

**HARNESSING THE RHIZOBIOME AND PATHOBIOME  
FOR MITIGATION OF FUSARIUM WILT IN  
*CAJANUS CAJAN***

**RASHI TYAGI**



**DEPARTMENT OF BIOCHEMICAL ENGINEERING & BIOTECHNOLOGY  
INDIAN INSTITUTE OF TECHNOLOGY DELHI**

**JANUARY 2026**

© **Indian Institute of Technology Delhi (IITD), New Delhi, 2026**

**Harnessing the rhizobiome and pathobiome for  
mitigation of Fusarium wilt in *Cajanus cajan***

**by**

**RASHI TYAGI**

**DEPARTMENT OF BIOCHEMICAL ENGINEERING  
AND BIOTECHNOLOGY**

*Submitted*

*In fulfilment of the requirements of the degree of  
Doctor of Philosophy*

to the



**INDIAN INSTITUTE OF TECHNOLOGY DELHI**

**JANUARY 2026**

*DEDICATED TO  
THE DIVINE  
CREATOR, MY  
GRANDPARENTS,  
PARENTS, AND  
SISTER*

## CERTIFICATE

This is to certify that the thesis entitled “**Harnessing the rhizobiome and pathobiome for mitigation of Fusarium wilt in *Cajanus cajan***” submitted by **Ms. Rashi Tyagi** has been prepared under my guidance with the rules and regulations of Indian Institute of Technology Delhi, India. The results presented in this thesis have not been submitted for any degree or diploma in any other institute or university.

(Dr. Shilpi Sharma)

Professor

Department of Biochemical Engineering and Biotechnology

Indian Institute of Technology Delhi

Hauz Khas, New Delhi – 110016, India

## **ACKNOWLEDGEMENT**

*Earning a doctoral degree has long been a cherished dream of mine, and after more than five years of dedication and hard work, I have finally reached this milestone. As I begin writing my thesis, I am profoundly aware that this achievement would not have been possible without the support of many remarkable individuals. The years 2022 and 2023 were particularly difficult, marked by challenges that at times felt overwhelming. Yet it was the unwavering encouragement, patience, and love of my family, friends, and mentor that helped me navigate those trying times and continue my journey. I am deeply grateful to each of them for their guidance, belief, and support, which have made this accomplishment a reality.*

*First and foremost, I owe my deepest gratitude to my Ph.D. supervisor, **Prof. Shilpi Sharma**, whose guidance has been the cornerstone of this journey. Her expertise has expanded my knowledge and profoundly shaped my research instincts. Beyond her academic brilliance, what amazes me most is her extraordinary strength, she is truly one of the strongest people I have ever known. Throughout my Ph.D., she has been a perfect blend of rigor and patience, guiding me with firmness when needed and offering understanding and encouragement when times were tough. She has been there for me through both professional challenges and personal struggles, sometimes scolding me like a mentor, and other times cheering me on like a friend. Her insightful feedback, unwavering support, and boundless wisdom have not only helped me grow as a researcher but also inspired me to become a better version of myself. I am profoundly grateful for her guidance, motivation, and for the strength she has shown me at every step of this journey.*

*I am grateful to the members of my SRC, **Prof. Lucinda E. Doyle**, **Prof. Preeti Srivastava**, and **Prof. P. Hariprasad**, for their constructive criticism, and encouragement, all of which greatly contributed to improving my work. Whenever I sought guidance on my research experiments, they generously offered their time and provided thoughtful suggestions, patiently meeting with me on numerous occasions. Their support and expertise were instrumental in shaping the progress of my research. I sincerely thank **Dr. Y. S. Shivay** (ICAR–IARI, New Delhi) and **Dr. Ankita Sarkar** (BHU, Varanasi) for kindly providing soil samples from their respective institutions for use in this study. Their valuable support and cooperation are gratefully acknowledged. I would also like to thank **Dr. Avinash Sharma** (NCCS, Pune) and his group especially **Yukti Oza**, **Namrata Jiya** and **Bhavuk Gupta** for helping us with the analysis of sequencing data generated in this study.*

*During my Ph.D. journey, I was fortunate to receive two fellowships that profoundly enriched both my research and personal experiences. The first was the Jeff Schell Fellowship in Agricultural Sciences, funded by the Bayer Foundation, which allowed me to spend six months in Germany under the guidance of **Prof. Rita Grosch**. This period was unforgettable, not only did I advance my research, but I also*

*immersed myself in European research culture, savoured local cuisines, and experienced German traditions. Prof. Grosch's warmth and guidance made me feel at home, and the two technicians, **Gundula Aust** and **Andrea Maikath**, were incredibly supportive, assisting me with experiments while creating a friendly, welcoming environment. I am also deeply grateful to my friends **Shristee Panthee**, **Dr. Abigail Moreno Pedraza**, **Dr. Monica Barman**, **Kalyan Pinninti**, and **Cheng-Han Hsieh** for making my stay in Germany so comforting. Their kindness, motivation, and companionship turned this academic journey into a memorable and heartwarming experience. Later, I had the privilege of receiving a scholarship from the University of Fribourg in Switzerland, where I spent five months conducting research under **Prof. Laure Weisskopf**. My experiments and research analyses were primarily guided by **Sébastien Bruisson**, from whom I learned immensely. Meanwhile, **Ola Abdelrahman** became like a true friend, offering support, encouragement, and companionship beyond the lab. I sincerely thank **Ms. Sandra Nast** for her guidance and support, especially in helping me navigate initial challenges at the municipality. Her assistance ensured a smooth stay and quick adaptation to life abroad. These experiences gave me invaluable insights into professionalism, collaboration, and the spirit of scientific inquiry, while also allowing me to build meaningful friendships. I am deeply grateful for these opportunities, the knowledge I gained, and the lasting impressions they left on both my personal and professional growth.*

