeneration caused by mutant SOD1 (adapted from reference).	26
Figure 1.9	Structure and the natural source of three naturally occurring polyphenols: curcumin, quercetin and baicalein.	30
Figure 2.1	(a) SDS PAGE gel of wild-type holo SOD1 purified by Ni-affinity column, (b) SEC profile of purified wild-type holo SOD1 (dimer).	50
Figure 2.2	Absorption spectrum of SOD1 for determination of its concentration.	52
Figure 2.3	Standard far UV CD spectra of protein representing its secondary structural elements α -helix (solid curve), antiparallel β -sheet. (long dashes), type 1 β -turn (dots) and irregular structure (dots and short dashes). Adapted from Reference	54
Figure 2.4	Far-UV CD spectrum of holo SOD1.	54

Figure 2.5	1H-15N HSQC spectrum of purified holo SOD1 (with reduced copper) at pH 5.0.	59
Figure 3.1	Different isomeric and tautomeric forms of curcumin.	66
Figure 3.2	Two different dimeric forms of curcumin.	71
Figure 3.3	Theoretical UV absorption spectra (left panel), theoretical emission spectra (right panel) of different forms of curcumin in binary mixtures of ethanol and water (5% ethanol) calculated using a DFT approach.	72
Figure 3.4	Absorption spectra of curcumin (25 μ M) in solutions of different volume percentages of ethanol at pH 7.0.	76
Figure 3.5	Calculation of molar extinction coefficient (ϵ) of curcumin in 5%, 25%, 50% and 100 % ethanol.	77
Figure 3.6	(a) Change in the λ_{app} and absorbance (height of peak) of curcumin at 429 nm for solutions with different volume percentages of ethanol. (b) The relative percentage of the diketo and enol forms of curcumin (25 μ M), with an increase in the volume percentage of ethanol. (c) Change in FWHM of the 429 nm peak for solutions with different volume percentages of ethanol.	78
Figure 3.7	The pure spectra of the components and concentration distribution of the components obtained by using MCR approach.	79
Figure 3.8	(a) Emission spectra of curcumin (25 μ M) in solutions with different volume percentages of ethanol at pH 7.0. (b) Normalized emission spectra of curcumin (25 μ M) in solutions with different volume percentages of ethanol.	80
Figure 3.9	Relative increase in the ϵ_{app} value of curcumin at 429 nm (red circle) and relative increase in the area under peak at 548 nm (black circle) as a function of volume % of ethanol. There is a small change in the ϵ_{app} value of curcumin at 429 nm as seen in Inset.	81
Figure 3.10	Absorption spectra of curcumin (25 μ M) at different pH in 5% ethanolic solution.	82
Figure 3.11	Resolved spectrum of curcumin (25 μ M) at pH 9.0 at 25 $^{\circ}$ C.	83

