

**INFLUENCE OF SOLUTION PROPERTIES
ON ELECTROSPINNING BEHAVIOUR OF
BIOCOMPATIBLE POLYMERS**

DEEPIKA GUPTA



**DEPARTMENT OF TEXTILE TECHNOLOGY
INDIAN INSTITUTE OF TECHNOLOGY DELHI**

DECEMBER 2016

©Indian Institute of Technology Delhi (IITD), New Delhi, 2016

**INFLUENCE OF SOLUTION PROPERTIES
ON ELECTROSPINNING BEHAVIOUR OF
BIOCOMPATIBLE POLYMERS**

by

DEEPIKA GUPTA

Department of Textile Technology

Submitted

in fulfillment of the requirements of the degree of Doctor of Philosophy

to the



Indian Institute of Technology Delhi

December 2016

CERTIFICATE

This is to certify that the thesis entitled '**Influence of solution properties on electrospinning behaviour of biocompatible polymers**' being submitted by **Ms. Deepika Gupta** to the Indian Institute of Technology Delhi for the award of degree **Doctor of Philosophy**, is a record of bonafide research work carried out by her. She has worked under our guidance and supervision and fulfilled the requirements for the submission of thesis which has attained the standard required for a Ph.D. degree of this institute. The results contained in this thesis have not been submitted, in part or in full to any other university or institute for the awards of any degree or diploma.

Dr. Ashwini K. Agrawal

Professor

Department of Textile Technology

Indian Institute of Technology Delhi

Dr. Manjeet Jassal

Professor

Department of Textile Technology

Indian Institute of Technology Delhi

Date:

Place: New Delhi

ACKNOWLEDGEMENTS

I am sincerely grateful to my Ph.D. supervisors, Prof. Ashwini K. Agrawal and Prof. Manjeet Jassal for their relentless support, unremitting encouragement and guidance throughout the course of this research work. Their valuable suggestions, instructions and utter confidence in me all through the study have been instrumental in successful completion of the thesis. I am highly obliged to them for giving me the opportunity to work for SMITA Research Lab.

I am thankful to the Head, Department of Textile Technology for providing the infrastructure and other facilities to carry out my research work.

I would like to express my sincere thanks to my Student Research Committee (SRC) members - Prof. Veena Koul, Prof. Bhuvanesh Gupta and Dr. Sourabh Ghosh for their valuable time and suggestions during the course of this work.

I am also thankful to Dr. Sujata Mohanty, Stem Cell Facility, AIIMS, New Delhi, and her team for the collaboration and boundless support in carrying out the cell culture experiments.

I acknowledge the financial support by the Department of Science and Technology, Govt. of India towards my scholarship and research grants under various projects.

I am also grateful to all the technical staff of the Department of Textile Technology especially Mr. V. K. Kala, Mr. Amarjeet, Mr. A. K. Sehgal and Mr. D. C. Sharma for their spontaneous support.

I would like to thank my seniors particularly Dr. Dhirendra Singh, Dr. Sidhharth Sirohi, Dr. Sandip Basu, Dr. Prashant Panda, Dr. Bhavana Sharma, and lab members Ms. Kiran, Ms. Ratyakshi, Ms. Kamlesh and Mr. Raghav, Ms. Surabhi, Ms. Kumud and other former members of SMITA Research Lab for their assistance and encouragement.

I can never thank my family enough for their unquestionable support and impeccable trust that helped me sail through the ups and downs of this period and complete this endeavor.

Deepika Gupta

ABSTRACT

Nanofibres are indispensable for applications like tissue engineering, filtration, sensors etc., due to their unique properties. Electrospinning is an excellent technique for producing nanofibres from polymer solutions or melts using high electric field. Control of nanofibre morphology and quality are important to augment their utility for various applications. Spinnability and morphology of fibres are affected by solution, ambient and processing parameters.

Process parameters that include flow rate, spinning voltage and spinning distance were recently reported to not have any significant effect on the fibre diameter, if the spinning is carried out at minimum electrospinning voltage (MEV). It has been suggested that solution properties have a stronger influence on controlling the fibre diameter. However, the studies in this direction are limited. The rheological properties can be expressed in terms of viscosity and elasticity, which are affected by the type of polymer, its molecular weight, its concentration and the nature of solvent used.

To isolate the elastic effect from the viscous effect, Boger fluids of poly(vinyl alcohol) (PVA) were prepared by blending small amounts of high molecular weight PVA (HMW-PVA) with highly concentrated solutions of low molecular weight PVA (LMW-PVA). The resulting solutions with increase in HMW-PVA content showed increasing elasticity, while maintaining the same viscosity, thereby, separating the two effects. On electrospinning, the elasticity of the solution (expressed in terms of relaxation time (λ) and Deborah number (De)) was found to have a direct correlation with diameter of the electrospun fibres.