*I am deeply grateful to my friends and colleagues whose support and camaraderie have been invaluable throughout my Ph.D. journey. **Salila Pradhan**, my friend and lab colleague, and **Ajay Yadav**, friend from another lab, have been constant sources of encouragement and cheer, making even the toughest days brighter. I am equally thankful to my lab mates, **Annapurna Ma'am**, **Swati Ma'am**, **Shivani Ma'am**, **Priyanka Ma'am**, **Shubham Sir**, **Sonal Ma'am**, **Vijal Ma'am**, **Priya**, **Kanika**, **Soumya**, **Argha**, **Tannu**, **Nischay**, **Himanshu**, **Vaibhav**, **Mathangi** and **Puja**, whose friendship, understanding, and light-heartedness provided both motivation and laughter when the weight of research felt heavy. I am deeply indebted to **Pratibha** for initiating this work and achieving promising results. Her sincere efforts and collaboration greatly facilitated the continuation of this research. A special note of gratitude goes to **Sonal Ma'am**, who has been a pillar of support in every sense, guiding me professionally, standing by me personally, and becoming a true friend along the way. **Shivani Ma'am**, **Vijal Ma'am**, and **Shubham Sir** also offered invaluable guidance, helping me navigate the challenges of research with patience. I am thankful to **Annapurna Ma'am** for our long, thought-provoking scientific discussions that expanded my perspective. I would also like to thank **Amitava Sir**, who, though visiting occasionally, shared valuable advice, guidance, and a delightful sense of humor, making every interaction memorable. I would also like to express my sincere appreciation to my interns, **Tuhin**, **Shantanu**, **Avani**, and **Harshita** for their valuable assistance and thoughtful contributions to various aspects of this research, each bringing unique perspectives and dedication during their respective tenures.*

*I would like to sincerely thank the **Department of Biochemical Engineering and Biotechnology** at the **Indian Institute of Technology Delhi** for providing me with excellent facilities and a supportive environment to carry out my Ph.D. research. I am also deeply grateful to the **Department of Science and Technology (DST)** for funding my fellowship in India, as well as to the **Bayer Foundation (Leverkusen, Germany)** and the **University of Fribourg (Fribourg, Switzerland)** for providing financial support that enabled my international research visits.*

*Each of you I have mentioned has inspired me to persevere, push beyond my limits, and pursue knowledge with determination. Your belief in me reminded me of my potential, even in moments of self-doubt. As I move forward in my career, I carry the lessons, memories, and connections I have gained, which will continue to shape my journey. I am truly grateful and humbled to have learned from such remarkable individuals.*

*Finally, my deepest gratitude goes to my beloved **grandparents (Sh. Om Prakash Tyagi, Smt. Omvati Tyagi, Late Sh. Shive Nandan Tyagi, Smt. Magan Tyagi)**, **parents (Sh. Vipin Kumar Tyagi and Smt. Seema Tyagi)**, and **sister (Pooja Tyagi)**. Growing up in a society where daughters are often not encouraged and married off early, I was fortunate to have a family that showed immense patience, trust, and unwavering support. Because of their belief in me, I was able to pursue opportunities that took me to two different foreign countries for a total of 11 months. Living in places where I did not know the language, was not easy, but the confidence instilled by my parents allowed me to manage and adapt. My father's patience, my mother's constant encouragement, and my sister's love and thoughtful, logical advice have been my greatest strengths. I am also deeply grateful to my grandparents, whose wisdom and warmth created a nurturing and inspiring environment for our whole family. My grandfathers' love and lifelong passion for education greatly influenced my decision to pursue higher studies, and their constant encouragement towards learning has been a guiding force throughout my journey. Their support in every situation, celebrating even my smallest successes and reminding me of the value of each step along the way has shaped who I am today. Without their encouragement and guidance, reaching this milestone would not have been possible. Their support has motivated me to work diligently toward my goals, striving for a secure and fulfilling future while reminding me to remain confident yet humble.*

*I sincerely thank everyone once again, from the bottom of my heart, for their continuous support, guidance, and friendship. I feel truly blessed to have had all of you by my side throughout this transformative Ph.D. journey. Above all, I thank God for guiding me through challenges and ensuring that my efforts bear fruit.*

*Rashi Tyagi*

## ABSTRACT

Conventional approaches to manage phytopathogens are associated with ecological drawbacks; chemical fertilizers pollute the environment, whereas bioinoculants often display inconsistent performance under field conditions. *Fusarium* is a notorious fungal pathogen causing a variety of diseases in different agricultural crops. Also, in soil ecosystems, environmental factors and pathogen pressure can disturb the rhizospheric microbiome, influencing plant fitness. A pathobiome comprises of microbial members that interact with both host plant and pathogen to promote disease establishment. In this study, the microbial diversity of rhizospheric soil from two varieties of *Cajanus cajan* (pigeonpea) grown in two agroclimatic zones were profiled under *Fusarium udum* infested and pathogen free control conditions. The core bacterial genera identified included *Steroidobacter*, *Pseudomonas*, and *Acidibacter*, while *Mortierella*, *Actinomortierella*, and *Naganishia* dominated the fungal core. A *Fusarium*-induced disruption of the rhizosphere microbiome was identified, leading to the characterization of a distinct pathobiome. While certain genera of pathobiome such as *Acidibacter* and *Actinomortierella* overlapped with the core microbiome, others including *Brevundimonas*, *Calenema*, *Delftia* and fungal taxa like *Aplosporella*, *Cristinia*, and *Kwoniella* were unique to *Fusarium*-infested soil samples. Correlation analysis revealed a negative association between *Bacillus* and *Fusarium*, prompting the generation of a *Bacillaceae* culture bank with strains exhibiting antagonism against *F. udum* both *in vitro* and *in planta*.

Building on this, a microbiome-assisted rhizosphere engineering approach was adopted to combat *Fusarium* wilt in *C. cajan*. Indigenous *Bacillaceae* strains from the culture bank with biocontrol and plant growth-promoting (PGP) traits were strategically assembled into synthetic microbial communities (SMCs). Compatibility testing, qualitative profiling for exhibition of several traits and then using a novel approach of iterative deconvolution to select strains exhibiting enhanced production of biocontrol traits when present in a community of other

bacterial strains aided the selection of bacterial strains for generation of several SMCs that were tested through *in vitro* and *in planta* assays. Evaluation of growth attributes and stress markers identified one SMC with superior disease suppression potential, which was further validated under natural conditions. Persistence and colonization patterns of the five member strains of the shortlisted SMC were also confirmed in different root regions through scanning electron microscopy and viable cell counting methods. Further, whole genome sequencing of these five constituent strains revealed multiple potential biosynthetic gene clusters (BGCs) that might be linked to biocontrol activity.