Figure 3.12	(a) Calibration graph for calculation of the molar extinction coefficient (ϵ) of curcumin pH 2.0, 7.0 and 9.0. Change in (b) ϵ_{app} value of curcumin at 429 nm and absorbance (height) at 429 nm, (c) FWHM at different pH.	84
Figure 3.13	Emission spectra of curcumin (25 μ M) at different pH in 5% ethanolic solution. We speculate that the shoulder at 488 nm might be due to the formation of a higher order of curcumin aggregates since curcumin is known to have lower solubility at acidic pH.	85
Figure 3.14	Effect of temperature on the absorption spectra of curcumin at (a) pH 2.0, (b) pH 7.0 and (c) pH 9.0 in 5% ethanolic solutions and at (d) pH 2.0, (e) pH 7.0 and (f) pH 9.0 in 50% ethanolic solutions.	86
Figure 3.15	(a) Ratio of A357/A429 versus temperature (Celsius) for curcumin at pH 7.0 in 5% ethanol. Absorption spectra of curcumin at pH 7.0 (b) and pH 9.0 (c) in 5% ethanol at different time intervals.	86
Figure 3.16	Effect of temperature on the emission spectra of curcumin at pH 2.0 (a and b) and pH 7.0 (c and d) in 5% ethanolic solutions with $\lambda_{ex} = 429$ nm (left panel) and 350 nm (right panel).	88
Figure 3.17	Effect of concentration on the emission spectra of curcumin at pH 2.0 (upper panel) and pH 7.0 (lower panel) in 5% ethanol solution at 25 °C.	90
Figure 3.18	Effect of temperature on the emission spectra of curcumin (70 μ M) at (a) pH 2.0 and (b) pH 7.0 in 5% ethanolic solutions with $\lambda_{ex} = 429$ nm.	91
Figure 3.19	Particle size distribution of curcumin at different concentrations. 25 μ M (a), 70 μ M (b), 100 μ M (c) in 5% ethanolic solution. Comparison of particle size of curcumin under different conditions at pH 2.0 (d).	92
Figure 3.20	Particle size distribution of curcumin at different concentrations 25 μ M (a), 70 μ M (b), 100 μ M (c) in 5% ethanolic solution. Comparison of particle size of curcumin under different conditions at pH 7.0 (d).	93
Figure 3.21	The loadings (left panel) and coefficient of loadings (right panel) of each principal component as determined by PCA analysis of absorption spectral variation.	93

Figure 3.22	Number of components and pure spectra for each component extracted by employing MCR approach.	94
Figure 3.23	Concentrations of keto and enol forms at pH 2.0 (a), 7.0 (b) and 9.0 (c) estimated by MCR analysis. (d) Variation of equilibrium constants for keto-enol tautomerization of curcumin with the temperature at pH 2.0 and pH 7.0.	95
Figure 3.24	Variation of ΔG , ΔH and $-T\Delta S$ with temperature at (a) pH 2.0 and (b) pH 7.0.	98
Figure 3.25	Schematic representation of the overall behaviour of curcumin in aqueous media under different conditions.	100
Figure 4.1	a) SOD1 unlabeled, NEM labeled SOD1 and NEM labeled DTT treated SOD1 were separated on an SDS- PAGE under non reducing conditions. b) Observed and calculated m/z and peptide sequences of SOD1 and NEM labeled DTT treated SOD1 clearly depicting the reduction of intra-molecular disulfide bond (37-69-S-S-144-153), c) Representative MALDI-TOF mass data of tryptic fragments of non-reduced SOD1 and inset shows the intact disulfide bridge (Cys57-S-S-Cys146) d) Representative MALDI-TOF mass data of trypsin-digested reduced SOD1 labeled with NEM and inset shows the reduced SOD1 fragment labeled with NEM (37-70-S-NEM).	107
Figure 4.2	SEC profiles of native SOD1 and DTT (100 mM) treated SOD1. (Inset) The standard calibration curve of: Lysozyme (13.3 kD), Myoglobin (16.7 kD), Bovine Carbonic Anhydrase (BCA, 29.3 kD), Bovine Serum Albumin (BSA, 66.6 kD). SOD1 monomer and dimer are represented by red and black squares, respectively.	109
Figure 4.3	Kinetic profiles of DTT-treated SOD1 (pH 7.4) aggregation at 37 °C, alone and in the presence of curcumin (100, 200, 300 μ M), monitored by ThT fluorescence.	114
Figure 4.4	AFM images of aggregates of DTT-treated SOD1 (a) alone and (b) in the presence of curcumin (300 μ M), respectively. Scale bars are 0.5 μ m. TEM images of DTT-treated SOD1 (c) alone and (d) in the presence of curcumin (300 μ M) respectively. Scale bars are 100 nm.	115