Nature of solvent is known to affect the solution rheology. Rheology and electrospinning behaviour of PVA in two different solvents, i.e. dimethyl sulfoxide

(DMSO) and water, was studied. The rheological properties for the same concentration of PVA were found to be significantly different in the two solvents. The viscosity, storage modulus and relaxation time (λ) were found to be higher in PVA-DMSO system. However, the fibre diameter obtained from PVA-DMSO system was lower than that from PVA-water system. Interestingly, still a linear correlation of the fibre diameter with λ could be established for the two solvents with two different slopes. On further investigation, a single linear equation could be drawn for the two systems between diameter and the number of entanglements per chain suggesting elasticity arising from chain entanglements was governing the fibre diameter.

Effect of solvent-solvent (DMSO-water) interaction on rheological properties and spinning behaviour of PVA solutions was also studied. Because of the strong interaction of water with DMSO, both the rheology and spinnability of PVA solutions were influenced by the composition of the binary solvent mixture. At the same concentration of PVA, the morphology of electrospun fibres changed from highly uniform thick fibres to very fine fibres with large number of beads or droplets.

Influence of incorporating small quantity (1-20 wt%) of non-solvent (1,4-dioxane) on rheology and spinning behaviour of PVA-water mixture was also studied. The spinning behaviour was dependent on the water-dioxane composition, where the diameter first decreased on addition of 1-3 wt% dioxane to 206 ± 21 nm and then it increased to 439 ± 21 nm on addition of 4 to 20 wt%. The diameter was found to have a better correlation with λ as both the fibre diameter and λ increased by 41 and 40%, respectively, compared to the viscosity, which increased by only 13% with the increase in dioxane content.

Poly(galacturonic acid) (PGuA) is an important natural biopolymer, however it has not been processed into fine films and fibres due to its anionic nature and rigid structure. This study aimed at modifying the solution properties of PGuA in aq. sodium hydroxide to enable their conversion into electrospun nanofibres. Addition of anionic surfactants, helped in individualizing the PGuA chains that led to formation of small spindle shaped fibres of length ranging from 2-10 μm and diameter from 287 to 997 nm. However, continuous fibres could not be formed. Using the understanding developed from the PVA Boger fluids, a small amount (1-3 wt% in solution) of high molecular weight PVA was added to modify the rheology of the PGuA solutions. This resulted in formation of continuous fibres. Correlation of fibre diameters of PGuA/PVA with the rheological properties suggested a strong dependence of diameter with the elasticity of the blend solutions.

Applicability and suitability of electrospun PGuA/PVA nanofibres as biocompatible substrates for tissue engineering was also investigated. PGuA based nanofibrous substrates obtained from 70:30 blends of PGuA and PVA were compared with conventionally used PGuA based films for bone tissue engineering. The crosslinked samples of PGuA/PVA nanofibres showed enhanced differentiation and mineralization of hMSCs (human mesenchymal stem cells) as compared to the films. Results indicated that PGuA/PVA based nanofibres facilitated better cell migration and boosted cell adhesion.

The present study was able to develop an understanding of the importance of elasticity, which is expected to facilitate better control of the fibre diameter in electrospinning process. Also, utilising the understanding obtained from this study, spinning of non-spinnable polymers can be achieved.

TABLE OF CONTENTS

CONTENTS	Page
	No.
Certificate	i
Acknowledgements	iii
Abstract	v
Table of Contents	ix
List of Figures	xv
List of Tables	xxv
List of Schemes	xxvii
List of Abbreviations	xxix
Chapter 1 Introduction & literature review	
1.1 Introduction	1
1.2 Electrospinning background, process, setups and various designs	2
1.3 Parameters affecting electrospinning process and nanofibre morphology	6
1.3.1 Effect of voltage	6
1.3.2 Effect of spinning distance	7
1.3.3 Effect of flow rate	8
1.3.4 Surface tension	10
1.3.5 Ambient parameters	10
1.3.6 Effect of solution properties	11
1.3.7 Effect of nature of solvent	15