Mechanistic insights into wilt suppression were gained by investigating the bacterial volatilome. Volatile organic compounds (VOCs) released by biocontrol strains demonstrated both direct antifungal activity and indirect effects via plant immunity. Volatile-mediated interactions between *F. udum* and bacterial strains were characterized, revealing pathogen-induced upregulation of specific bacterial VOCs. The application of some synthetic volatiles effectively suppressed Fusarium wilt under controlled conditions *in planta*. Beyond volatiles, diffusible metabolites were also examined; cell-free supernatants (CFS) of biocontrol strains reduced disease severity in *C. cajan*. Subsequent metabolomic profiling identified several bioactive compounds, and synthetic analogs of these metabolites were confirmed to suppress Fusarium wilt in *C. cajan*.

Overall, this work provides proof-of-concept for the pathobiome in plants and demonstrates microbiome-assisted rhizosphere engineering as a viable strategy for sustainable disease management. By leveraging VOCs and diffusible metabolites from indigenous *Bacillaceae* strains, this study establishes an effective alternative to environmentally damaging chemical pesticides and unreliable conventional bioinoculants.

## सार

फाइटोपैथोजेन्स (Phytopathogens) के प्रबंधन के पारंपरिक तरीके पारिस्थितिक कमियों से जुड़े हैं; रासायनिक उर्वरक पर्यावरण को प्रदूषित करते हैं, जबकि बायोइनोक्यूलेंट्स (bioinoculants) अक्सर क्षेत्र की स्थितियों के तहत असंगत प्रदर्शन प्रदर्शित करते हैं। फ्यूजेरियम (*Fusarium*) एक कुख्यात फंगल (fungal) रोगाणु है जो विभिन्न कृषि फसलों में कई तरह की बीमारियों का कारण बनता है। इसके अलावा, मिट्टी के पारिस्थितिक तंत्रों में, पर्यावरणीय कारक और रोगाणु दबाव राइजोस्फेरिक माइक्रोबायोम (rhizospheric microbiome) पर प्रतिकूल प्रभाव डाल सकते हैं, जिससे पौधे की फिटनेस (fitness) प्रभावित होती है। एक पैथोबायोम (pathobiome) में सूक्ष्मजीवी (microbial) सदस्य शामिल होते हैं जो रोग की स्थापना को बढ़ावा देने के लिए पौधे और रोगाणु दोनों के साथ वार्तालाप करते हैं। इस अध्ययन में, दो कृषि-जलवायु क्षेत्रों में उगाए गए अरहर (*Cajanus cajan*) की दो किस्मों से राइजोस्फेरिक (rhizospheric) मिट्टी की सूक्ष्मजीवी (microbial) विविधता को फ्यूजेरियम उदुम (*Fusarium udum*) संक्रमित और रोगाणु मुक्त स्थितियों के तहत प्रोफाइल (profile) किया गया था। फ्यूजेरियम (*Fusarium*)-प्रेरित राइजोस्फीयर माइक्रोबायोम (rhizosphere microbiome) में व्यवधान की पहचान की गई, जिससे एक विशिष्ट पैथोबायोम (pathobiome) का निर्धारण हुआ। पैथोबायोम की कुछ प्रजातियाँ, जैसे एसिडीबैक्टर (*Acidibacter*) और एक्टिनोमॉर्टिरेला (*Actinomortierella*), कोर माइक्रोबायोम (core microbiome) के साथ अतिव्याप्त थीं, जबकि ब्रेवुंडिमोनस, कैलेनेमा, डेल्टिया (*Brevundimonas*, *Calenema*, *Delftia*) और एप्लोस्पॉरेला, क्रिस्टिनिया और क्वोनिेला (*Aplosporella*, *Cristinia*, and *Kwoniella*) जैसे कवक वर्ग, फ्यूजेरियम (*Fusarium*)-संक्रमित मृदा नमूनों के लिए विशिष्ट थे। सहसंबंध विश्लेषण से बैसिलस (*Bacillus*) और फ्यूजेरियम (*Fusarium*) के बीच एक नकारात्मक संबंध का पता चला, जिससे एक बैसिलेसी (*Bacillaceae*) कल्चर बैंक (culture bank) का निर्माण हुआ, जिसमें ऐसे उपभेद थे जो इन विट्रो और इन प्लांटा (in plants) दोनों में एफ. उदुम (*F. udum*) के प्रति विरोध प्रदर्शित करते थे।

इसके आधार पर, अरहर में फ्यूजेरियम विल्ट (Fusarium wilt) से निपटने के लिए एक माइक्रोबायोम (microbiome)-सहायता प्राप्त राइजोस्फीयर इंजीनियरिंग (rhizosphere engineering) दृष्टिकोण अपनाया गया। कल्चर बैंक (culture bank) से जैव-नियंत्रण और पादप वृद्धि-प्रवर्तक (पीजीपी) गुणों वाले स्वदेशी बैसिलेसी (*Bacillaceae*) उपभेदों को रणनीतिक रूप से सिंथेटिक माइक्रोबियल (synthetic microbial) समुदायों (एसएमसी, SMC) में एकत्रित किया गया। संगतता परीक्षण, कई लक्षणों के प्रदर्शन के लिए गुणात्मक रूपरेखा तैयार करना और फिर अन्य जीवाणु उपभेदों के समुदाय में मौजूद होने पर जैवनियंत्रण लक्षणों के उन्नत उत्पादन को प्रदर्शित करने वाले उपभेदों का चयन करने के लिए पुनरावृत्त विसंवर्तन के एक नए दृष्टिकोण का उपयोग करने से कई एसएमसी की पीढ़ी के लिए जीवाणु उपभेदों के चयन में सहायता मिली, जिनका परीक्षण इन विट्रो (*in vitro*) और इन प्लांटा (*in planta*) परख के माध्यम से किया गया। विकास विशेषताओं और तनाव मार्करों के मूल्यांकन ने बेहतर रोग दमन क्षमता वाले एक एसएमसी की पहचान की, जिसे प्राकृतिक परिस्थितियों में आगे मान्य किया गया। स्कैनिंग इलेक्ट्रॉन माइक्रोस्कोपी (Scanning Electron Microscopy) और व्यवहार्य कोशिका गणना विधियों के माध्यम से विभिन्न जड़ क्षेत्रों में चयन किए गए एसएमसी (SMC) के पांच सदस्य उपभेदों की दृढ़ता और उपनिवेशण पैटर्न की भी पुष्टि की गई। इसके अतिरिक्त, इन पाँच घटक उपभेदों के संपूर्ण जीनोम अनुक्रमण से कई संभावित जैवसंश्लेषी जीन क्लस्टर सामने आए, जो जैव-नियंत्रण गतिविधि से जुड़े हो सकते हैं।