Figure 4.5	Deconvoluted ATR-FTIR spectra of DTT-treated SOD1 (pH 7.4), alone (upper panel) and with curcumin (lower panel), incubated at 37 °C for 72 h.	116
Figure 4.6	Deconvoluted ATR- FTIR spectra of DTT-treated SOD1 (pH 7.4), alone (left panel) and with curcumin (right panel) at 0 hr.	117
Figure 4.7	Kinetics of aggregation of DTT-treated SOD1 samples (alone and with curcumin) monitored by Rayleigh Light scattering at 37 °C, pH 7.4.	118
Figure 4.8	SDS-PAGE analysis of soluble fractions of DTT-treated SOD1, alone (B) and with curcumin (A), at different incubation times.	119
Figure 4.9	Particle size distribution of different soluble species of DTT-treated SOD1 (pH 7.4) at various incubation times at 37 °C.	119
Figure 4.10	Particle size distribution of DTT treated SOD1 at 0 h before aggregation. Inset is the enlarged view near the peak.	120
Figure 4.11	Fluorescence emission spectra of DTT-treated SOD1 in phosphate buffer, pH 7.4 at 25 °C in the presence of various concentrations of curcumin (0, 1, 3, 4, 5, 10, 20, 40 μM).	121
Figure 4.12	(a) Stern Volmer plot for tryptophan quenching of DTT treated SOD1 (5 μM, pH7.4) by curcumin at 25 °C. Modified Stern Volmer plots (b) and (c) for the estimation of the fraction of accessible tryptophan, number of binding sites and the binding constant.	122
Figure 4.13	(a) Time-resolved fluorescence lifetime decay profiles of DTT-treated (2 μM), alone and with curcumin (5 μM) (b) The Stern Volmer plot of fluorescence lifetime of DTT-treated SOD1 at various concentrations of curcumin (0, 0.5, 1, 5, 10, 20, 30 μM).	122
Figure 4.14	Left panel: Putative binding site of curcumin in the SOD1-curcumin complex obtained using AutoDock. Right panel: The interactions between the protein and curcumin as observed from Ligplot. In the middle is curcumin molecule shown as blue line surrounded by residues of SOD1. Green lines and half moon represent hydrogen bonding and hydrophobic interactions. Residues in black font are involved in hydrophobic bonding whereas residues in green font are involved in hydrogen bonding.	124

- Figure 4.15 Left panel: One of the putative binding site of curcumin in the SOD1 monomer (chain B) -curcumin complex obtained using AutoDock. Right panel: The interactions between the protein and curcumin as observed from Ligplot. In the middle is curcumin molecule shown as blue line surrounded by residues of SOD1 monomer. Green lines and half moon represent hydrogen bonding and hydrophobic interactions respectively. Residues in black font are involved in hydrophobic interactions whereas residues in green font are involved in hydrogen bonding. 125
- Figure 4.16 The fluorescence intensity of the curcumin on its addition to the aliquot of DTT treated SOD1 samples incubated alone for 0, 8, 22, 32 and 56 h at 37 °C. The inset shows the curcumin fluorescence spectra of the resulting solution after its addition. 126
- Figure 4.17 AFM images of reduced SOD1 alone, (a) after 6 h, (b) 24 h (c) 72 h. Scale Bars are 0.5 μM. 126
- Figure 4.18 MTT reduction by THP1 cells in the presence of aggregates of DTT-treated SOD1 incubated alone and with curcumin formed at incubation times of 0 h, 6 h, 24 h and 72 h. *p < 0.05, **p < 0.001. 128
- Figure 4.19 Schematic representation showing the effect of curcumin on the amyloidogenic pathway of reduced SOD1. 130
- Figure 5.1 (a) Absorption spectra of quercetin (30 μM) at different pH at 25 °C in 5% ethanolic aqueous solution, (b) the pure spectra of components of quercetin obtained by MCR approach (c) concentration distribution of pure components estimated by MCR analysis. 140
- Figure 5.2 (a) Structures of different forms of quercetin (b) Theoretical absorption spectra of different forms of quercetin in binary mixture of ethanol and water (5% ethanol) obtained by DFT approach. 141
- Figure 5.3 (a) Absorption spectra of baicalein (30 μM) at different pH at 25 °C in 5% ethanolic aqueous solution, (b) the pure spectra of components of baicalein obtained by MCR approach (c) concentration distribution of pure components estimated by MCR analysis. 142