1.4	Characterization of polymer solutions	18
1.5	Electrospinning of biocompatible synthetic polymers	19
1.6	Electrospinning of natural polymers	22
1.6.1	Use of soluble polymer derivatives	23
1.6.2	Choice of solvents	24
1.6.3	Use of synthetic carrier polymers:	24
1.6.4	Crosslinking methods of natural polymers	26
1.7	Application of electrospun biopolymer nanofibres in bone tissue engineering	27
1.8	Motivation and challenges	30
1.9	Objectives	32
1.10	Organization of thesis	34
Chapter 2 Effect of solution rheology on fibre morphology using PVA based Boger fluids		
2.1	Introduction	37
2.2	Experimental details	38
2.2.1	Materials	38
2.2.2	Preparation of PVA Boger solutions	38
2.2.3	Rheological characterization of PVA solutions	40
2.2.4	Electrospinning of PVA solutions	40
2.2.5	Characterization of PVA nanofibres	41
2.2.6	Statistical analysis	42
2.3	Results and discussion	42

2.3.1	Preparation and characterization of PVA Boger solutions	42
2.3.2	Electrospinning of PVA Boger solutions	51
2.3.3	Correlation of rheological properties and fibre diameter	54
2.3.4	Effect of flow rate, distance and voltage on fibre diameter	59
Chapter 3 Effect of solvents and their mixtures on solution properties and electrospinning behaviour of PVA		
3.1	Introduction	75
3A.1	Effect of two different solvents	76
3A.2	Experimental details	77
3A.2.1	Materials	77
3A.2.2	Preparation of PVA solutions	77
3A.2.3	Rheological characterization of PVA solutions	77
3A.2.4	Electrospinning of PVA solutions	77
3A.2.5	Characterization of PVA nanofibre morphology	78
3A.2.6	Mechanical testing	78
3A.3	Results and discussion	78
3A.3.1	Solution properties	78
3A.3.2	Electrospinning of PVA-DMSO and PVA-water solutions	90

3A.3.3	Correlation of solution properties of PVA-DMSO and PVA-water with fibre diameter	93
3A.3.4	Correlation of fibre diameter of PVA-DMSO and PVA-water with solution entanglement number $(n_e)_{\text{soln}}$	97
3A.3.5	Effect of spinning distance and flow rate on fibre diameter of PVA-DMSO and PVA-water	99
3A.3.6	Mechanical properties of nanofibrous webs of PVA-DMSO and PVA-water solutions	105
3B.1	PVA in DMSO-water mixtures	106
3B.2	Experimental details	106
3B.2.1	Materials	106
3B.2.2	Preparation of solutions	106
3B.2.3	Electrospinning and characterization	107
3B.3	Results and discussion	107
3B.3.1	Solution properties	107
3B.3.2	Electrospinning of PVA-DMSO-water solutions	115
3C.1	Rheology and electrospinning behaviour of PVA in water-dioxane mixtures	123
3C.2	Experimental details	123
3C.2.1	Materials	123
3C.2.2	Preparation of solutions	123

3C.2.3	Electrospinning and characterization methods	125
3C.3	Results and discussion	125
3C.3.1	Visual appearance of solutions	125
3C.3.2	Rheology of PVA-water-dioxane solutions	125
3C.3.3	Electrospinning of PVA-water-dioxane solutions	130
3C.3.4	Correlation of fibre diameter with solution rheology	133
Chapter 4 Rheology and electrospinning of poly(galacturonic acid)		
(PGuA) using solutions modified by high molecular weight		
PVA		
4.1	Introduction	135
4.2	Experimental details	136
4.2.1	Materials	136
4.2.2	Preparation of PGuA and PGuA/PVA solutions	136
4.2.3	Rheological characterization of PGuA solutions	137
4.2.4	Electrospinning of PGuA solutions	139
4.2.5	Raman spectroscopy of PGuA solutions	139
4.2.6	Characterization of electrospun PGuA solutions and PGuA/PVA nanofibres	139
4.3	Results and discussion	139
4.3.1	Dissolution and rheology of PGuA solutions	139
4.3.2	Electrospinning of PGuA/SDS/NaOH solutions	146
4.3.3	Rheology and electrospinning of PGuA/PVA solutions	152

4.3.4	Correlation of fibre diameter with the solution properties	161
Chapter 5 Application of nanofibrous scaffolds based on PGuA for bone tissue engineering		
5.1	Introduction	165
5.2	Experimental details	166
5.2.1	Materials	166
5.2.2	Preparation of scaffolds	166
5.2.3	Physical characterization of nanofibrous web and film scaffolds	167
5.2.4	Pre-conditioning of scaffolds and cell culture	168
5.2.5	Assessment of in vitro biocompatibility, cell attachment, proliferation and differentiation	169
5.2.6	Statistical analysis	172
5.3	Results and discussion	172
5.3.1	Characterization of scaffolds	172
5.3.2	Characterization of cell-scaffold interaction	177
5.3.3	Viability, proliferation and mineralization assay	188
Chapter 6 Conclusions		199
References		205
Appendix		239
Resume of the author		267