एफ. उदुम (*F. udum*) और जीवाणु उपभेदों के बीच वाष्पशील (Volatile organic compounds)-मध्यस्थ अंतःक्रियाओं का वर्णन किया गया, जिससे विशिष्ट जीवाणु के रोगजनक-प्रेरित अपरेगुलेशन का पता चला। कुछ सिंथेटिक वाष्पशील पदार्थों के अनुप्रयोग ने पौधों में नियंत्रित परिस्थितियों में फ्यूजेरियम विल्ट को प्रभावी ढंग से दबा दिया। वाष्पशील पदार्थों के अलावा, विसरित मेटाबोलाइट्स (metabolites) की भी जाँच की गई; जैवनियंत्रण उपभेदों के कोशिका-रहित सुपरनेटेंट्स ने अरहर में रोग की गंभीरता को कम किया। बाद में मेटाबोलोमिक प्रोफाइलिंग (metabolomic profiling) ने कई जैवसक्रिय

यौगिकों की पहचान की, और इन मेटाबोलाइट्स (metabolites) के सिंथेटिक एनालॉग्स (synthetic analogues) की पुष्टि अरहर में फ्यूजेरियम विल्ट (Fusarium wilt) को दबाने के लिए की गई।

कुल मिलाकर, यह कार्य पौधों में पैथोबायोम (pathobiome) के लिए अवधारणा का प्रमाण प्रदान करता है और स्थायी रोग प्रबंधन के लिए एक व्यवहार्य रणनीति के रूप में माइक्रोबायोम-सहायता प्राप्त राइजोस्फीयर इंजीनियरिंग को प्रदर्शित करता है। स्वदेशी बैसिलेसी उपभेदों से VOCs और विसरित मेटाबोलाइट्स (metabolites) का लाभ उठाकर, यह अध्ययन पर्यावरण को नुकसान पहुँचाने वाले रासायनिक कीटनाशकों और अविश्वसनीय पारंपरिक बायोइनोक्युलेंट्स (bioinoculants) का एक प्रभावी विकल्प स्थापित करता है।

<b>LIST OF CONTENTS</b>	
<b>TOPIC</b>	<b>Page No.</b>
<b>CERTIFICATE</b>	<b>i</b>
<b>ACKNOWLEDGEMENT</b>	<b>ii-iv</b>
<b>ABSTRACT</b>	<b>v-ix</b>
<b>LIST OF FIGURES</b>	<b>xvi-xx</b>
<b>LIST OF TABLES</b>	<b>xxi</b>
<b>ABBREVIATIONS</b>	<b>xxii-xxiv</b>
<b>1. INTRODUCTION AND OBJECTIVES</b>	<b>1-10</b>
<b>1.1. Introduction</b>	<b>2-6</b>
1.1.1. Stresses in agriculture	2-3
1.1.2. Limitations of current approaches to mitigate biotic stress	3-4
1.1.3. Rhizosphere engineering	4-5
<i>1.1.3.1. Top-down approach of rhizosphere engineering: In situ tailoring of rhizobiome</i>	4
<i>1.1.3.2. Bottom-up approach of rhizosphere engineering: Generation of Synthetic Microbial Community (SynCom/SMC)</i>	4-5
1.1.4. Model systems used in this study	5-6
<i>1.1.4.1. Fusarium as biotic stress in agriculture</i>	5
<i>1.1.4.2. Cajanus cajan as model crop system</i>	5-6
<b>1.2. Objectives</b>	<b>7-10</b>
<b>2. REVIEW OF LITERATURE</b>	<b>11-39</b>
<b>2.1. Environmental stresses affecting plants</b>	<b>12-13</b>
2.1.1. Abiotic stresses	12
2.1.2. Biotic stresses	12-13
<b>2.2. Rhizosphere: A hotspot for microbial diversity</b>	<b>13-16</b>
<b>2.3. Pathobiome</b>	<b>16-18</b>
<b>2.4. Fusarium: A notorious pathogen</b>	<b>18-19</b>
<b>2.5. Microbiome dynamics in the rhizosphere upon Fusarium infestation</b>	<b>19-20</b>
<b>2.6. Response of plants to Fusarium infection</b>	<b>20-22</b>
<b>2.7. Cajanus cajan: An important food legume</b>	<b>22-25</b>