Figure 5.4	(a) Structures of different forms of baicalein (b) Theoretical absorption spectra of different forms of baicalein in a binary mixture of ethanol and water (5% ethanol).	143
Figure 5.5	Effect of temperature on absorption spectra of quercetin (30 μ M) at three different pH, (a) pH 2.0, (b) pH 7.0 and (c) pH 9.0 in 5% ethanolic aqueous solutions.	144
Figure 5.6	Time dependent variation in absorption spectra of quercetin (30 μ M) at three different pH, (a) pH 2.0, (b) pH 7.0 and (c) pH 9.0 in 5% ethanolic aqueous solutions.	144
Figure 5.7	Effect of temperature on absorption spectra of baicalein (30 μ M) at three different pH, (a) pH 2.0, (b) pH 7.0 and (c) pH 9.0 in 5% ethanolic aqueous solutions.	146
Figure 5.8	Emission spectra of quercetin (30 μ M) at different pH in 5% ethanolic buffered solutions.	147
Figure 5.9	(a) Emission spectra of baicalein (30 μ M) at different pH in 5% ethanolic solution. (b) The combined plot of absorbance of baicalein at 362 nm and fluorescence intensity of baicalein at 360 nm as function of pH.	149
Figure 5.10	Particle size distribution of quercetin (30 μ M and 70 μ M) in ethanol and 5% ethanolic solution at pH 2.0 and 7.0 at 25 $^{\circ}$ C.	150
Figure 5.11	Particle size distribution of baicalein (30 μ M and 70 μ M) in ethanol and 5% ethanolic solution at pH 2.0 and 7.0 at 25 $^{\circ}$ C.	151
Figure 5.12	Particle size distribution of (a) quercetin (30 μ M) at pH 2.0, (b) quercetin (70 μ M) at pH 2.0, (c) quercetin (30 μ M) at pH 7.0 and (d) quercetin (70 μ M) at pH 7.0 at three different temperatures 25 $^{\circ}$ C, 50 $^{\circ}$ C and 70 $^{\circ}$ C.	151
Figure 5.13	Particle size distribution of (a) baicalein (30 μ M) at pH 2.0, (b) baicalein (70 μ M) at pH 2.0, (c) baicalein (30 μ M) at pH 7.0 and (d) baicalein (70 μ M) at pH 7.0 at three different temperatures 25 $^{\circ}$ C, 50 $^{\circ}$ C and 70 $^{\circ}$ C.	152

Figure 6.1	Kinetics of DTT and EDTA induced fibrillation of SOD1 alone and in the presence of (a) quercetin and (b) baicalein, monitored by ThT fluorescence.	162
Figure 6.2	TEM images of SOD1 fibrils (i) alone and in the presence of (ii) quercetin and (iii) baicalein.	163
Figure 6.3	Normalized titration curves of ThT fluorescence assay to monitor the kinetics of fibrillation of SOD1 in the absence and presence of (a) quercetin and (b) baicalein (c) macroscopic kinetics profile obtained by model simulations. Adapted from reference.	164
Figure 6.4	Kinetics of fibrillation of SOD1 after seeding with SOD1 fibrils in the absence and presence of quercetin and baicalein.	165
Figure 6.5	Time-dependent ThT assay for fibrillation of SOD1 and effect of the addition of flavonoids at different time intervals (6 h, 12 h, 30 h) alone and in the presence of (a) quercetin and (b) baicalein.	165
Figure 6.6	TEM images of SOD1 fibrils formed after incubation of 30 h (i) without flavonoids and preformed fibrils incubated with (ii) quercetin and (iii) baicalein.	166
Figure 6.7	Reducing SDS-PAGE of SOD1 aggregated samples of fibrillation and disaggregation assay after 30 h of incubation without flavonoid (LANE 2), with quercetin (LANE 3) and baicalein (LANE 5) added at 0 h, with quercetin (LANE 4) and baicalein (LANE 6) added to pre-existing fibrils. LANE 1 represents the standard molecular weight markers.	167
Figure 6.8	Representative micrographs of neuronal SHSY5Y cells morphology after treatment with buffer alone, quercetin or baicalein alone, SOD fibrils formed alone and with quercetin and baicalein and the existing fibrils treated with quercetin or baicalein. Control represents the untreated cells.	168
Figure 6.9	MTT reduction by the untreated (control) and treated neuronal SHSY5Y cells with SOD1 fibrils alone, SOD1 aggregated formed in the presence of flavonoids (quercetin or baicalein) and treated preformed fibrils with the flavonoids *p < 0.05, **p < 0.001, NS: Not significant.	169