LIST OF FIGURES

Figure No.	Caption of the Figure	Page No.
Figure 1.1	Diagrammatic representation of different concentration regions and its effect on fibre morphology	12
Figure 2.1	Schematic representation of the formation of Boger solutions	42
Figure 2.2	Viscosity vs. shear rate of PVA Boger solutions (a) 0 HMW, (b) 0.3 HMW, (c) 0.7 HMW, (d) 1.0 HMW and (e) 1.4 HMW	46
Figure 2.3	Frequency sweep curve of (A) Loss Modulus (B) Storage Modulus for PVA Boger solutions (a) 0 HMW, (b) 0.3 HMW, (c) 0.7 HMW, (d) 1.0 HMW and (e) 1.4 HMW	47
Figure 2.4	Frequency sweep curve for 10 wt% PVA solutions with salt NaCl (a) Control 10 wt% PVA, (b) 0.001 mMol NaCl-10 wt% PVA, (c) 0.01 mMol NaCl-10 wt% PVA, (d) 0.1 mMol NaCl-10 wt% PVA and (e) 1.0 mMol NaCl-10 wt% PVA	48
Figure 2.5	Plot of relaxation time vs. HMW-PVA content for (a) 0 HMW, (b) 0.3 HMW, (c) 0.7 HMW, (d) 1.0 HMW and (e) 1.4 HMW	50
Figure 2.6	SEM images of 2k-PVA at (a) 26 wt%, (b) 28 wt%, (c) 30 wt%, (d) 32 wt% and (e) 34 wt% at distance 15 cm, flow rate 0.5 ml/h	52
Figure 2.7	SEM images of electrospun PVA nanofibres (a) 0 HMW, (b) 0.3 HMW, (c) 0.7 HMW, (d) 1.0 HMW and (e) 1.4 HMW spun at flow rate 0.8 ml/h, distance 15 cm and voltages (MEV) 32, 25, 23, 19 and 17kV respectively	53
Figure 2.8	Fibre diameter vs. viscosity of PVA solutions (a) 0 HMW, (b) 0.3 HMW, (c) 0.7 HMW, (d) 1.0 HMW and (e) 1.4 HMW spun at MEV at flow rate of 0.5 ml/h and distance of 5 cm	55

Figure 2.9	Fibre diameter vs. relaxation time and Deborah number (De) of PVA (a) 0 HMW, (b) 0.3 HMW, (c) 0.7 HMW, (d) 1.0 HMW and (e) 1.4 HMW (i) 30.0%P (ii) 34.0%P. Fibres were spun at MEV at flow rate of 0.5 ml/h and distance of 5 cm	56
Figure 2.10	Extrapolated curve of G' and G'' showing crossover at an angular frequency of 10^4 s^{-1}	57
Figure 2.11	Plot of fibre diameter vs. overall concentration of HMW and LMW PVA content for (a) 0 HMW, (b) 0.3 HMW, (c) 0.7 HMW, (d) 1.0 HMW and (e) 1.4 HMW spun at MEV at flow rate of 0.5 ml/h and distance of 5 cm	58
Figure 2.12(a)	SEM images of 0 HMW solution at different spinning distances (H = 5, 10, 15, 20 and 25 cm) at flow rate of 0.1 ml/h	60
Figure 2.12(b)	SEM images of 1.0 HMW solution at different spinning distances (H = 5, 10, 15, 20 and 25 cm) at flow rate of 0.5 ml/h	61
Figure 2.12(c)	SEM images of 1.4 HMW solution at different spinning distances (H = 5, 10, 15, 20 and 25 cm) at flow rate of 0.1 ml/h	62
Figure 2.13(a)	Effect of distance on fibre diameter at flow rate of 0.1 ml/h for the solutions 0 HMW, 0.3 HMW, 0.7 HMW, 1.0 HMW and 1.4 HMW	63
Figure 2.13(b)	Effect of distance on fibre diameter at flow rate of 0.3 ml/h for the solutions 0 HMW, 0.3 HMW, 0.7 HMW, 1.0 HMW and 1.4 HMW	63
Figure 2.13(c)	Effect of distance on fibre diameter at flow rate of 0.5 ml/h for the solutions 0 HMW, 0.3 HMW, 0.7 HMW, 1.0 HMW and 1.4 HMW	64
Figure 2.13(d)	Effect of distance on fibre diameter at flow rate of 0.8 ml/h for the solutions 0 HMW, 0.3 HMW, 0.7 HMW, 1.0 HMW and 1.4 HMW	64
Figure 2.13(e)	Effect of distance on fibre diameter at flow rate of 1.0 ml/h for the solutions 0 HMW, 0.3 HMW, 0.7 HMW, 1.0 HMW and 1.4 HMW	65