2.7.1. Rhizobiome associated with <i>C. cajan</i>	22-24
2.7.2. Implications of <i>Fusarium</i> on <i>C. cajan</i>	24-25
<b>2.8. Traditional approaches to manage Fusarium wilt in <i>C. cajan</i></b>	<b>26-29</b>
2.8.1. Cultural management	26
2.8.2. Chemical management	26-27
2.8.3. Biological management	27-29
<b>2.9. Rhizosphere engineering to combat biotic stress</b>	<b>30-34</b>
2.9.1. Top-down approach	30
2.9.2. Bottom-up approach	30-34
<b>2.10. Screening for generation of SMC</b>	<b>34-38</b>
2.10.1. Microdroplet screening method	35
2.10.2. Artificial intelligence and machine learning in development of SMC	35
2.10.3. Deconvolution techniques	35-37
2.10.4. Neural networks	37-38
<b>2.11. Research gaps in the area</b>	<b>38-39</b>
<b>3. MATERIALS AND METHODS</b>	<b>40-65</b>
<b>3.1. Soil sampling to identify changes in microbiome of <i>C. cajan</i> upon <i>Fusarium</i> challenge</b>	<b>41-44</b>
3.1.1. DNA extraction and amplicon sequencing	43
3.1.2. Sequencing data analysis	43-44
<b>3.2. Development of a plant-pathogen disease model to evaluate fungal pathogenicity under controlled and natural conditions</b>	<b>44-45</b>
3.2.1. Soil drench	44-45
3.2.2. Root treatment	45
3.2.3. Sick soil	45
<b>3.3. Generation of culture bank of bacterial isolates with biocontrol activity</b>	<b>45-48</b>
3.3.1. Biocontrol activity of bacterial isolates and their cell-free supernatant (CFS) <i>in vitro</i>	46-47
3.3.2. Phylogenetic affiliation of antagonistic strains	47
3.3.3. Wilt protection ability of isolates <i>in planta</i>	47-48

3.3.4. Impact of bacterial bioformulations on plant growth attributes and stress markers	48
<b>3.4. Cross streak assay to check for inhibition between strains</b>	<b>49</b>
<b>3.5. Qualitative profiling of biocontrol and PGP properties shown by bacterial isolates</b>	<b>49</b>
<b>3.6. Quantitative analysis of biocontrol properties expressed by strains using iterative deconvolution</b>	<b>50-52</b>
3.6.1. IAA quantification	50
3.6.2. HCN quantification	50-51
3.6.3. Amylase estimation	51
3.6.4. Chitinase estimation	51
3.6.5. Cellulase estimation	51-52
3.6.6. Siderophore estimation	52
<b>3.7. Generation of SMC</b>	<b>52</b>
<b>3.8. Checking the root colonization ability of selected strains</b>	<b>52-53</b>
<b>3.9. Testing the disease protection ability of generated SMC (/s)</b>	<b>53-54</b>
3.9.1. Seed germination assay	53-54
3.9.2. Evaluation of disease suppression and PGP activity under controlled conditions <i>in planta</i>	54
<b>3.10. Quantitative assessment of stress markers</b>	<b>54-56</b>
3.10.1. Proline quantification	54-55
3.10.2. Measurement of total phenolics	55
3.10.3. Quantification of antioxidant enzymes	55-56
<b>3.11. Validation of suppression of Fusarium wilt and PGP activity of SMC under natural conditions</b>	<b>56-57</b>
<b>3.12. Tracking of SMC and member strains <i>in planta</i> under controlled conditions in a hydroponics setup</b>	<b>57</b>
<b>3.13. Whole genome sequencing of the five bacterial strains constituting the shortlisted SMC</b>	<b>57-58</b>
<b>3.14. Volatile-mediated management of Fusarium wilt in <i>C. cajan</i></b>	<b>58-62</b>
3.14.1. Testing the antagonistic activity of volatiles released by biocontrol strains against <i>F. udum</i> <i>in planta</i>	58-59

3.14.1.1. <i>Co-inoculation of plants with test pathogen and bacterial volatiles</i>	59
3.14.1.2. <i>Priming of seedlings with volatiles prior to pathogen inoculation</i>	59
3.14.2. Setup to capture volatiles released by individual strains using single microcosm	59-60
3.14.3. Setup to study volatile-mediated interaction between two microorganisms using connected microcosms	60
3.14.4. Conditions for gas Chromatography/Quadrupole time of flight (GC/QTOF) and analysis	60-61
3.14.5. Checking the inhibition of <i>F. udum</i> by selected synthetic volatile compounds <i>in planta</i>	61-62
<b>3.15. Role of bacterial metabolites in disease protection against <i>F. udum</i></b>	<b>62-64</b>
3.15.1. <i>In planta</i> disease protection ability of bacterial CFS	62-63
3.15.2. Extraction of bioactive diffusible metabolites	63
3.15.3. High resolution liquid chromatography electrospray ionization mass spectrometer (HR-LC-ESI-MS)	63-64
3.15.4. Management of Fusarium wilt using synthetic metabolites	64
<b>3.16. Statistical analysis</b>	<b>64-65</b>
<b>4. RESULTS</b>	<b>66-140</b>
<b>4.1. Infestation with <i>F. udum</i> triggered differential response in <i>C. cajan</i> varieties</b>	<b>67-73</b>
4.1.1. Variation in physicochemical parameters for different soil samples	67-68
4.1.2. Influence of test pathogen on the microbial diversity in rhizosphere of <i>C. cajan</i>	68-71
4.1.3. Composition of microbial communities	72-73
<b>4.2. Identification of core microbiome associated with <i>C. cajan</i></b>	<b>73-74</b>
<b>4.3. Microbial dysbiosis and pathobiome associated with Fusarium wilt of <i>C. cajan</i></b>	<b>74-80</b>
<b>4.4. Generation of culture bank of native bacterial strains antagonistic to <i>F. udum</i></b>	<b>80-82</b>