Figure 6.10	Time-dependent change in ANS emission wavelength maximum of EDTA and DTT treated SOD1 samples incubated at (37 °C) and pH (7.4) in the absence and presence of flavonoids (quercetin and baicalein).	170
Figure 6.11	The absorption spectra of flavonoid (a) quercetin and (b) baicalein in the presence of varying concentrations of SOD1 (0-40 μM).	171
Figure 6.12	Tryptophan fluorescence emission spectra of SOD1 in the presence of varying concentrations of flavonoid (0-40 μM) (a) quercetin and (b) baicalein.	172
Figure 6.13	The Stern Volmer plot of fluorescence lifetime of SOD1 at various concentrations of flavonoid (quercetin or baicalein) (0, 5, 10, 15 μM).	173
Figure 6.14	Stern Volmer plot for tryptophan quenching of SOD1 (10 μM, pH 7.4) by quercetin at 25 °C. Modified Stern Volmer plots (b) and (c) for the estimation of the fraction of accessible tryptophan, number of binding sites and the binding constant.	174
Figure 6.15	Stern Volmer plot for tryptophan quenching of SOD1 (10 μM, pH 7.4) by baicalein at 25 °C. Modified Stern Volmer plots (b) and (c) for the estimation of the fraction of accessible tryptophan, number of binding sites and the binding constant.	174
Figure 6.16	Putative binding site of quercetin (a) and baicalein (b) in the SOD1-Flavonoid complex obtained using AutoDock. The interactions between the SOD1 and quercetin (c) or baicalein (d) as observed from Ligplot. In the middle is quercetin or baicalein surrounded by residues of SOD1 and green lines and half-moon represent hydrogen bonding and hydrophobic interactions. Residues in black font are involved in hydrophobic bonding whereas residues in green font are involved in hydrogen bonding.	175
Figure 6.17	Putative binding site of (a) quercetin and (b) baicalein for 11 residues segment oligomeric state of SOD1 (ref, PDB:5dli) obtained using Autodock. Surface representation of binding sites (c) in yellow, highlighting the quercetin and (d) in pink, highlighting the baicalein.	176

- Figure 6.18 Putative binding site of (a) quercetin and (b) baicalein in non-native SOD1 trimer1 obtained using Autodock. Surface representation of binding sites (c) in yellow, highlighting the quercetin and (d) in magenta, highlighting the baicalein. 177
- Figure 6.19 Putative binding site of (a) quercetin and (b) baicalein in a SOD1 monomeric unit derived from SOD1 trimer 1 obtained using Autodock. Surface representation of binding sites (c) quercetin and (d) baicalein. 178
- Figure 6.20 Schematic representation of the role of flavonoids (quercetin and baicalein) in inhibition and disaggregation of SOD1 fibrils. 179