Figure 2.14(a)	SEM images of 0 HMW solution at different flow rates (FR = 0.1, 0.3, 0.5, 0.8 and 1.0 ml/h) at spinning distance of 15 cm	67
Figure 2.14(b)	SEM images of 1.0 HMW solution at different flow rates (FR = 0.1, 0.3, 0.5, 0.8 and 1.0 ml/h) at spinning distance of 15 cm	68
Figure 2.14(c)	SEM images of 1.4 HMW solution at different flow rates (FR = 0.1, 0.3, 0.5, 0.8 and 1.0 ml/h) at spinning distance of 15 cm	69
Figure 2.15(a)	Effect of flow rate on fibre diameter at distance of 5 cm for the solutions 0 HMW, 0.3 HMW, 0.7 HMW, 1.0 HMW and 1.4 HMW	70
Figure 2.15(b)	Effect of flow rate on fibre diameter at distance of 10 cm for the solutions 0 HMW, 0.3 HMW, 0.7 HMW, 1.0 HMW and 1.4 HMW	70
Figure 2.15(c)	Effect of flow rate on fibre diameter at distance of 15 cm for the solution 0 HMW, 0.3 HMW, 0.7 HMW, 1.0 HMW and 1.4 HMW	71
Figure 2.15(d)	Effect of flow rate on fibre diameter at distance of 20 cm for the solutions 0 HMW, 0.3 HMW, 0.7 HMW, 1.0 HMW and 1.4 HMW	71
Figure 2.15(e)	Effect of flow rate on fibre diameter at distance of 25 cm for the solutions 0 HMW, 0.3 HMW, 0.7 HMW, 1.0 HMW and 1.4 HMW	72
Figure 2.16	Dependence of fibre diameter on minimum electrospinning voltage at flow rate 0.8 ml/h for the solutions (a) 0 HMW, (b) 0.3 HMW, (c) 0.7 HMW, (d) 1.0 HMW and (e) 1.4 HMW at a distance of 5 and 15 cm	73
Figure 3.1	Viscosity vs. shear rate curves for PVA solutions at 35 °C in (a) DMSO and (b) water	81
Figure 3.2	Plot of specific viscosity vs. concentration for PVA solutions in (a) DMSO and (b) water	83
Figure 3.3	Graph of (a) loss and (b) storage modulus for PVA solutions in DMSO	85

Figure 3.4	Graph of (a) loss and (b) storage modulus for PVA solutions in water	86
Figure 3.5	Modified Cole-Cole plots for PVA solution in (a) DMSO and (b) water	87
Figure 3.6	Plot of $\tan(\delta)$ vs. angular frequency for PVA solutions in (a) DMSO and (b) water	89
Figure 3.7	SEM images of nanofibres obtained from PVA-DMSO solutions of concentrations (a) 10, (b) 12, (c) 14, (d) 16, (e) 18, and (f) 20 wt%	91
Figure 3.8	SEM images of nanofibres obtained from PVA-water solutions of concentrations (a) 6, (b) 8, (c) 10, (d) 12, (e) 14, and (f) 16 wt%	92
Figure 3.9	Plots of fibre diameter vs. viscosity for PVA-DMSO and PVA-water systems	94
Figure 3.10	Plot of diameter vs. relaxation time of PVA-water and PVA-DMSO systems	96
Figure 3.11	Effect of entanglement per chain on nanofibre diameter for PVA-water (blue diamonds) and PVA-DMSO (red squares) systems	98
Figure 3.12	Plots of diameter as a function of (a) spinning distance at constant flow rate of 0.5 ml/h and (b) flow rates at a constant height of 25 cm 18PVA-D and 12PVA-W solutions	100
Figure 3.13i	SEM images of nanofibres obtained from 18PVA-D for spinning distances of (a) 5, (b) 10, (c) 15, (d) 20 and (e) 25 cm at a flow rate of 0.5 ml/h	101
Figure 3.13ii	SEM images of nanofibres obtained from 12PVA-W for spinning distances of (a) 5, (b) 10, (c) 15, (d) 20 and (e) 25 cm at a flow rate of 0.5 ml/h	102
Figure 3.14i	SEM images of nanofibres obtained from 18PVA-D for flow rates of (a) 0.1,(b) 0.3,(c) 0.5, (d) 0.8 and (e) 1.0 ml/h for spinning distance of 25 cm	103