<b>4.5. Validation of disease protection ability of isolates in host plant</b>	<b>83-85</b>
<b>4.6. Assessment of compatibility and exhibition of biocontrol and PGP traits by the strains</b>	<b>85-88</b>
<b>4.7. Quantitative estimation of biocontrol traits through iterative deconvolution</b>	<b>89-92</b>
<b>4.8. Scoring of strains for constitution of SMCs and checking root colonization ability of selected strains</b>	<b>92-97</b>
<b>4.9. Assessing the biocontrol and PGP activity of SMCs <i>in vitro</i> and <i>in planta</i> under controlled conditions</b>	<b>98-103</b>
<b>4.10. Confirmation of the biocontrol and PGP activity of SMCs <i>in planta</i> under natural settings</b>	<b>104-109</b>
<b>4.11. Checking the persistence and survivability of SMC and individual strains in the roots of <i>C. cajan</i> in hydroponics</b>	<b>109-111</b>
<b>4.12. Analysis of whole genome sequences and metabolite clusters of individual bacterial strains involved in SMC</b>	<b>111-115</b>
<b>4.13. Volatile mediated biocontrol potential of individual strains</b>	<b>115-131</b>
4.13.1. Wilt protection potential of bacterial volatiles <i>in planta</i>	116
4.13.2. Exposure of the biocontrol strains to the volatiles of <i>F. udum</i> triggered increased emission of several volatile compounds	117-125
4.13.3. Biological activity of synthetic volatile compounds <i>in planta</i>	126-127
4.13.4. Impact of synthetic volatiles on stress markers and antioxidant enzymes	127-131
<b>4.14. Diffusible metabolite based antifungal ability of individual strains</b>	<b>131-140</b>
4.14.1. Biocontrol potential of bacterial metabolome <i>in planta</i>	131-135
4.14.2. Identification of bioactive diffusible metabolites released by bacterial strains	135-137
4.14.3. Wilt protection through synthetic diffusible metabolites	137-140
<b>5. DISCUSSION</b>	<b>141-162</b>
<b>5.1. Characterization of microbiome associated with pathogen-free and <i>Fusarium</i>-infested rhizosphere of <i>C. cajan</i></b>	<b>142-148</b>
<b>5.2. Rhizosphere engineering to control <i>Fusarium</i> wilt of <i>C. cajan</i> under controlled and natural conditions</b>	<b>148-156</b>

<b>5.3. Mechanism of fungal inhibition exhibited by individual biocontrol strains</b>	<b>156-162</b>
<b>6. SUMMARY AND CONCLUSIONS</b>	<b>163-167</b>
<b>6.1. Summary</b>	<b>164-165</b>
<b>6.2. Conclusions</b>	<b>166-167</b>
Bibliography	168-187
Appendix	188-192
CURRICULUM VITAE	193-194

## LIST OF FIGURES

Figure No.	Legend	Page No.
2.1	Rhizosphere with different zones demarcated - the endorhizosphere, the rhizoplane and the ectorhizosphere (McNear, 2013)	14
2.2	A framework for tailoring stable and effective synthetic microbial communities (SMCs) to enhance crop resiliency to environmental stresses (de Souza, 2020)	31
2.3	Pictorial representation of the approach of iterative deconvolution (created using Biorender.com)	37
3.1	Schematic representation of the experiments conducted to characterize the changes in microbiome of <i>C. cajan</i> upon <i>Fusarium</i> infestation (Objective 1) then, to design SMCs (Objective 2). Next, to check the disease protection ability of SMCs <i>in planta</i> (Objective 3), and understanding the pathogen suppression mechanism of individual strains (Objective 4)	42
4.1	Alpha-diversity of microbial communities in rhizosphere soil collected from <i>Fusarium</i> -infested and pathogen-free fields at two agroclimatic zones, (a) bacterial communities, and (b) fungal communities (Refer to Table 3.1 for treatment details)	69-70
4.2	PCoA plot depicting beta-diversity of microbial communities in rhizosphere soil collected from <i>Fusarium</i> -infested and pathogen-free fields at two agroclimatic zones, (a) bacterial community, and (b) fungal community (Refer to Table 3.1 for the treatment details)	71
4.3	Relative abundance of top bacterial (a), and fungal genera (b), observed in proportion of all sequences detected in <i>Fusarium</i> -infested and pathogen-free soil collected from different <i>C. cajan</i> varieties grown in the two agroclimatic zones	72-73
4.4	Core microbiome of <i>C. cajan</i> , bacterial (a), and fungal (b) communities	74
4.5	Shared and unique ASVs for pathogen-free and <i>Fusarium</i> -infested rhizosphere soil of Bahar collected from IARI, Delhi, (a) bacterial ASVs, and (b) fungal ASVs	75
4.6	Shared and unique ASVs for pathogen-free and <i>Fusarium</i> -infested rhizosphere soil of Bahar collected from both agroclimatic zones (Delhi and Varanasi), (a) bacterial ASVs, and (b) fungal ASVs	76
4.7	Shared and unique ASVs for pathogen-free and <i>Fusarium</i> -infested rhizospheric soil of two varieties, Bahar and C11 collected from Varanasi, (a) bacterial ASVs, and (b) fungal ASVs	77
4.8	Shared ASVs between <i>F. udum</i> -infested rhizosphere soil samples collected from different agroclimatic zones, Delhi and Varanasi, (a) bacterial ASVs, and (b) fungal ASVs	78

4.9	Correlation plot presenting interactions between microbial communities present in pathogen-free and <i>Fusarium</i> -infested rhizospheric soil samples from both agroclimatic zones: (a) between bacteria-fungi, (b) between fungi-fungi	79-80
4.10	<i>In vitro</i> assays for biocontrol potential of bacterial strains, (a) Representation of dual culture assay with potent isolates antagonistic to test pathogen; Bar graphs showing percent growth inhibition of mycelia of <i>F. udum</i> by different bacterial culture (b), and their CFS (c). Error bar shows standard deviation ( $n=3$ ), different letters indicate statistical differences	81
4.11	Plant growth experiment (a) Plot depicting antagonistic potential of all bacterial strains against test pathogen, <i>F. udum</i> ; Effect of seed pelleting on plant growth attributes including root length (b), total chlorophyll (c), and carotenoids (d) and, lipid peroxidation (e). Error bar represents standard deviation where $n=3$ , different letters express significant differences ( $p < 0.05$ )	84-85
4.12	Assessment of compatibility between bacterial isolates through dual culture assay on nutrient agar	86
4.13	Qualitative assays for biocontrol and PGP traits exhibited by strains	86
4.14	Deconvolution for production of metabolites responsible for biocontrol: (a) siderophore, (b) amylase, (c) cellulase, (d) IAA, (e) HCN and, (f) chitinase. Error bars represent standard deviation where $n=3$ . Blue bars represent activity of strains in isolation, while orange shows the strains in a community of non-producer strains. Statistical differences are denoted using paired t-test with ( $p < 0.05$ ) as no significant difference	90-92
4.15	Scanning electron micrograph showing adherence of bacterial strains occupying different parts of root system of <i>C. cajan</i> : (a) Control (without any bacterial amendment), (b) Ph7, (c) DC12, and (d) DI15	96-97
4.16	Seed germination assay with plot depicting the average vigour index for seeds treated with different bioformulations both in the absence and presence of fungal pathogen. Error bar represents standard deviation; different letters designate statistical differences	98
4.17	<i>In planta</i> assay for suppression of Fusarium wilt: Plot showing the percent (%) disease incidence observed in plants treated with different bioformulations. Error bar represents standard deviation where $n=3$ ; statistical differences are expressed with $p < 0.05$ as no significant difference	99
4.18	Heatmap for plant growth attributes, (a) root length, (b) dry weight, and (c) total chlorophyll content (created using GraphPad Prism 8.0.2)	101