LIST OF TABLES

Table No.	Title	Page No.
1.1	Protein misfolding diseases caused by loss of function of protein.	3
1.2.	List of neurodegenerative diseases associated with protein misfolding and amyloid formation.	4
1.3	List of some non-neurodegenerative amyloid diseases	4
1.4	The list of some of proteins whose intermediates in the fibril formation pathway have been characterized along with techniques used for their characterization. (Adapted from reference)	10
1.5	The list of some of proteins whose oligomers in their fibril formation pathway have been characterized along with techniques used for characterization. (Adapted from reference).	11
1.6	The list of some of proteins whose protofibrils/fibrils in their fibril formation pathway have been characterized along with techniques used for characterization. (Adapted from reference).	13
1.7	Examples of various polyphenols having inhibitory effect on protein aggregation.	19
2.1	Secondary structural elements of HSOD1 expressed in BL21.	55
3.1	Relative energies of different forms of curcumin calculated by using DFT approach.	72
3.2	The theoretical absorption parameters for curcumin calculated using DFT.	73
3.3	The theoretical emission parameters for curcumin calculated using DFT.	75
3.4	Parameters (ϵ and R^2) obtained from fitting of the calibration curve of curcumin in solutions with different percentage of ethanol.	77

3.5	Parameters (ϵ and R^2) obtained from fitting of the calibration curve of curcumin at different pH.	83
3.6	Thermodynamic parameters for keto-enol tautomerization equilibrium at pH 2.0.	96
3.7	Thermodynamic parameters for keto-enol tautomerization equilibrium at pH 7.0.	97
4.1	Secondary structure assignments of amide I peaks of aggregates obtained on incubation of DTT treated SOD1 alone and with curcumin.	117
4.2	Hydrodynamic radii of various species formed at different incubation times.	120
4.3	Parameters obtained by fitting of Stern-Volmer plot and modified Stern-Volmer plots.	123
5.1	The theoretical absorption parameters for quercetin calculated using DFT.	141
5.2	The theoretical absorption parameters for baicalein calculated using DFT.	143
5.3	Table 5.3 The theoretical emission parameters for quercetin calculated using DFT.	148
5.4	Table 5.4 The theoretical emission parameters for baicalein calculated using DFT.	149
6.1	Kinetic parameters obtained by global fitting of kinetic curves for DTT-EDTA induced fibrillation of SOD1 in the presence of quercetin and baicalein.	163
6.2	Parameters obtained by fitting of Stern-Volmer plot and modified Stern-Volmer plots.	175
6.3	Theoretical (from docking studies) and experimental (from quenching studies) binding parameters for quercetin-SOD1 dimer complex and baicalein-SOD1 dimer complex.	176
6.4	Theoretical binding parameters obtained from docking studies.	179

LIST OF ABBREVIATIONS

ALS	Amyotrophic Lateral Sclerosis
ANS	1-anilinonaphthalene-8-sulphonate
AAS	Atomic absorption spectrometry
AFM	Atomic Force Microscopy
ATR-FTIR	Attenuated total reflection fourier transform infrared spectroscopy
CD	Circular dichroism
DTT	Dithiothreitol
DFT	Density functional theory
DLS	Dynamic light scattering
EDTA	Ethylenedinitrilotetraacetic acid disodium salt (dihydrate)
ESPT	Excited state proton transfer
IPTG	Isopropyl-b-D-thio galactose
IMAC	Immobilized metal ion affinity chromatography
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5 diphenyltetrazolium bromide
MCR	Multivariate curve resolution
MALDI-TOF	Matrix-assisted laser desorption/ionization–time of flight
NTA	Nitrilotriacetic acid
NMR	Nuclear magnetic resonance
NDP	Nucleation dependent polymerization
NEM	N-ethylmaleimide
RMSD	Root mean square deviation
SOD1	Superoxide dismutase 1
SEC	Size exclusion chromatography
SDS-PAGE	Sodium dodecyl sulfate polyacrylamide gel Electrophoresis

TEM	Transmission electron microscopy
TDFT	Time dependent density functional theory
ThT	Thioflavin T
UV	Ultra violet
WT	Wild type