Figure 3.14ii	SEM images of nanofibres obtained from 12PVA-W for flow rates of (a) 0.1,(b) 0.3,(c) 0.5, (d) 0.8 and (e) 1 ml/h for spinning distance of 25 cm	104
Figure 3.15	Tensile strength and breaking strain of nanofibrous webs from 14PVA-W and 22PVA-D having similar fibre diameter and deposition	105
Figure 3.16	Photograph of 10 wt% PVA-DMSO-water solutions showing optical transparency	109
Figure 3.17	Viscosity vs. shear rate curve of 10 wt% PVA in different DMSO-water ratios	110
Figure 3.18	(a) Loss and (b) Storage modulus of 10 wt% PVA in various DMSO-water mixtures	111
Figure 3.19	Modified Cole-Cole plot for 10 wt% PVA-DMSO-water solutions	112
Figure 3.20	Plot of loss tangent vs. angular frequency for 10 wt% PVA-DMSO-water solutions	113
Figure 3.21	Viscosity and relaxation time curve of PVA-DMSO-water solutions	114
Figure 3.22	Photographs of electrospinning jet lengths (straight path of jet after Taylor cone) of 10 wt% PVA solutions in various DMSO-water ratios. Solution with 40 wt% DMSO shows maximum jet length	116
Figure 3.23i	SEM images of electrospun nanofibres obtained from 10 wt% PVA at different DMSO-water ratios, (a) 10PVA-W (b) 10PVA-20D (c) 10PVA-30D, (d) 10PVA-40D and (e) 10PVA-50D	117
Figure 3.23ii	SEM images of electrospun nanofibres obtained from 10 wt% PVA at different DMSO-water ratios, (f) 10PVA-60D (g) 10PVA-70D, (h) 10PVA-80D and (i) 10PVA-100D	118

Figure 3.24	Graph of diameter vs. DMSO mole fraction with schematic representation of polymer-solvent interaction at various mole fraction of DMSO. Region I of the graph shows uniform fibres due to free water molecules. Region II and III show the liquid-liquid demixing that leads to non-uniform fibres and beaded fibres. Region IV shows fibres with droplets due to free DMSO molecules	120
Figure 3.25	Photographs showing appearance of the PVA-water-dioxane solutions prepared at 30 °C. (a) 10PVA-W, (b) 10 PVA-2Dix, (c) 10PVA-4Dix, (d) 10PVA-6Dix, (e) 10PVA-10Dix, (f) 10PVA-20Dix, (g) 10PVA-30Dix, (h) 10PVA-50Dix and (i) 10PVA-60Dix	126
Figure 3.26	Viscosity vs. shear rate curve for PVA-water-dioxane solutions	127
Figure 3.27	(a) Loss and (b) storage modulus curves for PVA-water-dioxane solutions	128
Figure 3.28	Modified Cole–Cole plot for PVA-water-dioxane solutions	129
Figure 3.29	Phase behaviour of PVA-water-dioxane ternary system	130
Figure 3.30i	SEM images of nanofibres obtained from PVA-water-dioxane solutions	131
Figure 3.30ii	SEM images of nanofibres obtained from PVA-water-dioxane solutions	132
Figure 3.31	Plots of viscosity, relaxation time of PVA-water-dioxane solutions and diameter of the resultant fibres as a function of mole fraction of dioxane in the solutions.	134
Figure 4.1	Solution of 8 wt% PGuA in aq. NaOH (a) without SDS and (b) with SDS	140
Figure 4.2	Raman spectra of PGuA in different environments	141
Figure 4.3	Effect of SDS concentration on Raman spectra of PGuA in aq. NaOH solution	141
Figure 4.4	Effect of concentration of SDS on viscosity of 8 wt% PGuA/SDS/NaOH solutions	142
Figure 4.5	Influence of NaOH concentration on the gelling tendency and dissolution of PGuA	145