4.19	Heatmap showing comparison of production of stress markers and antioxidant enzyme activities, (a) lipid peroxidation, (b) proline, (c) SOD, and (d) PAL (created using GraphPad Prism 8.0.2)	102
4.20	Principal component analysis differentiating bioformulations of individual strains and SMCs based on different plant growth attributes and stress markers (created using PAST 4.03)	103
4.21	<i>In planta</i> assay for determination of biocontrol and plant growth promoting property of SMC and its member strains at two developmental stages, flowering and harvest in <i>C. cajan</i> at natural conditions, (a) Disease severity score, (b) Carotenoids, (c) Lipid peroxidation, (d) Proline, and (e) Phenolics. Error bar represents standard deviation where $n=5$ ; statistical differences are expressed with $p < 0.05$ as no significant difference. Different lowercase letters indicate statistically significant differences at flowering stage, while uppercase letters indicate significant differences among treatments at harvest	105-106
4.22	Determination of antioxidant enzymes for the <i>in planta</i> assay, (a) SOD, (b) GPX, (c) APX, and (d) PPO. The blue bars represent flowering while orange denotes harvest stage. Error bar represents standard deviation where $n=5$ ; statistical differences are expressed with $p < 0.05$ as no significant difference. Different lowercase letters indicate statistically significant differences at flowering stage, while uppercase letters indicate significant differences among treatments at harvest	108-109
4.23	Comparison of survivability of individual bacterial strain and in SMC over a time period across different root sections, base, mid and root tip. Graphs represent viability patterns for (a) individual strains, (b) in SMC. The graph was generated in R studio using 3-way ANOVA with Tukey HSD for statistical comparison	110
4.24	Genome-based phylogenetic tree of all the five shortlisted bacterial strains generated using the Type strain genome server (TYGS), inferred with GBDP. The branch lengths represent genome-to-genome distances (genomic divergence) between strains, while the numbers at nodes indicate bootstrap support values reflecting the confidence in each branching	112
4.25	Predicted biosynthetic gene clusters (BGCs) in all the five shortlisted bacterial strains identified using antiSMASH, (a) DI6, (b) DI15, (c) DI18, (d) DC12, and (e) Ph7	113-115
4.26	<i>In planta</i> biocontrol activity of volatiles released by bacterial strains under two treatment modalities, co-inoculation and priming. The plot shows the disease severity score observed in plants treated with volatilome of bacterial strains (differentiated by colours). Treatments are compared with their respective controls within each	116

	<p>modality using one-way ANOVA. Different lowercase letters indicate statistically significant differences among co-inoculation treatments (<math>p &lt; 0.05</math>), while uppercase letters indicate significant differences among priming treatments (<math>p &lt; 0.05</math>) based on Duncan post-hoc test. Error bars represent standard deviation where <math>n=6</math>. The experiment was performed with six biological replicates, each having three plants and each plant was scored separately</p>	
4.27	<p>Venn diagrams showing the distribution of the volatile organic compounds (VOCs) emitted by five biocontrol strains, (a) VOCs produced by the strains when grown individually in a single microcosm setup, and (b) VOCs emitted in higher amounts by the strains when exposed to volatiles released by the test pathogen, <i>F. udum</i>. The numeric values represent the compounds unique to individual strains as well as those shared among multiple strains. Each strain is represented by a different colour</p>	117-118
4.28	<p><i>In planta</i> study for the suppression of Fusarium wilt by pure synthetic volatiles, (a) Representative figure showing plants in the order, uninfected, infected, exposed to undecanal (Left to right), (b) Disease severity scores observed in plants exposed to different volatiles, and (c) Dry weight of plants exposed to different volatiles. Error bars represent standard deviation where <math>n=7</math> and each plant was scored and weighed separately; statistical differences were analysed through one-way ANOVA and different letters represent statistical differences (<math>p &lt; 0.05</math>) based on Duncan post-hoc test</p>	126-127
4.29	<p>Non-enzymatic stress markers exhibited by plants treated with different synthetic volatiles, (a) Lipid peroxidation, measured by quantifying malondialdehyde (MDA), (b) Proline, and (c) Total phenolics. Error bars represent standard deviation where <math>n=3</math> (test was conducted in triplicates using a composite sample of seven biological replicates); statistical differences were analysed through one-way ANOVA different letters represent statistical differences (<math>p &lt; 0.05</math>) based on Duncan post-hoc test</p>	128-129
4.30	<p>Antioxidant activities shown by plants subjected to various synthetic volatiles, (a) SOD, (b) APX, (c) GPX, and (d) PPO. Error bars represent standard deviation where <math>n=3</math> (test was conducted in triplicates using a composite sample of seven biological replicates); statistical differences were analysed through one-way ANOVA, the same letters express no significant difference (<math>p &lt; 0.05</math>) based on Duncan post-hoc test</p>	130-131
4.31	<p><i>In planta</i> study for the protection against Fusarium wilt by diffusible metabolites, (a) Representative figure showing plants in the order, uninfected, infected, inoculated with cell free supernatant of DC12 (Left to right), and (b) Disease severity scores observed in plants</p>	132