Figure 4.6(a)	SEM image of electrospun 8 wt% PGuA solution in aq. NaOH without SDS	146
Figure 4.6(b)	SEM images of electrospun 8 wt% PGuA/SDS/NaOH solutions with SDS concentrations of (a) 2 wt% (8PGuA-2SDS), (b) 5 wt% (8PGuA-5SDS), (c) 10 wt% (8PGuA-10SDS), (d) 15 wt% (8PGuA-15SDS), (e) 30 wt% (8PGuA-30SDS) and (f) 50 wt% (8PGuA-50SDS)	147
Figure 4.7	Viscosity vs. shear rate curve for PGuA/SDS/NaOH solutions with different concentrations	148
Figure 4.8	Specific viscosity vs. concentration curve for PGuA/SDS/NaOH solutions	150
Figure 4.9	SEM images of electrospun nanostructures obtained from PGuA/SDS/NaOH solutions of different concentrations (a) 6PGuA-10SDS, (b) 7PGuA-10SDS, (c) 9PGuA-10SDS and (d) 10PGuA-10SDS	151
Figure 4.10	Viscosity vs. shear rate curves for PGuA/PVA solutions	153
Figure 4.11	(a) Loss and (b) storage modulus vs. angular frequency curves of PGuA/PVA solutions	155
Figure 4.12	Tan(δ) plot of PGuA/PVA solutions	156
Figure 4.13	Modified Cole-Cole plot for PGuA/PVA solutions	158
Figure 4.14i	SEM images of PGuA/PVA blend solutions with various ratios of PGuA and PVA	159
Figure 4.14ii	SEM images of PGuA/PVA blend solutions with various ratios of PGuA and PVA	160
Figure 4.15	Plots of (a) viscosity, (b) relaxation time of PGuA/PVA solutions and (c) their resulting fibre diameter vs. PVA content in solutions	162
Figure 5.1	SEM images of calcium treated and GA crosslinked as-prepared scaffolds of (a) PGuA/PVA/Ca-NF, (b) PGuA/PVA/Ca-Film, (c) PGuA/Ca-Film and of scaffolds after dipping in DI water for 7 days (d) PGuA/PVA/Ca-NF, (e) PGuA/PVA/Ca-Film, (f) PGuA/Ca-Film	173
Figure 5.2	FTIR spectra of various as-prepared and crosslinked scaffolds	175

Figure 5.3	Stress vs. strain curves for as-spun and crosslinked PGuA/PVA nanofibre scaffolds	177
Figure 5.4(a)	SEM images of PGuA based scaffolds and cover slip showing cellular morphology of hMSCs for day 1 of cell culture	178
Figure 5.4(b)	SEM images of PGuA based scaffolds and cover slip showing cellular morphology of hMSCs for day 3 of cell culture	179
Figure 5.4(c)	SEM images of PGuA based scaffolds and cover slip showing cellular morphology of hMSCs for day 5 of cell culture	180
Figure 5.4(d)	SEM images of PGuA based scaffolds and cover slip showing cellular morphology of hMSCs for day 7 of cell culture	181
Figure 5.4(e)	SEM images of PGuA based scaffolds and cover slip showing cellular morphology of hMSCs for day 14 of cell culture	182
Figure 5.4(f)	SEM images of PGuA based scaffolds and cover slip showing cellular morphology of hMSCs for day 21 of cell culture	183
Figure 5.5	High magnification SEM images of bone nodules expressed on nanofibrous substrate, showing needle like crystal formation, on (a and b) day 3 and (c and d) day 5 of cell culture	184
Figure 5.6	Image showing (a) migration of a cell underneath the nanofibres in PGuA/PVA/Ca-NF (b) localized cell attachment and growth of hMSCs on PGuA/Ca-Film substrate	185
Figure 5.7	EDX of expressed mineral particles by cultured hMSCs over (a) PGuA/PVA/Ca-NF, (b) PGuA/PVA/Ca-Film, (c) PGuA/Ca-Film and (d) Cover slip	186
Figure 5.8	Fluorescent microscopy images for Live-Dead assay of (a) Control cover slip, (b) PGuA/PVA/Ca-NF, (c) PGuA/PVA/Ca-Film and (d) PGuA/Ca-Film. Images are taken at 200× magnification	189

Figure 5.9	MTT assay for assessment of metabolic activity and proliferation of hMSCs cultured over various PGuA based scaffolds and cover slip for day 3, 5, 7, 10 and 14. (**) denotes significant difference of $p < 0.01$ compared to control and film samples	190
Figure 5.10	ALP activity of hMSCs on various PGuA based scaffolds and cover slip for day 5, 7, 14 and 21. (*) denotes significance level of $p < 0.05$	191
Figure 5.11(a)	Optical images of ARS stained mineralized PGuA based scaffolds and cover slips cultured with hMSCs after day 7 of culture	193
Figure 5.11(b)	Optical images of ARS stained mineralized PGuA based scaffolds and cover slips cultured with hMSCs after day 14 of culture	194
Figure 5.11(c)	Optical images of ARS stained mineralized PGuA based scaffolds and cover slips cultured with hMSCs after day 21 of culture	195
Figure 5.12	Biochemical assessment of mineralization of hMSCs cultured over PGuA based substrates and coverslips by Alizarin Red staining. (*) denotes a significance level of $p \leq 0.05$ compared to control sample	196
Figure 5.13	Graph showing concentration of calcium mineral expressed by hMSCs cultured over PGuA based substrates and cover slip	197