	inoculated with cell free supernatants of individual strains. Error bars represent standard deviation where $n=7$ and each plant was scored and weighed separately; statistical differences were analysed through one-way ANOVA and different letters represent statistical differences ( $p < 0.05$ ) based on Duncan post-hoc test	
4.32	Stress markers exhibited by plants treated with cell free supernatants of different bacterial strains, (a) Lipid peroxidation, (b) Proline, and (c) Total phenolics, (d) SOD, and (e) GPX. Error bars represent standard deviation where $n=3$ (test was conducted in triplicates using a composite sample of seven biological replicates); statistical differences were analysed through one-way ANOVA different letters represent statistical differences ( $p < 0.05$ ) based on Duncan post-hoc test	134-135
4.33	Differential analysis of diffusible metabolites identified through HR-LC-ESI-MS: (a) Bar graph showing number of unique metabolites produced by each strain, and (b) Heatmap representing the metabolites shared among the five bacterial strains	136-137
4.34	<i>In planta</i> assay to assess the biocontrol and plant growth promoting ability of synthetic metabolites, (a) Disease severity scores observed in plants treated with synthetic metabolites, and (b) Total chlorophyll content. Error bars represent standard deviation where $n=7$ ; statistical differences were analysed through one-way ANOVA and different letters represent statistical differences ( $p < 0.05$ ) based on Duncan post-hoc test	138
4.35	Antioxidant enzymes assessed in plants treated with different synthetic metabolites, (a) SOD, (b) PAL, and (c) GPX. Error bars represent standard deviation where $n=3$ (test was conducted in triplicates using a composite sample of seven biological replicates); statistical differences were analysed through one-way ANOVA, different letters represent statistical differences ( $p < 0.05$ ) based on Duncan post-hoc test	139-140
6.1	Summary of results obtained for each of the objectives in this study	165

## LIST OF TABLES

Table No.	Legend	Page No.
2.1	Microbial traits and their role in plant growth promotion and biocontrol	15-16
2.2	Common pesticides and biocontrol agents against <i>F. udum</i>	27-28
2.3	Synthetic microbial communities and microbial consortia to combat biotic stress	32-34
3.1	Codes for soil samples	41
3.2	Disease severity score for Fusarium wilt of <i>C. cajan</i>	48
4.1	Physicochemical profiles of rhizosphere samples	67-68
4.2	Accession numbers and phylogenetic affiliation of biocontrol strains	82
4.3	Exhibition of biocontrol and PGP traits by bacterial isolates	87-88
4.4	Scoring of strains on the basis of antagonism and iterative deconvolution	93-94
4.5	Composition of the generated SMCs	95
4.6	Genome details of the shortlisted five bacterial strains	111
4.7	Biosynthetic gene clusters (BGCs) in the genomes of bacterial strains	115
4.8	List of volatiles emitted by bacterial strains in the absence of VOCs released by <i>F. udum</i>	118-120
4.9	List of volatiles released by bacterial strains with increased abundance when exposed to volatiles from <i>F. udum</i>	122-125

## ABBREVIATIONS

ADAP	Automated data analysis pipeline
AI	Artificial intelligence
BGCs	Biosynthetic gene clusters
Bioc	Biocompost
CFS	Cell free supernatant
CFU	Colony forming unit
CLSA	Closed-loop stripping analysis
DAS	Days after sowing
EPS	Exopolysaccharides
FACS	Fluorescence-activated cell sorting
FYM	Farmyard manure
FW	Fresh weight
GBDP	Genome BLAST distance phylogeny
GC-QTOF	Gas chromatography/quadrupole time of flight
HR-LCMS-QTOF	High resolution liquid chromatography mass spectrometry / quadrupole time of flight
ITS	Internal transcribed spacer
ISR	Induced systemic resistance
LPX	Lipid peroxidation
MDA	Malondialdehyde
ML	Machine learning
NRPS	Non-ribosomal peptide synthetase
OD	Optical density
OTU	Operational taxonomic unit
PBS	Phosphate buffer saline
PCoA	Principal coordinates analysis
PDA	Potato dextrose agar
PGP	Plant growth promoting
PGPR	Plant growth promoting rhizobacteria
PTFE	Polytetrafluoroethylene
RI	Retention index
RiPP	Ribosomally synthesized and post-translationally modified peptide
SEM	Scanning electron microscopy
SMC/SynCom	Synthetic microbial community
SPME	Solid phase microextraction
TYGS	Type strain genome server
VOC	Volatile organic compound

### Elements

Carbon	C
Nitrogen	N
Phosphorus	P
Potassium	K

## Units

Cm	Centimetre
°C	Degrees Celsius
G	Gram
H	Hours
m s <sup>-1</sup>	Meter per second
μL	Microliter
μm	Micrometre
μM	Micromolar
Mg	Milligram
mL	Milliliter
mM	Millimolar
Min	Minutes
M	Molar
m/z	Mass/charge
Nm	Nanometre
%	Percent
rpm	Revolution per minute
S	Seconds

## Molecules and Chemicals

ACC	Aminocyclopropane-1-carboxylic acid
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	Ammonium sulfate
CaCl <sub>2</sub>	Calcium chloride
CMC	Carboxymethyl cellulose
CoCl <sub>2</sub>	Cobalt (II) chloride
CTAB	Cetyltrimethylammonium bromide
DCM	Dichloromethane
ddH <sub>2</sub> O	Double-distilled water
DNS	Dinitro-salicylic acid
EDTA	Ethylenediaminetetraacetic acid
FCP	Folin-Ciocalteu Phenol
FeSO <sub>4</sub> ·7H <sub>2</sub> O	Ferrous sulfate heptahydrate
H <sub>2</sub> O <sub>2</sub>	Hydrogen peroxide
HCN	Hydrogen cyanide
IAA	Indole-3-acetic acid
KH <sub>2</sub> PO <sub>4</sub>	Potassium dihydrogen phosphate
MgSO <sub>4</sub> ·7H <sub>2</sub> O	Magnesium sulfate heptahydrate
MnSO <sub>4</sub>	Manganese (II) sulfate
Na <sub>2</sub> CO <sub>3</sub>	Sodium carbonate
Na <sub>2</sub> HPO <sub>4</sub> ·7H <sub>2</sub> O	Disodium hydrogen phosphate heptahydrate
NaCl	Sodium chloride
NaOCl	Sodium hypochlorite
NBT	Nitrobluetetrazolium
PVP	Polyvinylpyrrolidone
TBA	Thiobarbituric acid

TCA	Trichloroacetic acid
ZnCl <sub>2</sub>	Zinc chloride