LIST OF TABLES

Table No.	Title of the Table	Page No.
Table 1.1	List of common synthetic biocompatible polymers electrospun for various biomedical applications	20
Table 1.2	Electrospun polysaccharides for various biomedical applications	30
Table 2.1	Composition of all the solutions prepared for the formation of PVA Boger solutions	39
Table 2.2	Rheological properties of PVA solutions containing different proportions of HMW and LMW	45
Table 2.3	Rheological properties of 2k-PVA solutions of 30, 32 and 34 wt%	51
Table 3.1	Rheological parameters for PVA-DMSO solutions	79
Table:3.2	Rheological parameters for PVA-water solutions	80
Table 3.3	Rheological properties and spinning behaviour of 10 wt% PVA solutions in different ratios of DMSO-water mixtures	108
Table 3.4	Composition and rheological properties of PVA-water-dioxane solutions	124
Table 4.1	Compositions of PGuA solution with varying amounts of NaOH	137
Table 4.2	Compositions of PGuA solutions in SDS and aq. NaOH	138
Table 4.3	Viscosity of PGuA solutions	143
Table 4.4	Rheological properties of PGuA/PVA solutions	154
Table 5.1	Samples used for cell culture experiments	169
Table 5.2	Values of Ca/P ratio in expressed minerals obtained using EDX analysis of samples culture with hMSCs	187

LIST OF SCHEMES

Scheme No.	Title of the Scheme	Page No.
Scheme 1.1	Schematic representation of electrospinning process and parameters	3
Scheme 3.1	PVA chain interaction in (a) water and (b) DMSO	96
Scheme 3.2	Complex formation in DMSO and water molecules.	119
Scheme 4.1	Effect of SDS on individualization of PGuA chains.	144

LIST OF SYMBOLS AND ABBREVIATIONS

Symbol/Abbreviation	Meaning
ω	angular frequency
η	apparent viscosity
G^*	complex modulus
η^*	complex viscosity
J'	compliance
c	concentration
c^*	critical concentration
α	dilution coefficient
η'	dynamic viscosity
G	gauge
χ	interaction parameter
$[\eta]$	intrinsic viscosity
kV	kilovolt
G''	loss modulus
MPa	mega pascal
M	molar
Pa	pascal
Φ	polymer volume fraction
rad	radian
λ	relaxation time
γ°	shear strain
τ	shear stress
G'	storage modulus
wt%	weight percentage
η_0	zero shear viscosity
HFIP	1,1,1,3,3,3-hexafluoro-2-propanol
TFA	2,2,2-trifluoroethanoic acid

TFE	2,2,2-trifluoroethanol
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
ARS	alizarin red s
ALP	alkaline phosphatase
BSA	bovine serum albumin
CV	collector voltage
De	Deborah number
DH	degree of hydrolysis
DI	de-ionized
dia	diameter
DCM	dichloromethane
DMSO	dimethyl sulfoxide
DMEM	Dulbecco's modified Eagle's medium
EV	effective voltage
C_e	entanglement concentration
$(n_e)_{\text{soln}}$	entanglement number of solution
ECM	extra cellular matrix
FBS	fetal bovine serum
Gel	gelatin
GAGs	glycosaminoglycans
HMW	high molecular weight
hMSCs	human mesenchymal stem cells
HA	hydroxyapatite
LG	low glucose
LMW	low molecular weight
M_e	melt entanglement molecular weight
MSCs	mesenchymal stem cells
MEV	minimum electrospinning voltage
DMAc	<i>N,N</i> -dimethylacetamide
DMF	<i>N,N</i> -dimethylformamide
NV	needle voltage

NBF	neutral buffered formalin
NMP	<i>N</i> -methyl-2-pyrrolidone
NMMO	<i>N</i> -methylmorpholine <i>N</i> -oxide
PBS	phosphate-buffered saline
PVDF	poly(1,1-difluoroethene)
PAN	poly(1-acrylonitrile)
PANI	poly(aniline)
PCL	poly(<i>ε</i> -caprolactone)
PEG	poly(ethylene glycol)
PEO	poly(ethylene oxide)
PGuA	poly(galacturonic acid)
PHB	poly(hydroxyl butyrate)
PLA-PGA	poly(lactic-co-glycolic acid)
PGA	poly(L-glycolic acid)
PLA	poly(L-lactic acid)
PMA	poly(methacrylate)
PVP	poly(<i>N</i> -vinyl pyrrolidone)
PS	poly(styrene)
PVA	poly(vinyl alcohol)
RH	relative humidity
SDS	sodium dodecyl sulphate
$M_{e(soln)}$	solution entanglement molecular weight
THF	tetrahydrofuran
3-D	three dimensional
TE	tissue engineering
2-D	two dimensional
UV	ultraviolet
TCP	β -tricalcium bis(phosphate)